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## The Southern House Mosquito, *Culex quinquefasciatus*: profile of a smart vector

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### Abstract

*Culex quinquefasciatus* is the principal vector of bancroftian filariasis and a potential vector of *Dirofilaria immitis*. This mosquito species is also a potential vector of several arboviruses like West Nile virus (WNV), Rift Valley fever virus, avian pox and protozoa like *Plasmodium relictum* that causes bird malaria. This species has the ability to transmit other nematodes like *Saurofilaria sp.*, *Oswaldofilaria sp.* In the USA, it is a potential vector of St. Louis encephalitis virus (SLEV). Japanese encephalitis virus (JEV) has been isolated from this mosquito in several occasions in Asia. Furthermore, it can transmit several other arboviruses in the laboratory conditions. This article is an attempt to review the bio-ecology, medical and veterinary importance of *Culex quinquefasciatus*. It acts as an important “urban bridge vector” which bridges different reservoir/amplifier hosts to humans because of its encounter with different vertebrates. *Culex quinquefasciatus* also creates an ecological bridge between urban, periurban and rural areas owing to its presence and adaptability in diverse ecological niches. *Culex quinquefasciatus* emerged as a smart vector because of the adaptive fitness, ecological plasticity, invasive behaviour, host specificity and high reproductive potential along with expanded immune gene repertoire property at the genetic level. This mosquito possesses the necessary potential to initiate and facilitate the disease transmission by establishing an effective vector-host transmission cycle for diverse pathogens in different environments. Thus, in the changing ecological conditions this mosquito might enhance its epidemiological importance in the near future as a smart vector for those pathogens which were isolated from this mosquito species but are presently not having any public health importance.

**Keywords:** *Culex quinquefasciatus*, Pathogen, Bio-ecology, Smart vector, Medical and veterinary importance

### 1. Introduction

The cosmopolitan distribution of *Culex quinquefasciatus* is across continents and ecozones mostly south of 39° N latitude [1]. This mosquito species has been subsequently introduced to New Zealand, Australia and other geographical regions by the anthropogenic activity. It is one of the most widespread mosquito species [2] found in the Eastern Asian region [3], Indomalayan, Australasia [4], Neotropics, Afrotropics [5] and pan and subtropical America [6]. The transmission of arbovirus is facilitated by biological vectors [7]. The principal source of arbovirus transmission is mosquito vectors [8]. The Southern House Mosquito, *Culex quinquefasciatus* is the principal vector of bancroftian filariasis and a potential vector of *Dirofilaria immitis* [9, 10]. This species is considered as a potential vector of West Nile virus (WNV) [11]. Additionally, it can transmit Alfuy, Almpiwar, Corriparta, Sindbis, Ross River virus, Japanese Encephalitis virus (JEV) [12], St. Louis Encephalitis virus (SLEV) [13], Reticuloendotheliosis virus [14], Murray Valley encephalitis [6], Edge Hill, Eubenangee, Getah, Kokobera, Koongol, Kowanyama, Kunjin, Mapputta, Stratford, Trubanaman, Wongal, Reovirus type 3 and Chikungunya virus [4, 14] in laboratory condition. Thus, evidently this cosmopolitan mosquito is a potential vector of many important pathogens causing concern to public health authorities (Table.1). This article is an attempt to review the bio-ecology, medical and veterinary importance of Southern House Mosquito, *Culex quinquefasciatus*.

### 2. Taxonomic Status of *Culex quinquefasciatus*

*Culex quinquefasciatus* is a member of globally distributed *Culex pipiens* species complex. Additionally, the *Culex pipiens* species complex has many related species, ecotypes and hybrids which are situated in geographical introgression zones on multiple continents [15]. *Culex quinquefasciatus* was first described in 1823 by Thomas Say from a specimen collected

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along the Mississippi River in the southern United States. At that time, a number of similar species around the world like *Culex fatigans* [16] from the Old World tropics were used synonymous to *Cx. quinquefasciatus* [17, 18]. Females of *Culex pipiens* and *Cx. quinquefasciatus* are morphologically indistinguishable and hybrid zones for the two species are well documented. Owing to this, *Culex quinquefasciatus* has been considered and designated as a subspecies of *Cx. pipiens* with the name *Culex pipiens quinquefasciatus* [19]. Studies have indicated that *Culex pipiens* and *Cx. quinquefasciatus* are two distinct sympatric mosquito populations [20] and they also exhibit a distinct and clear genetic difference [21] which led to the elevation of *Cx. quinquefasciatus* to a species status.

### 3. Origin and spread of *Culex quinquefasciatus*

Although previous studies stated that *Culex quinquefasciatus* was native to the low land regions of West Africa from where the species has been spread to tropical and warm temperate regions by human activity [22, 23], recent studies indicated that *Culex quinquefasciatus* originated in Southeast Asia and then established in the New World through slave ships and colonized Africa [24]. There are four major phases that are thought to have involved in the global expansion of this species. First of all, an introduction of *Culex quinquefasciatus* to the New world and Australia in lieu of its spread in the Old World tropics through sailing vessels included in the slave trade before 1800 [22]. Secondly, this species expanded to New Zealand, the islands of Hawaii, the Seychelles and archipelagos in the Pacific and Indian oceans through sailing vessels linked with the whaling industry of the United States of America during 19<sup>th</sup> century [25]. Thirdly, during World War II, the expansion of this species occurred to small Pacific atolls of military significance by army aircraft and ships [26]. Finally, this species expanded to the remote Pacific islands and Indian oceans through commercial airline travel [27].

