



E-ISSN: 2320-7078

P-ISSN: 2349-6800

JEZS 2019; 7(4): 270-274

© 2019 JEZS

Received: 01-05-2019

Accepted: 03-06-2019

Ezhil Vadhana P

Division of Animal Genetics,
ICAR - Indian Veterinary
Research Institute, Izatnagar,
Bareilly, Uttar Pradesh, India

Ameya Santhosh

Division of Animal Genetics,
ICAR - Indian Veterinary
Research Institute, Izatnagar,
Bareilly, Uttar Pradesh, India

Pooja GS

Division of Animal Genetics,
ICAR - Indian Veterinary
Research Institute, Izatnagar,
Bareilly, Uttar Pradesh, India

Vani A

Division of Animal Genetics,
ICAR - Indian Veterinary
Research Institute, Izatnagar,
Bareilly, Uttar Pradesh, India

Sushil Kumar

Division of Animal Genetics,
ICAR - Indian Veterinary
Research Institute, Izatnagar,
Bareilly, Uttar Pradesh, India

Correspondence**Ezhil Vadhana P**

Division of Animal Genetics,
ICAR - Indian Veterinary
Research Institute, Izatnagar,
Bareilly, Uttar Pradesh, India

FecB: A major gene governing fecundity in sheep

Ezhil Vadhana P, Ameya Santhosh, Pooja GS, Vani A and Sushil Kumar

Abstract

Sheep being the major source of income for majority of farming community, holds indispensable position in the livestock and livelihood security of farmers. Major source of income in sheep farming is gained through meat and wool production which can be increased by improving the reproduction efficiency of animals. Selection and Breeding are the two major and basic tools involved in improving the reproduction efficiency. Most of the economic traits have quantitative inheritance, governed by minor genes, however few fecundity genes are exception. Major genes linked with fecundity and fertility have been identified by many researchers. *FecB*, one among them is chiefly involved nowadays in many introgression programmes. Since low heritability of reproduction traits severely hampers the selection, direct selection is very difficult. Marker assisted selection using these major genes linked with reproduction traits can be a best alternative for conventional method of selection.

Keywords: Marker, *FecB*, sheep, fecundity, introgression

Introduction

Fertility being the indispensable parameter controls the production and biological efficiency of sheep ^[1]. Variation in fertility between and within breeds is controlled by both genetic and environmental factors. Sheep farming is facing a major problem for maximizing production with limited resources in India. With about 43 registered breeds the total sheep population in the country is 65.06 million numbers in 2012, declined by about 9.07% over census 2007 ^[2]. Sheep (*Ovis aries*) being versatile species, possess more than 900 different breeds differing extensively in their physiological characteristics including ovulation rate and fecundity ^[3]. Some major genes are responsible for high fecundity in sheep. Use of sheep with *FecB* gene in crossing experiments offers a quantum increase in fecundity in one generation and therefore crossing experiments with the Booroola Merino were undertaken in various countries to evaluate *FecB* and its effects in other breeds ^[4]. In recent years following identification of *FecB* in the Garole as the likely source of the original mutation, Since then, when Garole was found to be the source of *FecB*, the Garole has been used for crossing experiments with many breeds of sheep ^[5] in India. Several breeds also shown to carry gene fecundity genes, that have a major effect on fecundity.

In some occasions, major or closely linked minor genes contribute to the differences in the ovulation rate among the animals ^[6]. Molecular genetics work on sheep has identified the presence of prolificacy genes that have been linked to reproductive efficiency. Molecular genetics and marker assisted selection (MAS) have paramount importance in genetic improvement of reproductive efficiency ^[7] in sheep. In animal production, marker based investigation have led to identification of several SNPs for traits viz. reproduction ^[8, 9], production ^[10, 11], thermotolerance ^[12, 13] and resistance to diseases like tuberculosis ^[14, 15, 16, 17, 18, 19], paratuberculosis ^[20, 21, 22, 23] brucellosis ^[24] & mastitis ^[25], along with generalized immune response ^[26, 27, 28, 29, 30] traits. Multiple genes have been identified of having diversified effects on the prolificacy and efficacy of sheep's reproduction. Age of ewe at service and lambing, and litter size were economically important traits that can enhance the productive value of sheep in terms of improving the twinning rate indirectly improving the meat and wool production. Indirect selection of those marker genes that have been linked to the reproduction traits can be a best alternative for conventional method of direct selection in terms of time and cost incurred.

