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Instantaneous insemination in the millipede *Centrobolus inscriptus* (Spirobolida: Trigiulidae) determined by artificially-terminated mating

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Abstract

Male millipedes may control the duration of copulation and maintain genital contact with females in relation to the intensity of sperm competition. Post-insemination associations were considered in arthropods and the timing of insemination in the millipede *Centrobolus inscriptus* calculated by artificially-terminated radiolabelled mating. Copula pairs were separated after 1, 2, 3, 4, 5, 10, 20, and 30 minutes of copulation. Ejaculate volumes inseminated did not correlate significantly with the manipulated copulation durations ($r = -0.35$, $df = 8$, $P = 0.35$). High (*ca.* 2500 dpm) and low values in the first 5 minutes evidenced "instantaneous-insemination" and self-sperm displacement. Instantaneous insemination was demonstrated using artificially terminated mating in *Centrobolus inscriptus* by showing no relationship between ejaculate volume and copulation duration except for the high and low volumes at the beginning of mating when the male loads and seats the gonopods before adaptive mate-guarding by prolonged copulation.

Keywords: Arthropoda; *Centrobolus inscriptus*; Diplopoda; postinsemination; mate-guarding

1. Introduction

Mate guarding is known in the Arthropod classes Diplopoda [1-14, 58-59], Araneae [44, 47-48], Acari [45-46, 61, 65], Crustacea [49-50, 56-57, 60, 62], Scorpiones [52], Opiliones [53], and Insecta, orders Orthoptera [31, 51], Odonata [26, 34, 35], Phasmida [36], Lepidoptera [21, 37], Diptera [30, 38], Coleoptera [19, 20, 22, 27, 29, 39, 40, 63-64], Hymenoptera [28, 41], Hemiptera [15-18, 23-25, 42-43]. Post-insemination associations and the mate-guarding hypothesis has been previously reviewed for Insecta [32]. Millipedes (Class: Diplopoda) have highly polygynandrous mating systems that are very complex because males and females can mate multiply and prolong copulations with maintained genital contact where males control the duration of copulation and maintain genital contact with females in relation to the intensity of sperm competition [1-4]. The instantaneous timing of insemination has been detected using radiolabeled ejaculates in *Alloporus uncinatus* (Spirostreptida: Spirostreptidae), followed by the adaptive mate-guarding phase with maintained genital contact [13]. The collection of sperm at the bottom of the spermathecae from the beginning of insemination with time after the start of copulation indicated prolonged copulation was the adaptive mate-guarding phase [13]. The objective of the present study is to determine the timing of insemination in the millipede *Centrobolus inscriptus*, replicating the artificially-terminated radio-labelled mating experiments which were performed in *A. uncinatus* [13], to test if prolonged genital contact is the adaptive mate-guarding phase.

2. Materials and Methods

Millipedes were hand collected (1996-1998) from indigenous coastal forest at Twin Streams Farm, Mtunzini, South Africa (28°55'S, 31 °45'E). Live specimens of each sex were transported to Cape Town and kept at 25°C temperature; 70% relative humidity; 12:12 hours light-dark cycle. Food was provided in the form of fresh vegetables *ad libitum*. Unisexual groups were housed in plastic containers containing moist vermiculite (± 5 cm deep) before the mating experiments commenced. The radioisotope labelling technique allowed two types of ejaculates to be discerned [11]. A Hamilton syringe was used to inject 50 μ L aliquots of tritiated [*methyl*] thymidine (85 Ci / mmol, Amersham, UK) between the tergites of the 10th and 11th diplosegments of individual males (L). A second class of males did not receive the treatment and were left unlabelled males (UL). Females were killed in ethyl-acetate jars after their last copulation and the paired sperm storage organs were dissected out under a magnifying lens

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(5X). Left and right spermathecae were placed in separate 7ml scintillation vials and vortexed for 30s with 0.1 ml concentrated HCl to promote rapid tissue homogenisation [12]. Acid was neutralised with 0.1 ml 5M NaOH prior to adding 3.5 ml scintillation fluid (Scintillator 299, Packford). The volume of labelled ejaculate present in the female spermathecae was quantified in disintegrations per minute (dpm) of radioisotope using a 1600 scintillation counter (low count reject = 0; dpm multiplier = 1). Thus dpm values were used as volumetric indications of the labelled ejaculate present in the female spermathecae. Radiolabelled males were used in artificially-terminated matings to elucidate the timing of insemination [12]. Copula pairs were separated after 1, 2, 3, 4, 5, 10, 20, and 30 minutes of copulation. Statistical analyses were performed using Statgraphics (version 6.0). The non-parametric nature of disintegration counts [11], coupled with small sample sizes, did not allow data to be normally transformed. Pearson's correlation coefficients were used to analyse the relationships between ejaculate volume, copulation time and body mass.