## 4. Bio-ecology of Southern House Mosquito, *Culex quinquefasciatus*

### 4.1. Reproductive Biology and Breeding Behaviour

Gravid *Cx. quinquefasciatus* females lay a single egg raft averaging 155 eggs during each gonotrophic cycle; the number of eggs depends on mosquito age, blood source and blood volume [28]. Egg rafts are laid on the surface of a suitable water body selected, using the chemical cues derived from the conspecific egg rafts [29]. Larval to adult development is dependent on temperature, nutrition and population density and can be as short as at 7 days under optimal conditions (30 °C) [30]. Females mate within 2-6 days of emergence and may begin to seek hosts within 48 hours of emergence [28]. The duration of larval stages was 118 hours for males and 135 hours for females [31]. Females of *Cx. quinquefasciatus* emerge in large number than males in the regions where the seasons are more distinct [32]. Since this mosquito must require blood meal for reproduction and does not undergo a reproductive diapause, hence this species is active and reproduces year-round. In India, it may complete 2-3 gonotrophic cycles in a lifetime during the hotter season and 4-8 cycles in the cooler season [33]. It has been observed that infection with *Wuchereria bancrofti* could disrupt the relationship between mosquito size and egg production during the first gonotrophic cycle of *Culex quinquefasciatus* [33]. This mosquito was found in domestic collections of water and in places like flooded open cement drains, flooded latrines, overflow water from houses, kitchens, as well as in ground-pools, ditches and shallow wells. This species usually selects organically rich and polluted surface

waters and artificial containers for breeding [6]. The species also breeds in shallow ponds, streams, phytotelmata [34] and also in artificial habitat such as drains, wells, septic tanks and other small containers [35]. While conducting a survey in metro rail construction sites in Kolkata, researchers found that *Culex quinquefasciatus* mainly preferred polluted water for breeding in winter and post-winter months [36]. In North Central Nigeria, Africa the Bancroftian filariasis is transmitted by *Culex quinquefasciatus* in urban and semi-rural areas where increased pollution of freshwater bodies and the introduction of pit latrines favour the breeding of the mosquito [37, 38]. A wide variety of sites, mostly characterized by coloured, foul water with high nutrient values and low dissolved oxygen content, such as pumping and irrigation wells, canals, wastewater treatment ponds, sewage overflows, rain pools, rice paddy fields, fish ponds, septic tanks, drains, cesspools, agricultural trenches, vegetable farms etc. generally are preferred as the breeding sites by this mosquito [39, 40].

### 4.2. Biting Behavior

*Culex quinquefasciatus* has a predilection for urban environments and feeds on human indoors as well as outdoors [41]. This mosquito is nocturnal and was found to be predominant of the total mosquitoes caught off human baits in Calcutta and adjoining South and North 24 Parganas districts [42, 43]. It constitutes 97% of the nocturnal man-biting mosquitoes of Calcutta [44]. In addition to human, the female can efficiently bite amphibians [4], pigs, horses, cattle, sheep, dogs, rabbits [14]. The annual transmission potential of bancroftian filariasis higher in the urban area than that in the rural areas [45]. *Culex quinquefasciatus* is considered as an opportunistic feeder in rural Bengal and feeds on blood of human (26.45%), ruminant (46.25%), pig (14.19%) and bird (6.45%) [46]. In Southern India this is a highly anthropophilic species, with 50-76% feeding on human [47]. Interestingly, it has been found that the attraction ratio of *Cx. quinquefasciatus* of man: cow was 101.1:1 in Burdwan district, West Bengal [48].

### 4.3. Seasonal Effects on population density

In tropical areas, population of *Culex quinquefasciatus* reaches the peak density during or just following the rainy season [28]. Interestingly, the peak population was found in the warmest months of the year in subtropical and warm temperate areas [49, 50]. In summer, the higher man hour density of this vector mosquito has been reported in Bankura, West Bengal [51]. Adult female *Cx. quinquefasciatus* could survive till one month with a source of carbohydrates and at a constant temperature of 28 °C [52]. It has been documented that at lower temperatures (15 °C), longevity is increased dramatically [53].

### 4.4. Physico-chemical factors

The embryonic development of *Cx. quinquefasciatus* requires the optimal temperature ranging from 24 to 29 °C. It has been found that at Low ( $\leq 12$  °C) and high ( $\geq 32$  °C) temperatures the survivorship drops with an upper lethal threshold at 35 °C. Slight alkaline water ( $< \text{pH } 8$ ) with little ( $\leq 0.5\%$ ) NaCl, CaCl<sub>2</sub> and NaCO<sub>3</sub> are the favourable condition for the larvae [49].

### 4.5. Associations with other species

Many other mosquito species might co-exist with *Cx. quinquefasciatus* in peridomestic habitat, such as *Culex nigripalpus* in the southern United States [54] and *Culex australicus*, *Culex annulirostris*, *Culex pervigilans*, *Aedes polynesiensis*, *Aedes notoscriptus*, *Aedes hebrideus*, *Aedes*

*pernotatus* and *Tripteroides melanesiensis* in the tropical Pacific [35, 55]. The larvae of *Culex quinquefasciatus* are occasionally found with *Aedes aegypti* and *Aedes albopictus* in domestic and peridomestic water containers [28]. *Cx. quinquefasciatus* larvae are often found in association with psychodid moth fly larvae in septic habitats [54, 56].

## 5. Genetics and Molecular Biology

*Cx. quinquefasciatus* has been extensively used for considerable genetic research because of its long-standing taxonomic controversy and importance as a vector [57]. The species contains three metacentric chromosomes among which Chromosome 1 is considered the shortest, Chromosome 2 is intermediate in length and Chromosome 3 is the longest [58]. The complete genome sequencing studies of *Culex quinquefasciatus* reveal that the number of their protein coding genes (18,883) is 22% greater than that of *Aedes aegypti* and 52% greater than that of *Anopheles gambiae*. In addition to this, the species exhibit multiple gene-family expansion which include olfactory and gustatory receptors, salivary gland genes and genes associated with xenobiotic detoxification [59]. It has been reported that the large cytochrome P450 repertoire of this mosquito species has a significant role associated with the mosquito resistance [60, 61]. Interestingly, research showed that *Culex quinquefasciatus* has the largest number of olfactory receptor among dipteran species which explains culicine olfactory behavioural diversity which is in turn related with their host and oviposition site choice [59]. The ability of this mosquito species to feed on birds, humans and livestock indicates that it has a large number of proteins which could increase their capacity to imbibe blood from diverse hosts [59]. In this context, it could be mentioned that researchers have also discovered a 16.7 kDa family of proteins following salivary transcriptome analysis [62].

## 6. *Culex quinquefasciatus* as an Invasive species

*Culex quinquefasciatus* has reputation as an important invasive species which exhibits significant detrimental impacts on resident species or ecosystem as well as human or vertebrate animal health [63]. Interestingly, being an urban mosquito, *Culex quinquefasciatus* has significantly also invaded lowland rainforest [64]. Moreover, the desiccation resistant egg is a significant characteristic of invasiveness of a species [63]. The serosal cuticle contributes to mosquito egg desiccation resistance [65]. The serosa has been considered as a protective layer around the insect egg. It secretes a chitinized cuticle, the serosal cuticle underneath the maternal eggshell [66, 67]. In mosquitoes, desiccation resistance coincides with the time of serosal cuticle secretion [68]. Studies indicated that *Culex quinquefasciatus* acquire desiccation resistance ability during embryogenesis through the formation of the serosal cuticle [69].