Fecundity genes

Mutations with major effects on ovulation rate and litter size have been identified in genes of the transforming growth factor beta (TGF- β) super family and a TGF- receptor,

namely bone morphogenetic protein (BMP-15), growth differentiation factor-9 (GDF9) and BMP receptor –IB (ALK6) [31, 32, 33] and some other genes have also been identified. *FecB* or BMPR1B (Bone Morphogenetic Protein Receptor 1B) or ALK6 (activin receptor-like kinase 6) present in chromosome 6, *FecX* or BMP15 (Bone morphogenetic protein 15) present in chromosome X, *FecG* or GDF9 (Growth Differentiation Factor 9) present in chromosome 5 and B4GALNT2 (beta-1,4-N-acetyl-galactosaminyl transferase 2) present in chromosome 11 p [31, 33, 34].

***FecB* or Booroola gene**

Origin of mutation was traced back by Turner [35] and expanded by Piper and Bindon [4]. Turner and Young [36] visualized high prolificacy of separate line of Booroola merino (Australia) compared to other merino flock and those animals were selected for breeding experiments for improving the prolificacy. Simultaneously with Australian Flock Registers they traced back and found that the origin of *FecB* should be from Garole Breed of India which arrived in Australia from Kolkata in 1792 so called Bengal sheep at that time. A major gene, located in the BMPR1B gene, which is linked to the increase in fecundity of Booroola Merino was named *FecB* by the Committee on Genetic Nomenclature of Sheep and Goats. Possess a mutation (A746G) in its coding region leading to amino acid substitution (Q249R) in its protein sequence. In Booroola ewes, *FecB* gene, which is mapped to sheep chromosome 6q23–31, has additive effect on ovulation rate and partially dominant effect on litter size, can be inherited as a single autosomal locus and is syntenic to human chromosome 4q21–25. Heterozygous carrier parents produced approximately 1.5 extra eggs, giving birth to approximately 1.0 extra lamb per lambing. Homozygous carriers produced about 3.0 extra eggs producing about 1.5 extra lambs per lambing [37]. In addition to its effect on granulosa cells, *FecB* also affects the development of oocyte and its function.

Mechanism of Action

The members of the transforming growth factor beta (TGF- β) superfamily of peptides including bone morphogenetic proteins (BMPs) and activins were bound for the coordination of development and homeostasis of many tissues in various animal species right from insects (fruit fly) to mammals [38]. The TGF- β signal transduction network possess a receptor serine/threonine kinase at the cell surface, after binding of a ligand to its type 2 receptor in concert with a type 1 receptor, leads to a formation of heterodimeric receptor complex and phosphorylation of the type 1 receptor. Once activated, the type 1 receptor phosphorylates a receptor-regulated Smad which then heterodimerizes with a common Smad (Smad-4) and moves to the nucleus where the Smad complex associates with nuclear transcription factors and activates transcription of target genes [39].

Granulosa cells and oocytes expresses BMPR1B through all the stages of development and to a lesser extent by the theca layer of antral follicles also [40]. It has been reported as a potent receptor for various BMP (Bone Morphogenetic Protein) factors [41]. Basal and FSH-stimulated progesterone production can be inhibited by Ovine granulosa cells from small antral follicles, BMP-2, 4 and 6 receptors. BMP-2, 4 and 6 also enhances gonadotropin-induced estradiol production and decreases LH-induced androstenedione production [42]. All the above discussed factors were

promising stimulators of theca cells proliferation. *FecB* mutation (Q249R), being hypothesized as a partial “loss of function” mutation, is associated with the loss of responsiveness of BMP-4 receptor to the inhibitory action of progesterone production [43], and hence by decreasing BMP activity indirectly increases the ovulation rate. Through loss of function activity, allows numerous maturation of LH-responsive follicles with lower number of granulosa cells, producing same amounts of estradiol and inhibin as non-carriers [44]. Marked differentiative response to BMP, gonadotropin, and IGF-I stimulation has been shown by the granulosa and theca cells of carrier ewes on tissue culture studies and thereby explains the mechanism of deregulating the normal follicle selection process by its profound effect in inducing precocious maturation of ovarian follicles. The mechanism by which the carrier ewes differs from the non-carriers are not because of the total number of antral follicles but due to the markedly extended recruitment period along with low prevalence of atresia which leads to maturation and ovulation of large number of small ovulatory follicles. Some research studies strongly denote that the effect of *FecB* on ovulation rate is by altering the response of ovary rather than the level of gonadotropins applied to the ovary.

