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Susmita Das
Department of Zoology,
University of Kalyani, Nadia,
West Bengal, India

Kausik Mondal
Associate Professor,
Department of Zoology,
University of Kalyani
Dist- Nadia, West Bengal,
India

Salma Haque
Department of Zoology,
University of Kalyani, Nadia,
West Bengal, India

Correspondence
Kausik Mondal
Associate Professor,
Department of Zoology,
University of Kalyani
Dist- Nadia, West Bengal,
India

A review on application of probiotic, prebiotic and synbiotic for sustainable development of aquaculture

Susmita Das, Kausik Mondal and Salma Haque

Abstract

Aquaculture is one of the fastest emerging food producing sectors of the world. World aquaculture has immensely grown during the last few years as well as becoming an economically significant zone. However in this culture, aquatic organisms are subjected to stress condition that weakens fish immune system as well as increases the susceptibility to diseases. Thus, affects both economic development and socio-economic status of the local people in many countries. Disease control in aquaculture industry has been achieved by following different methods using traditional ways, synthetic chemicals and antibiotics. Application of antimicrobial drugs and pesticides has steered to the evolution of resistant strains of bacteria. Therefore, alternative methods are much more essential to maintain a healthy microbial environment in the aquaculture systems. The present review summarizes and discusses the effects of probiotic, prebiotic or synbiotic administration on growth performance, stress tolerance, intestinal microbiota, immune response and health of aquatic organisms.

Keywords: Aquaculture, probiotic, prebiotic, synbiotic, aquaculture

1. Introduction

Aquaculture is one of the world's promising and fastest developing food-producing sectors with the largest potential to accomplish growing demand of aquatic food^[1]. World aquaculture has immensely grown during the last few years as well as becoming an economically significant zone. With the rising commercialization and intensification of aquaculture production, diseases and deterioration of environmental conditions are major problems in fish farming and face massive economic losses^[2]. For prevention and control of diseases, antibiotics used as traditional strategy during the last decades and also for fish growth as well as efficacy of feed conversion. However, the development of antimicrobial resistant pathogens were recognized but there is a huge risk of transmission of resistance bacteria from aquatic environment to human^[1]. As per them, using antibiotic is harmful for aquatic animals because it kills beneficial microbiota in gastrointestinal system of animals and it also accumulates in fish products to be unsafe for human consumption. Considering these factors, the improvements of non-antibiotic agents are more suitable for health management in this culture^[3]. Dietary supplement such as probiotic, prebiotic and synbiotic provide nonspecific disease protection and also act as growth promoting factors. Although various studies evaluating the effectiveness of probiotic, prebiotic and synbiotic in fish has received till present very little attention and available data are still insufficient. Therefore, the objective of this review is to compile the existing data on the use of probiotic, prebiotic and synbiotic in aquaculture, highlighting the most important properties demonstrated to date.

2. Probiotic

Probiotics shows a new dimension in disease resistance and improving water quality in aquaculture industry. The Greek word probiotic means "for life", was introduced by Parker^[4]. According to him, probiotics are "Organisms and substance, which contribute to intestinal and microbial balance". Fuller,^[5] defined probiotics as "live microbial feed supplement which beneficially affect the host animal by improving its intestinal microbial balance". Probiotics are used in aquaculture to improve growth performance^[6], nutrition^[7], decrease diseases^[8] and develop immune system^[9].

2.1 Micro Organisms of Probiotic

Now a days, Probiotics are commonly used as therapeutic, prophylactic and growth supplements in aquaculture as well as “functional food” for human health [1]. Probiotics include Gram-positive bacteria, Gram-negative bacteria and many other organisms like yeast, bacteriophages and unicellular algae [10]. Pandiyan *et al.*, [1] also added that Lactic Acid Bacteria have been broadly used as probiotic strains which are usually present in the intestine of healthy fishes such as the *Lactobacilli* and *Bifidobacteria*. Some gram positive bacteria like *Bacillus*, *Enterococcus*, *Streptococcus* act as common probiotic strains which are the main gastrointestinal microbiota [11]. On the other hand, Gram negative facultative anaerobes are dominant in gastrointestinal tract of fishes and shellfishes, as well as, symbiotic anaerobes prevail in posterior part of digestive tract of tropical herbivorous fishes [1].