3. Results

Ejaculate volumes (Figure 1) inseminated did not correlate significantly with the manipulated copulation durations ($r = -0.35$, $df = 8$, $P = 0.35$) but the first few data points indicate very high volumes (ca. 2500 dpm) deposited within the first five minutes interspersed with the range of lower volumes. Male body mass was not related to ejaculate volume ($r = 0.54$, $df = 8$, $p = 0.13$) and was not related to copulation duration ($r = -0.64$, $df = 8$, $P = 0.06$).

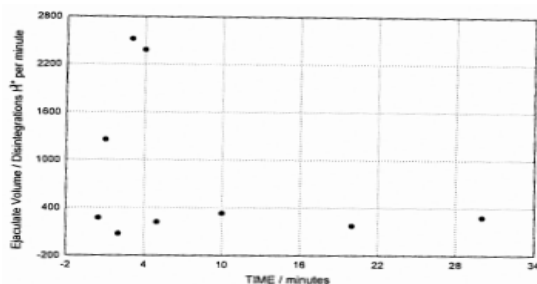


Fig 1: The ejaculate volumes inseminated by radiolabelled males of *Centrobolus inscriptus* when matings were terminated at different stages.

4. Discussion

The lack of correlation between ejaculate volume inseminated and manipulated copulation duration evidenced "instantaneous-insemination" during which time the male loads and seats the gonopods before engaging in the adaptive mate-guarding phase [12]. This was supported by high and low volumes of ejaculate in the first five minutes of copulation supporting self-sperm displacement and matches the predicted results found in the spirostreptid *A. uncinatus* [13]. The absence of an allometric relationship here supports the previous findings in millipedes. This is an interesting result and probably indicates this is the situation in all Julid millipedes where there is a post-insemination association. It was critical to discern between the different stages or phases of copulation in order to show mate-guarding as an adaptation to counter sperm-competition through the integration of pre- or syn-copulatory and post-copulatory events [33].

5. Conclusion

Instantaneous insemination was demonstrated using artificially terminated mating in *Centrobolus inscriptus* by showing no relationship between ejaculate volume and copulation duration except for high and low volumes at the beginning of mating when the male loads and seats the gonopods before adaptive mate-guarding by prolonged copulation.

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7. References

1. Telford SR, Dangerfield JM. Sex in millipedes: laboratory studies on sexual selection. *Journal of Biological Education*. 1990; 24(4):233-238.
2. Telford SR, Dangerfield JM. Manipulation of the sex-ratio and duration of copulation in the tropical millipede *Alloporus uncinatus* - a test of the copulatory guarding hypothesis. *Animal Behaviour* 1990; 40(5):984-986.
3. Telford SR, Dangerfield JM. Mating tactics in the tropical millipede *Alloporus uncinatus* (Diplopoda: Spirostreptidae). *Behaviour* 1993; 124(1):45-56.
4. Telford SR, Dangerfield JM. Males control the duration of copulation in the tropical millipede *Alloporus uncinatus* (Diplopoda: Julida). *South African Journal of Zoology*. 1994; 29(4):266-268.
5. Telford SR, Dangerfield JM. Sexual selection in savanna millipedes: Products, patterns and processes. In: Geoffroy, J. J., Mauries, J.P. & Nguyen Duy-Jacquemin, *Mémoires du Muséum National d'Histoire Naturelle* 1996; 169:565-576.
6. Telford SR, Dangerfield JM. Males control the duration of copulation in the tropical millipede *Alloporus uncinatus* (Diplopoda: Julida). *South African Journal of Zoology* 1994; 29(4):266-268.
7. Barnett M, Telford SR, Villiers de CJ. The genital morphology of the millipede *Orthoporus pyrrocephalus* (Diplopoda: Spirostreptidae) - a possible mechanism of sperm displacement. *Proceedings of the Electron Microscopy Society of South Africa* 1991; 21:15-17.
8. Cooper MI, Telford SR. Copulatory sequences and sexual struggles in millipedes. *Journal of Insect Behaviour*. 13(2):217-230.
9. Barnett M, Telford SR, Villiers de CJ. Sperm competition in a millipede? An investigation into the genital morphology of the southern African spirostreptid millipede *Orthoporus pyrrocephalus*. *Journal of Zoology London*. 1993; 231(3):511-522.
10. Barnett M, Telford SR. Sperm competition and the evolution of millipede genitalia. In JC Geoffroy, JP Mauries, M Nguyen Duy-Jacquemin. *Mémoires du Muséum National d'Histoire Naturelle* 1996; 169:331-339.
11. Barnett M, Telford SR, Tibbles BJ. Female mediation of sperm competition in the millipede *Alloporus uncinatus* (Diplopoda: Spirostreptidae). *Behavioural Ecology and Sociobiology* 1995; 36(6):413-419.
12. Barnett M, Telford SR. The timing of insemination and its implications for sperm competition in a millipede with prolonged copulation. *Animal Behaviour* 1994; 48(2):482-484.