## 7. Insecticide Resistance

*Culex quinquefasciatus* control was mainly conducted through the use of neurotoxic insecticides belonging to the Organochlorines (OC), the Organophosphates (OP) and the Pyrethroids (PYR) families in the western Indian Ocean islands [70, 71, 72]. The larvae of *Culex quinquefasciatus* used to grow easily in sewers and waste water collections [73] where in addition to insecticides, they are also subject to a wide range of xenobiotics. The mosquito control failures in the field have been resulted from resistant *Culex quinquefasciatus* individuals [74]. The enzymatic detoxification (i.e. metabolic resistance) and target site modification are the two major insecticide resistance mechanisms found in mosquitoes [74].

The most common target modification in *Culex quinquefasciatus* are the L1014F mutation (kdrR allele) in the voltage-gated sodium channel gene, conferring resistance to PYR and DDT. The G119S ace-1 mutation (ace-1R allele) conferring resistance to OP and carbamates, while A302S Rdl mutation (RdlR allele) conferring resistance to the OC dieldrin [71, 75, 76].

## 8. Discussion

The importance of Southern House Mosquito, *Culex quinquefasciatus* as a mosquito vector from the public health standpoint relies on the fact that *Culex quinquefasciatus* is the principal vector of bancroftian filariasis and a potential vector of *Dirofilaria immitis* [9, 10, 77, 78]. It is also a competent vector of several arboviruses like West Nile virus (WNV) in northeastern United States [79] and Asia [80]. St. Louis encephalitis virus (SLEV) in eastern and south-central North America [13], Rift Valley fever virus [81], and several protozoa like *Plasmodium relictum* that causes bird malaria [82]. Studies revealed that this mosquito species can efficiently transmit protozoan *Hepatozoon breinli* in the laboratory setup. *Culex quinquefasciatus* is also an important vector of avian pox [83, 84, 85] and *Plasmodium cathemerium* [4], a protozoan parasite of birds. Additionally, this species has also the ability to transmit the other nematodes like *Saurofilaria sp.*, *Oswaldofilaria sp.* [86]. The Southern House Mosquito is considered as a cyclopropagative vector for avian malaria and a mechanical vector for avian pox in Hawaii [87]. This vector is a well known domestic mosquito species of rural, semi-urban and urban areas [86, 88] and also has the reputation as an important invasive species [63]. The cosmopolitan distribution of *Culex quinquefasciatus* is across continents and ecozones mostly south of 39 °N latitude. This suggests that this invasive mosquito species has the plasticity to adapt in diverse habitats and this could be expanded owing to its amplified immunity gene repertoire [1]. Interspecific competition with the resident species in newly introduced areas may be the possible ecological process that might help this mosquito to become as a significant invasive mosquito species. In newly invaded areas, this mosquito species might also expand because of the absence of the suitable competitors and predators [63] (Table 2). Interestingly, researchers have found substantial mosquito samples from container-type breeding sites (metal and plastic containers, “calabash”, and tyres) and least from phytotelmata [64]. The vast distribution of breeding sites (tires, plastic and metal containers, calabashes, puddles, leaf axiles, depressions on trees, etc.) possibly indicates the significant changes in the oviposition behaviour of *Culex quinquefasciatus* [64]. Studies indicated that the habitat quality in which mosquitoes are oviposited is the major factor behind enhanced offspring survival and growth. Thus, when the potential habitats differ in their suitability for juveniles, females used to select the habitats which can maximize the fitness of the juveniles [89]. The oviposition habitat selection in mosquitoes have been demonstrated in response to physical and chemical suitability for the larval development [90], which in turn might be the possible reason behind the variation in preference among metallic, plastic containers and calabashes as the suitable oviposition habitat [63]. Moreover, *Culex quinquefasciatus* mosquito can be able to survive in polluted waters where there are no natural predators, which leads to the excessive growth of their population [91]. *Culex quinquefasciatus*, thus indicates its adaptive flexibility [92, 93] and ecological plasticity. The changing climate might be associated with the substantial increase in the vector distribution, enhanced transmission

potential especially in temperate climates, elongation of transmission season and shortening of vector gonotrophic cycle and incubation time [94]. *Culex quinquefasciatus* is a nocturnal mosquito [4]. Interestingly, the biting density, natural infection and infectivity rates of *Culex quinquefasciatus* were found significantly higher in the third quadrant of the night. This was true both in urban and rural environments [95]. Researchers demonstrated that the human foot region attracts *Culex quinquefasciatus* more than other parts of the exposed lower limb of the human host [96] (Figure. 1). This mosquito is thought to be capable of transmitting WNV [97].

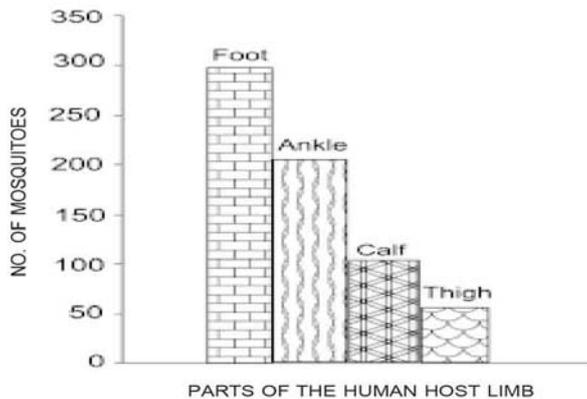


Fig 1. *Culex quinquefasciatus* attracted to different parts of the human limb [96].

Although some members of *Culex pipiens* complex bite only birds and other members bite mainly mammals including humans [98], investigators reiterated that the Nearctic population of *Culex pipiens* complex feed on both bird and mammals. This selective feeding behaviour might be a possible reason behind the high frequencies of infection found in human, horse and bird by WNV in North America [99]. Albeit members of the *Culex vishnui* complex are important vectors of Japanese encephalitis [100], JE virus has also been isolated from *Culex quinquefasciatus* in several occasions [101, 102, 103]. Bhattacharya *et al.*, [100] demonstrated that *Culex vishnui* supported JE virus for a longer period than *Culex quinquefasciatus*, however the Southern House Mosquito was able to generate a notable titre level for a considerable period of time (Figure.2).