2.2 Types of Probiotics

Probiotics are classified into following two groups:

2.2.1 Feed Probiotics

Some bacterial, fungal strains can be blended with feeding pellets or by encapsulating into live feed stock or administered orally to feed rearing animals to prevent disease and enhance essential microbial flora of the gut [11, 12]. Viability of strains should be tested before feeding animals. Probiotics like lactic acid bacteria applied in the feed of fry of Atlantic cod, showed adequate growth, survival and immune response [13].

2.2.2 Water probiotics

According to Prasad *et al.*, [11] water probiotics are applied to reduce organic pollutants and various contaminants in water by directly applying to rearing medium. These improve water quality by converting organic matter to smaller units. Breakdown of organic matters evolve simpler substances like glucose and amino acids that are used as food for beneficial bacteria which reduce the accumulation of organic pressure and provide congenial environment to farmed stock. Probiotic bacteria such as *Bacillus* sp. can convert organic matter to CO₂ so that organic effluent can be minimized in aquatic system. By using nitrifying bacteria, the quantity of nitrate, nitrite, ammonia are reduced to a large extent. These lead to purify the water in the hatchery enhancing larval survival and growth [14, 15].

2.3 Significance of Probiotics in Aquaculture

Probiotics use in aquaculture show great impact on aquatic organisms. Probiotics decrease accumulation of organic load and maintain water quality in an efficient way. A modern probiotic organism can easily fulfill the desires of sustainable aquaculture development because it can heighten two major key factors of growth performance and disease resistance [16]. Prasad *et al.*, [11] reported that *Lactic Acid Bacteria*, a popular probiotic strain, can be applied to control bacterial pathogen. In addition, another well-known probiotic organism, *Bacillus* sp. is used to diminish metabolic waste in aquatic system. Many strains of *Aeromonas* sp., *Pseudomonas* sp., *Vibrio* sp. act against infectious hematopoietic necrosis virus to show antiviral activity [17]. These probiotic organisms may be used singly or in combination such as incorporation of individual or combined supplementation of *Lactobacillus rhamnosus* and *Lactobacillus sporogenes* enhance health and disease resistance of common carp [18, 19, 20, 21]. Probiotics do not cause

water pollution because of their eco-friendly nature, thus more and more suitable for aquaculture system. They not only promote animal health but also maintain consumer health safety [11]. Uses of probiotics and their target aquatic organisms are briefly demonstrated in table 1.

There are some possible benefits linked to the administering of probiotics which have already been suggested as:

2.3.1 Improvement of Water Quality

According to Michael *et al.*, [22] the contamination of various nitrogenous compounds such as ammonia, nitrite and nitrate has been a serious problem in aquaculture system. In high concentrations, these compounds may be tremendously harmful and cause mass mortality. Ma *et al.*, [23] described that *Lactobacillus* sp. simultaneously eliminates nitrogen and pathogens from polluted shrimp farms. According to Stanier *et al.*, [24] gram-positive bacteria are usually more efficient in transforming organic matter back to CO₂ than gram-negative bacteria, which would convert organic carbon to bacterial biomass or slime. The aerobic gram-positive bacteria, such as *Bacillus* sp. were associated with development of water quality, reduction of pathogenic population in culture environment, enhancement of survival and growth rate, and the better health condition of juvenile *Penaeus monodon* [25,26].

2.3.2 Control of Diseases

Probiotics or their products have been found useful for health benefits and disease prevention in fish culture system. These include microbial adjunct that stop pathogens from multiplying in the gastrointestinal tract, on the superficial surfaces and in culture environment of the culture species [27]. Administration of probiotic can reduce the use of antibiotics and synthetic chemicals in the fish feed [5]. The effect of probiotic organisms is achieved through improving the immune system of culture organism, enhancing their disease resistance potential or generating inhibitory-substance that prevent the pathogenic organisms from disease formation in the host [16]. As per Rengpipat *et al.*, [28] the use of *Bacillus* sp. showed disease protection by initiating both cellular and humoral immune resistances in tiger shrimp (*P. monodon*).