13. Rowe M. Copulation, mating system and sexual dimorphism in an Australian millipede, *Cladethosoma clarum*. *Australian Journal of Zoology*. 2010; 58(2):127-132.
14. Galipaud M, Bollache L, Dechaume-Moncharmont F-X. Assortative mating by size without a size-based preference: the female-sooner norm as a mate-guarding criterion. *Animal Behaviour* 2013; 85(1):35-41.
15. Sillén-Tullberg B. Prolonged copulation: A male 'postcopulatory' strategy in a promiscuous species, *Lygaeus equestris* (L.) (Heteroptera: Lygaeidae). *Behavioural Ecology and Sociobiology* 1981; 9(4):283-289.
16. Clark SJ. The effects of operational sex-ratio and food-deprivation on copulation duration in the water strider (*Gerris-remigis* Say). *Behavioural Ecology and Sociobiology* 1988; 23(5):317-322.
17. Campbell V, Fairbairn DJ. Prolonged copulation and the internal dynamics of sperm transfer in the water strider *Aquarius remigis*. *Canadian Journal of Zoology* 2001; 79(10):1801-1812.
18. McLain DK. Prolonged copulation as a post-insemination guarding tactic in a natural population of the ragwort seed bug. *Animal Behaviour* 1989; 38(4):659-664.
19. Harano K, Tanaka S, Yasui H, Wakamura S, Nagayama A, Hokama Y *et al.* Multiple mating, prolonged copulation and male substance in a scarab beetle *Dasylepida ishigakiensis* (Coleoptera: Scarabaeidae). *International Journal of Tropical Insect Science* 2010; 30(3):119-126.
20. Wing SR. Prolonged copulation in *Photinus macdermotti* with comparative notes on *Photinus collustrans* (Coleoptera: Lampyridae). *The Florida Entomologist* 1985; 68(4):627-634.
21. Watanabe M, Nakanishi Y, Bonno M. Prolonged copulation and spermatophore size ejaculated in the sulfur butterfly, *Colias erate* (Lepidoptera: Pieridae) under selective harassments of mated pairs by conspecific lone males. *Journal of Ethology*. 1997; 15(1):45-54.
22. Harari AR, Landolt PJ, O'Brien CW, Brockmann HJ. Prolonged mate guarding and sperm competition in the weevil *Diaprepes abbreviates* (L.). *Behavioural Ecology*, 14(1):89-96.
23. Schöfl G, Taborisky M. Prolonged tandem formation in firebugs (*Pyrrhocoris apterus*) serves mate-guarding. *Behavioural Ecology and Sociobiology* 2002; 52(5):426-433.
24. Carroll SP. The adaptive significance of mate guarding in the soapberry bug, *Jadera haematoloma* (Hemiptera: Rhopalidae). *Journal of Insect Behaviour* 1991; *Journal of Insect Behaviour* 1991; 4(4):509-530.
25. Vitta ACR, Lorenzo MG. Copulation and mate guarding behavior in *Triatoma brasiliensis* (Hemiptera: Reduviidae). *Journal of Medical Entomology*. 2009; 46(4):789-795.
26. Andres JA, Rivera AC. Copulation duration and fertilization success in a damselfly: an example of cryptic female choice? *Animal Behaviour* 2000; 59(4):696-703.
27. Saeki Y, Kruse KC, Switzer PV. Physiological costs of mate guarding in the Japanese beetle (*Popillia japonica* Newman). *Ethology* 2005; 111(9):863-867.