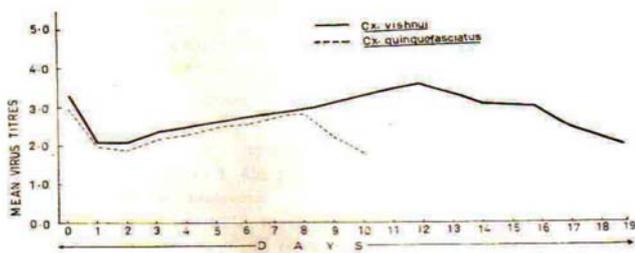


Fig 2. Duration of virus detection in *Culex vishnui* and *Culex quinquefasciatus* mosquitoes [100].

JE is predominantly a rural disease. Interestingly, JEV vectors are found in urban areas however, small in numbers [104]. Diverse pathogenic parasites, arbovirus and nematodes have been isolated from this mosquito species. Although the finding of any pathogen in particular species of mosquito is however not itself a proof that the species is a vector to the

man or animal. The attainment of the status of being called a potential vector, itself is a dynamic phenomenon and there is a possibility that with the passage of time, *Culex quinquefasciatus* may become epidemiologically significant for some isolated pathogens (Table. 1) which are presently not having any public health importance. The arbovirus like JEV, WNV, SLEV survive by virtue of alternation between vertebrate and invertebrate hosts in a "cycle" where man gets the infection tangentially on accidental intrusion in this pathway. Animal hosts such as pigs and birds are recognized animal reservoirs and play an important role in the maintenance and amplification of these viruses. The mosquito vectors may carry the virus for life after getting the infection [105]. The Southern House Mosquito acts as an important "urban bridge vector" [106] which bridges different reservoir/amplifier hosts to humans because of its encounter with different vertebrates. This species also creates an ecological bridge between urban, periurban and rural areas owing to its presence in diverse ecological niches. Mosquito borne zoonotic viral disease like JE, is historically a rural disease which may spill over to urban areas in the changing ecological conditions; in such a situation *Culex quinquefasciatus* could play an important role not only as a bridge vector between man and pigs/birds (reservoir host) but also bridging the rural-urban eco-epidemiological zones. Hence *Culex quinquefasciatus* may emerge as a potential vector of urban Japanese encephalitis. Thus, ubiquitous *Culex quinquefasciatus* can act as both way traffic for the dissemination of certain zoonotic arboviral diseases in different environments. *Culex quinquefasciatus* is one of the most frequently encountered domestic and peridomestic mosquito among the other *Culex sp.* and mostly found in aboard aircraft [107]. The anthropophilic and endophilic blood feeding habits bring this species close to human and human habitation [103]. Thus, the remarkable adaptive fitness, ecological plasticity, invasive behaviour, host specificity, and high reproductive potential [103] along with expanded immune gene repertoire property at the genetic level [1] have made *Culex quinquefasciatus* a smart vector. This species possesses the necessary potential to initiate and facilitate the disease transmission by establishing an effective vector-host transmission cycle for diverse pathogens in different environments. Thus, it could be relevant to mention that in the changing ecological conditions Southern House Mosquito might enhance its epidemiological importance in the near future as a smart vector for those pathogens which were isolated/detected from this mosquito species. There can be a possible association between the invasiveness of this mosquito and the increased risk of introduction of pathogens in newer areas [108]. In this context, it could be mentioned that the insecticide resistance of *Culex quinquefasciatus* is a matter of concern as it might increase the risk of resurgence of parasitic and arboviral outbreaks. Global warming and globalization are likely to reshape the ecology of vector mosquitoes, such as ubiquitous *Culex quinquefasciatus* which might have a wide ranging consequences on the epidemiology of the vector borne diseases. This notion is also important in the context of emergence and invasion of arbovirus like Zika virus. Thus, a continuous surveillance and molecular studies are required to ascertain the prospective role of Southern House Mosquito, *Culex quinquefasciatus* in the possible transmission of different pathogens.

**Table 1:** List of virus and Protozoan and Nematode Parasite found and isolated from *Culex quinquefasciatus* in natural and artificial conditions

Pathogens detected/isolated in <i>Culex quinquefasciatus</i>	References
<b>Virus</b>	
West Nile virus (Flaviviridae)	11
Japanese encephalitis virus (Flaviviridae)	12, 100, 101
St. Louis encephalitis virus (SLEV) (Flaviviridae)	13
Rift Valley fever virus (Bunyaviridae)	81
Edge Hill virus (Flaviviridae)	4, 14
Eubenangee virus (EUBV) (Reoviridae)	
Getah virus (Togaviridae)	
Kokohera virus (KOKV) (Flaviviridae)	
Koongol virus (Bunyaviridae)	
Kowanyama virus (Bunyaviridae)	
Kunjin virus (Flaviviridae)	
Mapputta virus (Bunyaviridae)	
Stratford virus (STRV) (Flaviviridae)	
Trubanaman virus (TRUV) (Bunyaviridae)	
Wongal virus (WONV) (Bunyaviridae)	
Chikungunya virus (Togaviridae)	
<b>Protozoa</b>	
<i>Hepatozoon breinli</i>	26, 86
<i>Plasmodium cathemerium</i>	4
<i>Plasmodium relictum</i>	82
<b>Nematode</b>	
<i>Wuchereria bancrofti</i>	3
<i>Dirofilaria immitis</i>	10
<i>Saurofilaria sp.</i>	86
<i>Oswaldofilaria sp.</i>	
<i>Brugia malayi</i>	36

**Table 2:** Features of invasiveness of *Culex quinquefasciatus* mosquito.

Location	Time of Introduction	Possible Ecological Processes	References
New Zealand	19 <sup>th</sup> Century	Absence of Competitors and predators	109
Australia	19 <sup>th</sup> Century	Interspecific competition via a gut symbiotic fungi	110, 111, 112
North merica	19 <sup>th</sup> Century	interspecific competition with native <i>Culex</i> may have contributed declines of <i>Cx. tarsalis</i> in California	113

## 9. Conflict of Interest

There is no conflict of interest.