***FecB* on Growth traits**

Some of the researches observed that the lambs carrying the Booroola mutation exhibit relatively lower birth weight and growth rates compared with non-carrier lambs [45,46]. Lamb survival also has been reported to be lower among Booroola merino cross sheep carrying mutation. Lambs of carrier ewes require comparatively higher energy and protein per kilogram in an average [47]. Since lambs carrying the Booroola mutation are often from large litters, it is not clear whether their low birth weight, reduced postnatal growth and smaller mature body size is the result of the lambs being born in large litters or the result of a direct negative effect of the *FecB* locus on growth ability. Quantitative trait locus for low weaning weight has been suggested to segregate with the booroola mutation [48]. And this may explain some of the inferiority in the growth rate of booroola lambs. Report says that the gene has negative impact on fetal development [49] resulting in comparatively low birth weight.

Metabolic effects of *FecB*

Global metabolomics studies showed that in carrier animals, the mutation has marked effect on FF (Follicular Fluid) than in OVS (Ovine Vein Serum). Changes in the FF might affect the growth of the developing oocyte. Biochemical studies showed that the higher oxidative pressure of rapidly dividing cells in the homozygous animals, resulted in a greater antioxidant capacity of the FF. Additionally, some key metabolites in ovine FF samples were found to significantly correlate with ovulation rate. Altogether, *FecB* mutation not only affects ovulation rate but also the metabolic profile of FF [50].

***FecB* on male reproductive system**

In addition to its effect on female reproduction, *FecB* also have its effect on male reproduction by significantly affecting percent linearity and rapid motility of sperms, which did not change significantly with age. Although, homozygous carrier animals have increased sperm concentration than heterozygous which in turn was higher than non-carriers, this effect was non-significant. The age and *FecB* mutation have

significant effects on velocity and percentage of motile sperms without influencing the sperm morphometry^[51]. A report says that *FecB* is also responsible for the rate of increase of FSH concentration in adult rams after castration^[52] which is the reason for higher sperm concentration of homozygous and heterozygous Garole x Malpura rams.

Breeds possessing *FecB*

A number of prolific breeds all over the world were screened for *FecB*, besides Garole and Booroola, the mutated gene was present in Javanese (Indonesia), Small Tailed Han (China)^[53,54,55], and Hu (China) sheep^[54,55,56]. *FecB* mutation in Garole^[57], Kendrapada^[58] and Nilagiri sheep^[59] were reported in India. Studies shows that the mutation in fixed only in the Garole and Hu sheep population but segregating in other breeds wherein which it is present and because of its segregating nature, it has spread to about 48 breeds and synthetics in about 19 different countries^[60].

Conclusion

By improving the prolificacy, economic contribution of the *FecB* gene, can be noticed as early as in first generation itself after introgression, where the farmers income is mainly based on the lamb production. Introgression of the Booroola gene has an obvious potential for quick and large improvement of sheep breeding through cross breeding programmes which can improve the economic status of farmers who depend entirely on Sheep farming. Being affected by Environmental conditions, Ewe parity, Breed effect, Other genes, Flock management, Nutrition and Uterine capacity Introgression into new breed has its own utility and disadvantages. Constant genetic profiling of various prolific breeds in search of genes associated with fecundity, fertility and prolificacy and their molecular characterization can be done for better understanding. Detailed research is needed to search and evaluate the mutations in other fragments of the major genes and related genes in various other breeds and populations such that marker assisted selection can be rendered as a great tool for increasing the prolificacy of sheep and thereby helping the farmers for increasing their livelihood security.