28. Brown MJF, Baer B. The evolutionary significance of long copulation duration in bumble bees. *Apidologie*, 36(2):157-167.
29. Chaudhary DD, Mishra G, Omkar O. Prolonged matings in a ladybird, *Menochilus sexmaculatus*: A mate guarding mechanism? *Journal of Asia-Pacific Entomology*, 18(3): 453-458.
30. Martin OY, Hosken DJ. Strategic ejaculation in the common dung fly *Sepsis cynipsea*. *Animal Behaviour* 2002; 63(3):541-546.
31. Parker GA, Smith JL. Sperm competition and the evolution of the precopulatory passive phase in *Locusta migratoria migratorioides*. *Physiological Entomology* 1975; 49(2):155-171.
32. Alcock J. Postinsemination associations between males and females in Insects: The mate-guarding hypothesis. *Annual Review of Entomology* 1994; 39(1):1-21.
33. Parker GA, Pizzari T. Sperm competition and ejaculate economics. *Biological Reviews* 2010; 85(4):897-834.
34. Cordero A. The adaptive significance of prolonged copulations of the damselfly, *Ischnura graellsii* (Odonata: Coenagrionidae). *Animal Behaviour* 1990; 40(1):43-48.
35. Michiels NK. Consequences and adaptive significance of variation in copulation duration in the dragonfly *Sympetrum danae*. *Behavioural Ecology and Sociobiology* 1992; 29(6):429-435.
36. Sivinski J. Predation and sperm competition in the evolution of coupling durations, particularly in the stick insect *Diapheromera veliei*. In *Competition in a diverse group of Insects*, Gwynne DT, Morris GK, Westview, USA, 1983, 147-162.
37. Drummond BA III. Multiple mating and sperm competition in the Lepidoptera. In Smith RL, *Sperm competition and the evolution of Insect mating systems*, Academic Press, New York, 1984, 291-370.
38. Thornhill R. Sexual selection within mating swarms of the lovebug *Plecia nearctica* (Diptera: Bibionidae). *Animal Behaviour* 1980; 28(2): 405-412.
39. Peschke K. Male aggression, female mimicry and female choice in the rove beetle, *Aleochara curtula*. *Ethology* 1987; 75(4):265-284.
40. Snead S, Alcock J. Aggregation formation and assortative mating in two meloid beetles. *Evolution* 1985; 39(5):1123-1131.
41. Rutowski R, Alcock J. Temporal variation in male copulatory behaviour in the solitary bee *Nomadopsis puellae* (Hymenoptera: Andrenidae). *Behaviour* 1980; 73(3/4):175-188.
42. Carroll SP, Loe JE. Male-biased sex ratios, female promiscuity, and copulatory mate guarding in an aggregating tropical bug, *Dysdercus bimaculatus*. *Journal of Insect Behaviour*. 1990; 3:33-48.
43. McLain DK. Female choice and adaptive significance of prolonged copulation in *Nezara viridula* (Hemiptera: Pentatomidae). *Psyche* 1980; 87(3-4):325-336.
44. Sziranyi A, Kiss B, Samu F. The function of long copulation in the wolf spider *Pardosa agrestis* (Araneae, Lycosidae) investigated in a controlled copulation duration experiment. *The Journal of Arachnology*. 2005; 33:408-414.