## 10. Acknowledgements

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## 11. References

- Bartholomay LC, Waterhouse RM, Mayhew GF, Campbell CL, Michel K. Pathogenomics of *Culex quinquefasciatus* and meta-analysis of infection responses to diverse pathogens. *Science* 2010; 330(6000):88-90.
- Dobrotworsky NV. The Mosquitoes of Victoria. Melbourne University Press; London and New York; Cambridge University Press, 1965, 237.
- Bram RA. Contribution to the mosquito fauna of South East Asia II. The genus *Culex* in Thailand (Diptera: Culicidae). Contributions of the American Entomological Institute 1967; 2(1):1-296.
- Lee DJ, Hicks MM, Debenham ML, Griffiths M, Marks EN, Bryan JH *et al.* The Culicidae of the Australasian Region. Canberra, Australia: Australian Government Publishing Service 1989; 7:281.
- White GB. Notes on a catalogue of Culicidae of the Ethiopian region. *Mosquito Systematics* 1975; 7:303-44.
- Weinstein P, Laird M, Browne G. Exotic and Endemic Mosquitoes in New Zealand as Potential Arbovirus Vectors [occasional paper]. Wellington (NZ): Ministry of Health, 1997.
- Higgs S, Beaty BJ. Natural cycles of vector-borne pathogens. In: Marquardt, W.C., *et al.* (Eds.) *Biology of disease vectors*. Second. Ed., 2005, 167-185.
- Lindahl J. Japanese Encephalitis Virus in Pigs and Vectors in the Mekong Delta with Special Reference to Urban Farming. Published PhD Thesis. Swedish University of Agricultural Sciences, Uppsala, 2012.
- Belkin JN. Mosquito studies (Diptera, Culicidae). VII. The Culicidae of New Zealand. Contributions of the American Entomological Institute 1968; 3(1):1-28.
- Lai CH, Tung KC, Ooi HK, Wang JS. Competence of *Aedes albopictus* and *Culex quinquefasciatus* as vector of *Dirofilaria immitis* after blood meal with different microfilarial density. *Veterinary Parasitology* 2000; 90:231-237.
- Goddard LB, Roth AE, Reisen WK, Scott TW. Vector competence of California mosquitoes for West Nile Virus. *Emerging Infectious Diseases* 2002; 8(12):1385-1391.
- Reuben R, Tewari S, Hiriyani J, Akiyama J. Illustrated keys to species of *Culex* (Culex) associated with Japanese Encephalitis on Southeast Asia (Diptera: Culicidae) *Mosquito Systematics* 1994; 26(2):75-96.

13. Tsai TF, Mitchell CJ. St. Louis encephalitis, In: Monath TP (ed) The Arboviruses: Epidemiology and Ecology, CRC Press, Boca Raton, USA 1989; 4:113-143.
14. Holder P, Browne G, Bullians M. The mosquitoes of New Zealand and their animal disease significance. *Surveillance* 1999; 26(4):12-15.
15. Farajollahi A, Fonseca DM, Kramer LD, Kilpatrick AM. Bird biting mosquitoes and human disease: a review of the role of *Culex pipiens* complex mosquitoes in epidemiology. *Infection, Genetics and Evolution* 2011; 11(7):1577-1585.
16. Wiedemann CRW. Ausser europaische zweiflugelige Insekten. Hamm 1828; 1:608. (Cited in [http://entnemdept.ufl.edu/creatures/aquatic/southern\\_house\\_mosquito.htm](http://entnemdept.ufl.edu/creatures/aquatic/southern_house_mosquito.htm). visited on 10.04.2015).
17. Stone A. Corrections in the taxonomy and nomenclature of mosquitoes (Diptera: Culicidae). *Proceedings of the Entomological Society of Washington* 1956; 56(6):333-343.
18. Belkin JN. *Cx. quinquefasciatus* or *fatigans* for the tropical (southern) house mosquito (Diptera: Culicidae). *Proceedings of the Entomological Society of Washington* 1977; 79(1):45-52.
19. Barr AR. The Distribution of *Culex p. pipiens* and *Cx. p. quinquefasciatus* in North America. *American Journal of Tropical Medicine and Hygiene*. 1957; 6(1):153-165.
20. Cornel AJ, McAbee RD, Rasgon J, Stanich MA, Scott TW, Coetzee M. Differences in extent of genetic introgression between sympatric *Culex pipiens* and *Culex quinquefasciatus* (Diptera: Culicidae) in California and South Africa. *Journal of Medical Entomology*. 2003; 40(1):36-51.
21. Smith JL, Fonseca DM. Rapid assays for identification of members of the *Culex (Culex) pipiens* complex, their hybrids, and other sibling species (Diptera: Culicidae). *American Journal of Tropical Medicine and Hygiene*. 2004; 70(4):339-345.
22. Belkin JN. The Mosquitoes of the South Pacific (Diptera, Culicidae) Univ of California Press, 1962; 1:608.