References

1. Notter DR. Effects of ewe age and season of lambing on prolificacy in US Targhee, Suffolk and Polypay sheep. *Small Ruminant Research*. 2000; 38(1):1-7.
2. Sagar NG, Kumar S, Baranwal A, Prasad AR. Introgression of fecundity gene (*FecB*) in non-prolific sheep breeds: a boon for farmers. *International Journal of Science, Environment and Technology*. 2017; 6(1):375-380.
3. Terrill CE. The distribution of breeds of sheep as related to domestication and development of modern genotypes. In *The Domestication of Sheep: Their Ancestors, Geography, Time Period, and People Involved*. The International Sheep and Goat Institute, Utah State University, Logan, Utah, 1979, 41-112.
4. Piper LR, Bindon BM. The Booroola Merino. In: Fahmy MH (ed.) *Prolific sheep*, Wallingford, UK. CAB International, 1996, 152-160.
5. Nimbkar C, Ghalsasi PM, Nimbkar BV, Walkden-Brown SW, Maddox JF, Gupta VS *et al*. Reproductive performance of Indian crossbred Deccani ewes carrying the *FecB* mutation. In *Proceedings of the Association for the Advancement of Animal Breeding and Genetics*.

- 2007; 17:430-433.
6. Davis GH. Major genes affecting ovulation rate in sheep. *Genetics Selection Evolution*. 2005; 37(1):11.
7. Abdoli R, Zamani P, Deljou A, Rezvan H. Association of BMPR-1B and GDF9 genes polymorphisms and secondary protein structure changes with reproduction traits in Mehraban ewes. *Gene*. 2013; 524(2):296-303.
8. Yathish HM, Kumar S, Chaudhary R, Mishra C, Sivakumar A, Kumar A *et al*. Nucleotide variability of protamine genes influencing bull sperm motility variables. *Animal Reproduction Science*. 2018; 193:126-139
9. Chauhan IS, Gupta AK, Khate K, Chauhan A, Rao TK, Pathak S *et al*. Genetic and non-genetic factors affecting semen production traits in Karan Fries crossbred bulls. *Tropical Animal Health and Production*. 2010; 42(8):1809-15.
10. Kumar A, Singh RV, Chauhan A, Ilayakumar K, Kumar S, Kumar A *et al*. Genetic association analysis reveals significant effect of β -casein A1/A2 loci and seasonal/periodic variations on production & reproduction traits in Frieswal crossbred cows. 2019. DOI:10.1080/09291016.2019.1571705
11. Datta S, Adhikari AMJB, Chauhan A, Verma A, Gupta ID, Chauhan I *et al*. Nucleotide Sequence Variation in Leptin Gene of Murrah Buffalo (*Bubalus bubalis*). *Exploratory Animal and Medical Research*. 2012; 2(2):130-36.
12. Kashyap N, Kumar P, Deshmukh B, Dige MS, Sarkar M, Kumar A *et al*. Influence of ambient temperature and humidity on ATP1A1 gene expression in Tharparkar and Vrindavani cattle. *Indian Journal of Animal Research*. 2014; 48(6):541-544.
13. Kashyap N, Kumar P, Deshmukh B, Bhat S, Kumar A, Chauhan A *et al*. Association of ATP1A1 gene polymorphism with thermo tolerance in Tharparkar and Vrindavani cattle, *Veterinary World*. 2015; 8(7):892-897.
14. Baqir M, Bhusan S, Sharma D, Kumar A, Saminathan M, Dhama K *et al*. Bovine IL12RB1, IL12RB2, and IL23R Polymorphisms and Bovine Tuberculosis (bTB) Infection status. *Journal of Pure and Applied Microbiology*. 2014; 8(5):4117-4124.
15. Bhaladhare A, Chauhan A, Sonwane A, Kumar A, Kumar P, Kumar S *et al*. Association of Single Nucleotide Polymorphisms in IFNGR1 and IFNGR2 genes with bovine tuberculosis *Indian Journal of Animal Research*, 2019. DOI: 10.18805/ijar.B-3733
16. Baqir M, Bhushan B, Kumar S, Sonawane A, Singh R, Chauhan A *et al*. Association of polymorphisms in SLC11A1 gene with bovine tuberculosis trait among Indian cattle. *Journal of Applied Animal Research*. 2016; 44(1):380-383.
17. Bhaladhare A, Sharma, D, Chauhan A, Kumar A, Sonwane A, Singh RV *et al*. Association study of Single Nucleotide Polymorphisms (SNP) in Toll-like Receptor 9 gene with bovine tuberculosis. *Indian Journal of Animal Research*. 2018; 52(4):533-537.
18. Bhaladhare A, Sharma D, Kumar A, Sonwane A, Chauhan A, Singh R *et al*. Single nucleotide polymorphisms in toll-like receptor genes and case-control association studies with bovine tuberculosis. *Veterinary World*. 2016; 9(5):458-464.
19. Kumar S, Chauhan A, Baranwal A, Sonwane A, Kumar S, Singh RV. Host genetic resistance to mycobacterial