45. Radwan J. Sperm competition in the mite *Caloglyphus berlesei*. *Behavioural Ecology and Sociobiology* 1991; 29(4):291-296.
46. Radwan J, Siva-Jothy MT. The function of post-insemination mate association in the bulb mite, *Rhizoglyphus robini*. *Animal Behaviour* 1996; 52(4):651-657.
47. Elgar MA, Bathgate R. Female receptivity and male mate-guarding in the jewel spider *Gasteracantha minax* Thorell (Araneidae). *Journal of Insect Behaviour*. 1996; 9(5):729-738.
48. Calbacho-Rosa L, Peretti AV. Copulatory and post-copulatory sexual selection in Haplogyne spiders, with emphasis on Pholcidae and Oonopidae. In Peretti AV, Aisenberg A, Cryptic female choice in Arthropods, Springer, Switzerland, 2015, 109-144.
49. Elgar MA. Sperm competition and sexual selection in spiders and other Arachnids. In Birkhead TR, Møller, AP, Sperm competition and sexual selection. Academic Press, New York, 1998, 307-339.
50. Jivoff P. The relative roles of predation and sperm competition on the duration of the post-copulatory association between the sexes in the blue crab, *Callinectes sapidus*. *Behavioural Ecology and Sociobiology* 1997; 40(3):175-185.
51. Jivoff P, Hines AH. Female behaviour, sexual competition and mate guarding in the blue crab, *Callinectes sapidus*. *Animal Behaviour* 1998; 55(3):589-603.
52. Tunj C, Beveridge M, Simmons LW. Female crickets assess relatedness during mate guarding and bias storage of sperm towards unrelated males. *Journal of Evolutionary Biology*. 2013; 26(6):1261-1268.
53. Benton TG. Determinants of male-mating success in a scorpion. *Animal Behaviour* 1992; 43(1):125-135.
54. Buzatto BA, Machado G. Male dimorphism and alternative reproductive tactics in harvestmen (Arachnida: Opiliones). *Behavioural Processes* 2014; 109(A):2-13.
55. Machado G, Requena GS, Toscano-Gadea C, Stanley E, Macías-Ordóñez R. Male and female mate choice in Harvestmen: General patterns and inferences on the underlying processes. In Peretti AV, Aisenberg A, Cryptic female choice in Arthropods, Springer, Switzerland, 2015, 169-201.
56. Jormalainen V, Merilaita S. Female resistance and duration of mate-guarding in three aquatic peracarids (Crustacea). *Behavioral Ecology and Sociobiology* 1995; 36(1):43-48.
57. Benvenuto C, Knott B, Weeks SC. Mate-guarding behavior in clam shrimp: a field approach. *Behavioural Ecology* 2009; 20(6):1125-1132.
58. Adolph SC, Geber MA. Mate-guarding, mating success and body size in the tropical millipede *Nyssodesmus python* (Peters) (Polydesmida, Platyrrhacidae). *Southwestern Naturalist* 1995; 40(1):56-61.
59. Heisler IL. *Nyssodesmus python*. In Costa Rican Natural History, Janzen DH, University of Chicago Press, Chicago, 1983, xi + 816.
60. Glatzel T, Schminke HK. Mating behaviour of the groundwater copepod *Parastenocaris phyllura* Kiefer, (Copepoda: Harpacticoida). *Contributions to Zoology* 1938; 66(2):103-108.
61. Fashing NJ. Mate-guarding in the genus *Creutzeria* (Astigmata: Histiostomatidae), an aquatic mite genus inhabiting the fluid-filled pitchers of *Nepenthes* plants (Nepenthaceae). *Systematic & Applied Acarology* 2008; 13(3):163-171.
62. Goshima S, Kogab T, Muraib M. Mate acceptance and guarding by male fiddler crabs *Uca tetragonon* (Herbst). *Journal of Experimental Marine Biology and Ecology*. 1996; 196(1-2):131-143.
63. Sato H, Hiramatsu K. Mating behaviour and sexual selection in the African ball-rolling scarab *Kheper platynotus* (Bates) (Coleoptera: Scarabaeidae). *Journal of Natural History*. 1993; 27(3):657-668.
64. Knox TT, Scott MP. Size, operational sex ratio, and mate-guarding success of the carrion beetle, *Necrophila Americana*. *Behavioural Ecology* 2006; 17(1):88-96.
65. Oku K. Sexual selection and mating behavior in spider mites of the genus *Tetranychus* (Acari: Tetranychidae). *Applied Entomology and Zoology* 2014; 49(1):1-9.