23. Vinogradova EB. *Culex pipiens pipiens* mosquitoes, taxonomy, distributions, ecology physiology genetics, applied importance and control. Sofia, Bulgaria: Pensoft. 2000, 646.
24. Fonseca DM, Smith JL, Wilkerson RC, Fleischer RC. Pathways of expansion and multiple introductions illustrated by large genetic differentiation among worldwide populations of the southern house mosquito. *American Journal of Tropical Medicine and Hygiene*. 2006; 74(2):284-289.
25. Lounibos LP. Invasions by insect vectors of human disease. *Annual Review of Entomology* 2002; 47:233-266.
26. Ward RA. Mosquito fauna of Guam: case history of an introduced fauna. In: Commerce and the spread of pests and disease vectors [ed. by Laird, M.]. New York: Praeger Publishers, USA, 1984; 143-162.
27. Bataille A, Cunningham AA, Cedeño V, Cruz M, Eastwood G, Fonseca DM *et al.* Evidence for regular ongoing introductions of mosquito disease vectors into the Galápagos Islands. *Proceedings of the Royal Society of London. Series B, Biological Sciences* 2009; 276(1674):3769-3775.
28. Subra R. Biology and control of *Culex pipiens quinquefasciatus* Say, 1823 (Diptera, Culicidae) with special reference to Africa. *Insect Science and its Application* 1981; 1(4):319-338.
29. Laurence BR, Pickett JA. An oviposition attractant pheromone in *Culex quinquefasciatus* Say (Diptera: Culicidae). *Bulletin of Entomological Research* 1985; 75(2):283-290.
30. Rueda LM, Patel KJ, Axtell RC, Stinner RE. Temperature-dependent development and survival rates of *Culex quinquefasciatus* and *Aedes aegypti* (Diptera: Culicidae). *Journal of Medical Entomology*. 1990; 27(5):892-898.
31. Meillon B, De Sebastian A, Khan ZH. The duration of egg, larval and pupal stages of *C. pipiens fatigans* in Rangoon, Burma. *Bull. Wld. Hlth. Org.* 1967a; 36:7-14.
32. Hayes J. Seasonal changes in population structure of *Culex pipiens quinquefasciatus*, Say (Diptera: Culicidae). Study of an isolated population. *J Med Ent.* 1975; 12:167-178.
33. Lima CA, Almeida WR, Hurd H, Albuquerque CMR. Reproductive aspects of the Mosquito *Culex quinquefasciatus* (Diptera: Culicidae) infected with *Wuchereria bancrofti* (Spirurida: Onchocercidae). *Mem Inst Oswaldo Cruz, Rio de Janeiro* 2003; 98(2):217-222.
34. Derraik JGB. Mosquitoes breeding in phytotelmata in native forest in the Wellington region, New Zealand. *New Zealand Journal of Ecology*. 2005; 29(2):185-191.
35. Laird M. Background and findings of the 1993-94 New Zealand mosquito survey. *New Zealand Entomologist* 1995; 18:77-90.
36. Chatterjee KK, Biswas D, Chandra G, Bhattacharyya A, Ghosh A, Hati AK. A longitudinal survey on mosquito larvae in relation to metro rail construction in Calcutta. *Indian Med Gaz.* 1988; CXXII(2):56-59.
37. Badaki JA. Parasitological and social aspects of Lymphatic filariasis in Taraba State. Ph.D Thesis in the Department of Zoology, University of Jos, Nigeria, 2010.
38. Service MW. *Medical entomology for students*. Edn 5, Cambridge University Press, New York, 2012, 303.
39. Matthys B, N'Goran EK, Koné M, Koudou BG, Vounatsou P, Cisse G. Urban agricultural land use and characterization of mosquito larval habitats in a medium-sized town of Côte d'Ivoire. *J Vector Ecol.* 2006; 31(2):319-333.
40. Opoku AA, Ansa-Asare OD, Amoako J. The occurrences and habitat characteristics of mosquitoes in Accra, Ghana. *West African J Appl Ecol.* 2007; 11:81-86.
41. Sirivanakarn S. *Medical entomology studies-III. A revision of the subgenus Culex in the Oriental region* (Diptera: Culicidae). *Contrib. Am. Entomol. Inst. (Ann Arbor)* 1976, 12(2):1-272.
42. Bhattacharya S, Hati AK. Man-*Culex quinquefasciatus* contact in relation to transmission dynamics of Bancroftian filariasis. Abstract book, 6th International congress of Parasitology held in Brisbane, Australia, 1986.
43. Bhattacharya S, Santra SC. Mosquito-borne diseases and vector diversity in Kolkata with special reference to malaria. In: *Perspectives in Environmental Health- Vector and Water-Borne diseases* (Eds: A. Mukhopadhyay and A K De) Published by Originals, Delhi-110052, 2005, 53-70.
44. Mukhopadhyay AK, Hati AK. Man-biting activity of *Anopheles stephensi* in Calcutta. *Bull. Cal. Sch. Trop. Med.* 1978; 26:5-7.
45. Hati AK, Chandra G, Bhattacharyya A, Biswas D, Chatterjee KK, Dwivedi HN. Annual transmission