- infections in bovines. *Journal of Entomology and Zoology Studies*. 2018; 6(6):1102-1106.
20. Kumar S, Singh RV, Kumar S, Chauhan A, Kumar A, Bharati J *et al.* Association of Bovine CLEC7A gene polymorphism with host susceptibility to paratuberculosis disease in Indian cattle. *Research in Veterinary Science*. 2019; 123:216-222
 21. Kumar S, Kumar S, Singh R, Chauhan A, Agrawal S, Kumar A *et al.* Investigation of Genetic Association of Single Nucleotide Polymorphisms in SP110 Gene with Occurrence of Paratuberculosis Disease in Cattle. *International Journal of Livestock Research*. 2017; 7(3):81-88.
 22. Yadav R, Sharma AK, Singh R, Sonwane A, Kumar A, Chauhan A *et al.* An association study of SNPs with susceptibility to Bovine Paratuberculosis infection in cattle. *The Indian Journal of Animal Sciences*. 2014; 84(5):490-493.
 23. Kumar S, Kumar S, Singh RV, Chauhan A, Kumar A, Bharati J *et al.* Genetic association of polymorphisms in bovine TLR2 and TLR4 genes with Mycobacterium avium subspecies paratuberculosis infection in Indian cattle population. *Veterinary Research Communications*, 2019a. <https://doi.org/10.1007/s11259-019-09750-2>
 24. Prakash O, Kumar A, Sonwane A, Rathore R, Singh RV, Chauhan A *et al.* Polymorphism of cytokine and innate immunity genes associated with bovine brucellosis in cattle. *Molecular Biology Reports*. 2014; 41(5):2815-2825.
 25. Dige MS, Ahlawat SPS, Kumar P, Chauhan A, Inamdar B, Kokate LS *et al.* Dissecting the Partial Genomic Region of CXCR to Correlate with CMT in Vrindavani Cattle. *Indian Journal of Animal Research*. 2013; 47(4):335-339.
 26. Mishra C, Kumar S, Panigrahi M, Yathish HM, Chaudhary R, Chauhan A *et al.* Single Nucleotide Polymorphisms in 5' Upstream Region of Bovine TLR4 Gene Affecting Expression Profile and Transcription Factor Binding Sites. *Animal Biotechnology*. 2017; 29(2):119-128.
 27. Baranwal A, Sonwane A, Chauhan A, Panigrahi M, Sharma AK. Quantification and Comparison of TLR2 activity in monocyte-derived macrophages of zebu and crossbred cattle. *Iranian Journal of Veterinary Research*. 2018; 19(4):283-289.
 28. Chaudhary R, Kumar S, Yathish HM, Sivakumar A, Mishra C, Kumar A *et al.* Identification of SNPs in Beta 2 Microglobulin (b2M) Gene and their Association with IgG Concentration in Neonatal Buffalo Calves. *Journal of Pure and Applied Microbiology*. 2016; 10(2):1387-1394.
 29. Chaudhary R, Kumar S, Yathish HM, Mishra C, Chauhan A, Sahoo NR *et al.* Estimation of immunoglobulin G levels in colostrum of Murrah buffaloes. *International Journal of Science, Environment and Technology*. 2016; 5(4):2003-2007.
 30. Chaudhary R, Kumar S, Yathish HM, Sivakumar A, Mishra C, Kumar A *et al.* Nucleotide variability in Beta 2 Microglobulin (β 2M) gene and its association with colostral IgG levels in buffaloes (*Bubalus bubalis*). *Indian Journal of Animal Research*. 2018; 52(1):51-55.
 31. Galloway SM, McNatty KP, Cambridge LM, Laitinen MP, Juengel JL, Jokiranta T *et al.* Mutations in an oocyte-derived growth factor gene (BMP15) cause increased ovulation rate and infertility in a dosage-sensitive manner. *Nature Genetics*. 2000; 25(3):279.