- potential of bancroftian filariasis in an urban and a rural area of West Bengal, India. *Am J Trop Med Hyg.* 1989; 40(4):365-367.
46. Bhattacharya S, Chakraborty S, Mitra AK, Ghosh KK, Das S, Chakraborty MS *et al.* Blood meal analysis of certain species of mosquitoes of rural West Bengal. *Bull. Cal. Sch. Trop. Med.* 1982; 30(2):43-45.
  47. Reuben R, Thenmozhi V, Samuel P, Gajanana A, Mani T. Mosquito blood feeding patterns as a factor in the epidemiology of Japanese encephalitis in southern India. *American Journal of Tropical Medicine and Hygiene.* 1992; 46(6):654-663.
  48. Chakraborty S, Choudhuri DK, Bhattacharya S, Hati AK. Bait preference pattern of *Culex vishnui* mosquito in rural West Bengal. *Bull. Cal. Sch. Trop. Med.* 1986; 34(1-4):15-17.
  49. Mitchell CJ, Francy DB, Monath TP. Arthropod vectors. In: *St. Louis encephalitis* [ed. by Monath, T. P.]: Chapter 7, American Public Health Association, Inc, Washington, DC, USA, 1980, 313-373.
  50. Ahumada JA, Lapointe D, Samuel MD. Modeling the population dynamics of *Culex quinquefasciatus* (Diptera: Culicidae), along an elevational gradient in Hawaii. *Journal of Medical Entomology.* 2004; 41(6):1157-1170.
  51. Pramanik M, Chandra G. Studies on seasonal fluctuation of different indices related to filarial vector, *Culex quinquefasciatus* around foothills of Susunia of West Bengal, India. *Asian Pacific Journal of Tropical Medicine.* 2010; 3(9):727-730.
  52. Vrzal EM, Allan SA, Hahn DA. Amino acids in nectar enhance longevity of female *Culex quinquefasciatus* mosquitoes. *Journal of Insect Physiology.* 2010; 56(11):1659-1664.
  53. Gunay F, Alten B, Ozsoy ED. Estimating reaction norms for predictive population parameters, age specific mortality, and mean longevity in temperature-dependent cohorts of *Culex quinquefasciatus* Say (Diptera: Culicidae). *Journal of Vector Ecology.* 2010; 35(2):354-362.
  54. Hribar LJ, Vlach JJ, DeMay DJ, James SS, Fahey JS, Fussell EM. Mosquito larvae (Culicidae) and other Diptera associated with containers, storm drains, and sewage treatment plants in the Florida Keys, Monroe County, Florida. *Florida Entomologist* 2004; 87(2):199-203.
  55. Laird M. The natural history of larval mosquito habitats: Academic Press Ltd, xxvii, London, UK, 1988, 555.
  56. Su TY, Webb JP, Meyer RP, Mulla MS. Spatial and temporal distribution of mosquitoes in underground storm drain systems in Orange County, California. *Journal of Vector Ecology.* 2003; 28(1):79-89.
  57. Fonseca DM, Smith JL, Wilkerson RC, Fleischer RC. Pathways of expansion and multiple introductions illustrated by large genetic differentiation among worldwide populations of the southern house mosquito. *American Journal of Tropical Medicine and Hygiene.* 2006; 74(2):284-289.
  58. McAbee RD, Christiansen JA, Cornel AJ. A detailed larval salivary gland polytene chromosome photomap for *Culex quinquefasciatus* (Diptera: Culicidae) from Johannesburg, South Africa. *Journal of Medical Entomology.* 2007; 44(2):229-237.
  59. Arensburger P, Megy K, Waterhouse RM, Abrudan J, Amedeo P, Antelo B *et al.* Sequencing of *Culex quinquefasciatus* establishes a platform for mosquito comparative genomics. *Science* (Washington). 2010; 330(6000):86-88.
  60. Kasai S. Molecular cloning, nucleotide sequence and gene expression of a cytochrome P450 (CYP6F1) from the pyrethroid resistant mosquito, *Culex quinquefasciatus* Say. *Insect Biochem Mol Biol.* 2000; 30:163-71.
  61. Komagata O. Overexpression of cytochrome P450 genes in pyrethroid-resistant *Culex quinquefasciatus*. *Insect Biochem Mol Biol.* 2010; 40:146-152.
  62. Ribeiro JMC, Arcà B. From sialomes to the sialoverse: An insight into the salivary potion of blood feeding insects. *Adv Insect Physiol* 2009; 37:59-118
  63. Juliano SA, Lounibos LP. Ecology of invasive mosquitoes: effects on resident species and on human health, *Ecology Letters.* 2005; 8:558-574.
  64. Okiwelu SN, Noutcha MAE. Breeding Sites of *Culex quinquefasciatus* (Say) during the Rainy Season in Rural Lowland Rainforest, Rivers State, Nigeria. *Public Health Research* 2012; 2(4):64-68.
  65. Jacobs Chris GC. Surviving embryogenesis: the extraembryonic serosa protects the insect egg against desiccation and infection, Leiden University dissertation, 2014, 7-13.
  66. Hinton HE. *The Biology of Insect Eggs.* (1st ed.). Oxford: Pergamon Press, 1981.
  67. Lamer A, Dorn A. The serosa of *Manduca sexta* (Insecta, Lepidoptera): ontogeny, secretory activity, structural changes, and functional considerations. *Tissue & Cell* 2001; 33(6):580-595.
  68. Goltsev Y, Rezende GL, Vranizan K, Lanzaro G, Valle D, Levine M. Developmental and evolutionary basis for drought tolerance of the *Anopheles gambiae* embryo. *Developmental Biology* 2009; 330(2):462-470.
  69. Vargas HC, Farnesi LC, Martins AJ, Valle D, Rezende GL. Serosal cuticle formation and distinct degrees of desiccation resistance in embryos of the mosquito vectors *Aedes aegypti*, *Anopheles aquasalis* and *Culex quinquefasciatus*. *J Insect Physiol.* 2014; 62:54-60.
  70. Robert V, Lagneau C. Mission de conseil et d'appui à la mise en place de la lutte intégrée contre les vecteurs à Mayotte. IRD EID Méditerranée. 2009, 53.
  71. Tantely ML, Tortosa P, Alout H, Berticat C, Berthomieu A. Insecticide resistance in *Culex pipiens quinquefasciatus* and *Aedes albopictus* mosquitoes from La Reunion Island. *Insect Biochem Mol Biol.* 2010; 40:317-324.
  72. Ministry of Health and Quality of Life Mauritius, World Health Organization, University of California, San Francisco, Eliminating Malaria, Case-study 4. *Prev Reintroduction Mauritius GENEVA*, 2012, 75.
  73. Vinogradova EB. *Culex pipiens pipiens* mosquitoes: taxonomy, distribution, ecology, physiology, genetics, applied importance and control. Pensoft Publishers, 2000.
  74. Pocquet N, Milesi P, Makoundou P, Unal S, Zumbo B. Multiple Insecticide Resistances in the Disease Vector *Culex p. quinquefasciatus* from Western Indian Ocean. *PLoS ONE* 2013; 8(10):e77855.
  75. Weill M, Lutfalla G, Mogensen K, Chandre F, Berthomieu A. Comparative genomics: Insecticide resistance in mosquito vectors. *Nature* 2003; 423:136-137.
  76. Martinez-Torres D, Chevillon C, Brun-Barale A, Berge JB, Pasteur N. Voltage-dependent Na<sup>+</sup> channels in pyrethroid-resistant *Culex pipiens* L mosquitoes. *Pestic Sci.* 1999; 55:1012-1020.

77. Chandra G, Seal B, Hati AK. Age composition of the filarial vector *Culex quinquefasciatus* (Diptera: Culicidae) in Calcutta, India. *Bulletin of Entomological Research* 1996; 86(3):223-226.
78. Farid HA, Hammad MM, Morsy ZS, Kamal IH, Weil GJ, Ramzy RMR. Detection of *Wuchereria bancrofti* in mosquitoes by the polymerase chain reaction: a potentially useful tool for large scale control programmes. *Trans Royal Soc Trop Med Hyg.* 2001; 95:29-32
79. Lanciotti RS, Roehrig JT, Deubal V. Origin of the West Nile virus responsible for an outbreak of encephalitis in the northeastern United States. *Science* 1999; 286:2333-7.
80. Hubálek Z, Halouzka J. West Nile fever--a reemerging mosquito-borne viral disease in Europe, *Emerg Infect Dis.* 1999; 5(5):643-650.
81. Meegan JM. The Rift Valley fever epizootic in Egypt 1977-78. 1. Description of the epizootic and virological studies. *Trans R Soc Trop Med Hyg.* 1979; 73:618-623.
82. Atkinson CT, Woods KL, Dusek RJ. Wildlife disease and conservation in Hawaii: pathogenicity of avian malaria (*Plasmodium relictum*) in experimentally infected Iiwi (*Vestiaria coccinea*). *Parasitology* 1995; 111:S59-S69.
83. Derraik JGB. A survey of the mosquito (Diptera: Culicidae) fauna of the Auckland Zoological Park. *New Zealand Entomologist* 2004; 24:51-55.
84. Derraik JGB, Slaney D. Container aperture size and nutrient preferences of mosquitoes (Diptera: Culicidae) in the Auckland region, New Zealand. *Journal of Vector Ecology.* 2005; 30(1):73-82.
85. Laird M. New Zealand's mosquito fauna in 1995: History and Status. University of Auckland, 1996, 1-25.
86. New Zealand Biosecure Entomological laboratory Report *Culex (Culex) quinquefasciatus* (Say), New Zealand Status, Version, 2008; 2:1.
87. Atkinson CT, Dusek RJ, Woods KL, Iko WM. Pathogenicity of avian malaria in experimentally-infected Hawaii Amakihi. *J Wildl Dis.* 2000; 36:197-204.
88. WHO. Report of the WHO is formal consultation on the evaluation on the testing of insecticides CTD / WHO PES / IC / 96.1, 1972, 69.
89. Kiflawi M, Blaustein L, Mangel M. Predation-dependent oviposition habitat selection by the mosquito *Culiseta longiareolata*: a test of competing hypotheses. *Ecol. Lett.* 2003; 6:35-40.
90. Trimble RM. Laboratory observations on oviposition by the predaceous tree hole mosquito, *Toxorhynchites rutilus septentrionalis*. *Canadian J Zool.* 1979; 57:1104-1108.
91. Dibo MR, Menezes RM, Ghirardelli CP, Mendonça AL, Chiaravalloti Neto F. The presence of Culicidae species in medium-sized cities in the State of São Paulo, Brazil and the risk of West Nile fever and other arbovirus infection. *Rev Soc Bras Med Trop.* 2011; 44:496-503.
92. Wilke ABB, Vidal PO, Suesdek L Marrelli. Population genetics of Neotropical *Culex quinquefasciatus* (Diptera: Culicidae). *Parasites & Vectors* 2014; 7:468.
93. Karlekar SR, Deshpande MM, Andrew RJ. Present susceptibility status Of *Culex quinquefasciatus*, Say to three insecticides in Nagpur district of India, *Ind. J Sci Res and Tech.* 2013; 1(2):12-14.
94. Weaver SC, Reisen WK. Present and Future Arboviral Threats, *Antiviral Res.* 2010; 85(2):328-36.
95. Chandra G, Majumdar G, Hati AK. An alternative approach for assessing filarial endemicity. In: *Studies on some vectors of public health importance* (Editor: AK Hati) Published by the Department of Medical Entomology, Calcutta School of Tropical Medicine, Govt. West Bengal, 1995, 146-148.
96. Oduola AO, Awe OO. Behavioural biting preference of *Culex quinquefasciatus* in human host in Lagos metropolis Nigeria *J Vect Borne Dis.* 2006; 43:16-20.
97. Hill S, Connelly CR. Southern House Mosquito *Culex quinquefasciatus* Say 2013; EENY-457 (University of Florida, IFAS Extension), 1-5. Retrieved from: <https://edis.ifas.ufl.edu/pdffiles/IN/IN83700.pdf> (Accessed on: 04-01-2016)
98. Fonseca DM, Keyghobadi N, Malcolm CA, Mehmet C, Schaffner F, Mogi M *et al.* Emerging vectors in the *Culex pipiens* complex. *Science.* 2004; 303:1535-1538.
99. Kuno G, Chang G-JJ. Biological Transmission of Arboviruses: Reexamination of and New Insights into Components, Mechanisms, and Unique Traits as Well as Their Evolutionary Trends. *Clinical Microbiology Reviews.* 2005; 18(4):608-637.
100. Bhattacharya S, Chakraborty S, Ghosh KK, Chakraborty MS, Mitra P, Chakraborty SK *et al* Persistence of Japanese Encephalitis Virus in *Culex vishnui* and *Culex quinquefasciatus* Mosquitoes-a Comparative Experimental Study, *Feedback-A Newsletter, Regional Computer Centre, Jadavpur University Campus, Calcutta.* 1987; 10(7):3-6.
101. Mourya DT, Ilkal MA, Mishra AC, Jacob PG. Isolation of Japanese encephalitis virus from mosquitoes collected in Karnataka state, India from 1985 to 1987. *Trans R Soc Trop Med Hyg.* 1989; 83:550-2.
102. Thenmozhi V, Mariappan T, Krishnamoorthy R, Baskarn G, Krishnamoorthi R, Balaji T, Tyagi BK. A first note on Japanese encephalitis virus isolation from *Culex quinquefasciatus* Say in Northern West Bengal, *International Journal of Mosquito Research.* 2014; 1(1):1-4.
103. Nitatpattana N, Apiwathnasorn C, Barbazan P, Leemingsawat S, Yoksan S, Gonzalez JP. First isolation of Japanese encephalitis from *Culex quinquefasciatus* in Thailand. *Southeast Asian J Trop Med Public Health.* 2005; 36(4):875-8.
104. Lindhal J. Urbanization and Vector-Borne Disease Emergence – a Possibility for Japanese Encephalitis Virus. *British Journal of Virology.* 2014; 1(1):10-20.
105. Bhattacharya S, Basu P. Japanese Encephalitis Virus (JEV) infection in different vertebrates and its epidemiological significance: a Review. *International Journal of Fauna and Biological Studies* 2014; 1(6):32-37.
106. Do QH, Vu TQ, Huynh TK, Dinh QT, Deubel V. Current situation of Japanese encephalitis in the south of Vietnam, 1976-1992. *Trop. Med.* 1994; 36:202-214.
107. Smith A, Carter ID. International transportation of mosquitoes of public health importance. In: *Commerce and the spread of pests and disease vectors* [ed. by Laird, M.]. New York: Praeger Publishers, USA; 1984, 1-21.
108. Kilpatrick AM, Daszak P, Goodman SJ, Rogg H, Kramer LD, Cedeño V *et al.* Predicting pathogen introduction: West Nile virus spread to Galápagos. *Conservation Biology.* 2006; 20(4):1224-1231.
109. Weinstein P, Laird M, Browne G. Exotic and Endemic Mosquitoes in New Zealand as Potential Arbovirus Vectors. *Occas. Pap. Minist. Health, Wellington, New Zealand,* 1997, 16.
110. Mokany A, Shine R. Competition between tadpoles and mosquito larvae, *Oecologia.* 2003a; 135:615-620.

111. Mokany A, Shine R. Biological warfare in the garden pond: tadpoles suppress the growth of mosquito larvae. *Ecol. Entomol.* 2003b; 28:102-108.
112. Mokany, A. & Shine, R. Oviposition site selection by mosquitoes is affected by cues from conspecific larvae and anuran tadpoles. *Aust. Ecol.* 2003c; 28:33-37.
113. Smith PT, Reisen WK, Cowles DA. Interspecific competition between *Culex tarsalis* and *Culex quinquefasciatus*. *J Vector Ecol.* 1995; 20:139-146.