 32. Hanrahan JP, Gregan SM, Mulsant P, Mullen M, Davis GH, Powell R *et al.* Mutations in the genes for oocyte-derived growth factors GDF9 and BMP15 are associated with both increased ovulation rate and sterility in Cambridge and Belclare sheep (*Ovis aries*). *Biology of Reproduction*. 2004; 70(4):900-909.
 33. Souza CJ, MacDougall C, Campbell BK, McNeilly AS, Baird DT. The Booroola (*FecB*) phenotype is associated with a mutation in the bone morphogenetic receptor type 1 B (BMPRIB) gene. *Journal of Endocrinology*. 2001; 169(2):1-6.
 34. Drouilhet L, Mansanet C, Sarry J, Tabet K, Bardou P, Woloszyn F *et al.* The highly prolific phenotype of Lacaune sheep is associated with an ectopic expression of the B4GALNT2 gene within the ovary. *PLoS Genetics*. 2013; 9(9):1003809.
 35. Turner HN. Origins of the CSIRO Booroola. In: CSIRO, Division of Animal Production, Genetics Research Laboratories, PO Box 184, North Ryde, NSW 2113, Australia, editor/s. *The Booroola Merino*. Proceedings of a workshop held at Armidale, NSW; 24; Melbourne, Vic: CSIRO, 1980-1982, 1-7.
 36. Turner HN, Young SS. *Quantitative genetics in sheep breeding*. South Melbourne, Vict: Macmillan Co. of Australia Pty Ltd. London, 1969, 332.
 37. Fabre S, Pierre A, Mulsant P, Bodin L, Di Pasquale E, Persani L *et al.* Regulation of ovulation rate in mammals: contribution of sheep genetic models. *Reproductive Biology and Endocrinology*. 2006; 4(1):20.
 38. Raftery LA, Sutherland DJ. TGF- β family signal transduction in *Drosophila* development: from Mad to Smads. *Developmental biology*. 1999; 210(2):251-268.
 39. Massagué J. How cells read TGF- β signals. *Nature reviews Molecular cell biology*. 2000; 1(3):169.
 40. Glistler C, Kemp CF, Knight PG. Bone morphogenetic protein (BMP) ligands and receptors in bovine ovarian follicle cells: actions of BMP -4, -6 and -7 on granulosa cells and differential modulation of Smad-1 phosphorylation by follistatin. *Reproduction*. 2004; 127(2):239-254.
 41. Ten Dijke P, Korchynskiy O, Valdimarsdottir G, Goumans MJ. Controlling cell fate by bone morphogenetic protein receptors. *Molecular and cellular endocrinology*. 2003; 211(1-2):105-113.
 42. Souza CJ, Gonzalez-Bulnes A, Campbell BK, McNeilly AS, Baird DT. Mechanisms of action of the principal prolific genes and their application to sheep production. *Reproduction, Fertility and Development*. 2004; 16(4):395-401.
 43. Fabre S, Pierre A, Pisselet C, Mulsant P, Lecerf F, Pohl J *et al.* The Booroola mutation in sheep is associated with an alteration of the bone morphogenetic protein receptor-IB functionality. *Journal of Endocrinology*. 2003; 177(3):435-444.
 44. Shackell GH, Hudson NL, Heath DA, Lun S, Shaw L, Condell L *et al.* Plasma gonadotropin concentrations and ovarian characteristics in Inverdale ewes that are heterozygous for a major gene (*FecX1*) on the X chromosome that influences ovulation rate. *Biology of reproduction*. 1993; 48(5):1150-1156.
 45. Nowak Z, Charon KM. Identification of fecundity gene (*FecB*) carriers using microsatellite markers and its effect on sheep weight. *Journal of applied genetics*. 2001;

- 42(1):49-57.
46. Kolte AP, Mishra AK, Kumar S, Arora AL, Singh VK. A study on effect of carrying *FecB* gene on body weight in Garole and Garole x Malpura sheep. Asian Australasian Journal of Animal Sciences. 2005; 18(10):1379.
 47. Visscher AH, Dijkstra M, Lord EA, Suss R, Rosler HJ, Heylen K *et al.* Maternal and lamb carrier effects of the Booroola gene on food intake, growth and carcass quality of male lambs. Animal Science. 2000; 71(2):209-217.
 48. Walling GA, Dodds KG, Galloway SM, Beattie AE, Lord EA, Lumsden JM *et al.* The consequences of carrying the Booroola fecundity (*FecB*) gene on sheep live weight. In Proceedings of the British Society of Animal Science, 2000, 43.
 49. Smith POWS, Hudson NL, Shaw L, Heath DA, Condell L *et al.* Effects of the Booroola gene (*FecB*) on body weight, ovarian development and hormone concentrations during fetal life. Journal of Reproduction and Fertility. 1993; 98(1):41.
 50. Guo X, Wang X, Di R, Liu Q, Hu W, He X *et al.* Metabolic Effects of *FecB* Gene on Follicular Fluid and Ovarian Vein Serum in Sheep (*Ovis aries*). International journal of molecular sciences. 2018; 19(2):539.
 51. Kumar D, Joshi A, Naqvi SM, Kumar S, Mishra AK, Maurya VP *et al.* Sperm motion characteristics of Garole x Malpura sheep evolved in a semi-arid tropical environment through introgression of *FecB* gene. Animal reproduction science. 2007; 100(1-2):51-60.
 52. Price CA, Hudson NL, McNatty KP. Effect of testosterone and bovine follicular fluid on concentrations of luteinizing hormone and follicle-stimulating hormone in plasma of castrated rams that are homozygous carriers or non-carriers of the Booroola fecundity gene. Journal of reproduction and fertility. 1992; 95(3):947-957.
 53. Liu SF, Jiang YL, Du LX. Studies of BMPR-IB and BMP15 as candidate genes for fecundity in Little Tailed Han sheep. Yi chuan xue bao= Acta genetica Sinica. 2003; 30(8):755-760.
 54. Wang G, Mao X, George DH, Zhao Z, Zhang L, Zeng Y. DNA tests in Hu sheep and Han sheep (small tail) showed the existence of Booroola (*FecB*) mutation. Journal of Nanjing Agricultural University. 2003; 26(1):104-106.
 55. Davis GH, Balakrishnan L, Ross IK, Wilson T, Galloway SM, Lumsden BM *et al.* Investigation of the Booroola (*FecB*) and Inverdale (*FecXI*) mutations in 21 prolific breeds and strains of sheep sampled in 13 countries. Animal Reproduction Science. 2006; 92(1-2):87-96.
 56. Guan F, Liu SR, Shi GQ, Yang LG. Polymorphism of *FecB* gene in nine sheep breeds or strains and its effects on litter size, lamb growth and development. Animal Reproduction Science. 2007; 99(1-2):44-52.
 57. Ranga PK, Singh RV. Genetic polymorphism of the Booroola fecundity (*FecB*) gene in Garole sheep. Use In Helen Newton Turner International Workshop on Using the Booroola (*FecB*) Gene in Sheep Breeding Programs. Pune, 2008, 221.
 58. Kumar S, Mishra AK, Kolte AP, Dash SK, Karim SA. Screening for Booroola (*FecB*) and Galway (*FecXG*) mutations in Indian sheep. Small Ruminant Research. 2008; 80(1-3):57-61.
 59. Sudhakar A, Rajendran R, Rahumathulla PS. Detection of Booroola (*FecB*) mutation in Indian sheep-Nilagiri. Small ruminant research. 2013; 113(1):55-57.
 60. Davis GH. The Booroola gene: origin, distribution, utilization and management of the *FecB* mutation. In Helen Newton Turner International Workshop on Using the Booroola (*FecB*) Gene in Sheep Breeding Programs. Pune, 2008, 22-31.