2.3.3 Growth Promoters

Probiotics have been used in aquaculture to enhance the growth of cultivated species and yet no side effect on the host. Yassir *et al.*, [29] used probiotic bacteria as growth promoter on tilapia (*Oreochromis niloticus*) in his work and the highest growth performance with *Micrococcus luteus* was noted and the best feed conversion ratio was seen with the same probiotic organism. So they mentioned *M. luteus* as a growth promoters in fish culture. Lactic acid bacteria also referred as growth promoters due to effect on the growth rate in juvenile carp [30].

2.3.4 Source of Nutrients and Enzymatic Contribution to Digestion

According to Michael *et al.*, [22] various microorganisms have a valuable effect in the digestive system of aquatic organisms. It has also been stated that *Bacteroides* and *Clostridium* sp. have supplied nutrients like fatty acids and vitamins to the host in fish aquaculture. Some microorganisms such as *Agrobacterium* sp., *Pseudomonas* sp., *Microbacterium* sp. and *Staphylococcus* sp. may contribute to nutritional processes in Arctic charr (*Salvelinus alpinus*) [31].

2.3.5 Improvement of the Immune Response

As per Michael *et al.*,^[22] among the various beneficial effects of probiotics, modulation of immune system is one of the most important benefits of probiotics. They also reported that fish larvae, shrimps and other invertebrates have immune systems that are dependent mainly on non-specific immune responses for their resistance to infection. Sakai *et al.*,^[32] has been demonstrated that oral administration of *Clostridium butyricum* bacteria to rainbow trout improved their resistance to vibriosis by developing the phagocytosis of leucocytes.

2.3.6 In Aquaculture Management

These beneficial probiotic organisms can be administered through feeding, injection or immersion to improve aquaculture management^[22].

3. Prebiotic

Prebiotics are non-digestible food ingredient that stimulate the growth or activity of beneficial gut commensal bacteria in host thus improves host health^[33]. Gibson *et al.*,^[34] reported that a food ingredient which acts as prebiotic must possess the following criteria such as showing resistance to gastric acidity, hydrolysis by digestive enzyme, fermentation by gastrointestinal microflora and increase the abundance of intestinal bacteria related to health.

3.1 Prebiotic Organisms

Nowadays, carbohydrates use as most efficient prebiotics which can be classified on the basis of molecular size or step of polymerization^[35]. According to Mahious and Ollevier^[36], Fooks *et al.*,^[37] and Gibson *et al.*,^[34] various food substances such as non-digestible carbohydrates, some proteins and peptides, certain lipids act as prebiotic ingredient. Prebiotic compounds like fructooligosaccharides, mannanoligosaccharides, inulin or B-glucanare considered as the most effective prebiotics in aquaculture^[33] (Table 2). Prebiotics are mainly fermented by *Bifidobacteria*, *Lactobacillus* and *Bacteriodes*^[38].

3.2 Significance of Prebiotics in Aquaculture

Prebiotics are essential dietary supplement which enhance growth performance as well as microbial activities of digestive tract, also boost immune system and improve stress resistance that are discussed below.

3.2.1 Use of Prebiotics on Growth Parameters

A huge number of prebiotics are used as feed supplement to achieve better growth performance. Growth parameters vary on the basis of aquatic organisms as well as prebiotic supplementation. A diet containing 20 g kg⁻¹ oligofructose, a fructooligosaccharide produced by partial enzymatic hydrolysis of inulin, resulted in increased growth of turbot larvae, but 20 g kg⁻¹ inulin itself had no effect on growth^[39].

3.2.2 Application of Prebiotics on Immune System

As per Yousefian and Amiri^[38], last few decades, antibiotics were used to control bacterial diseases but this type of chemical substance is advisable to avoid in aquaculture. In recent years in the aquaculture industry, alternative strategies have been developed for disease control as well as reduction in the widespread use of antibiotics. Prebiotic is a well-known group of these strategies which enhances non-specific immune response. According to Bailey *et al.*,^[40] prebiotic can modify microbes of GI tract by increasing immune responses.

3.2.3 Application of Prebiotics on Microbes of Gastrointestinal Track

The gastrointestinal track of all invertebrates and vertebrates plays a vital role for providing habitat to different kinds of microorganisms^[41]. Various prebiotic oligosaccharides such as inulin and oligofructose are fermented in the colon where they stimulate the growth of bacterial populations related with a well-functioning colon and this stimulation occurs because oligosaccharides are readily fermented by beneficial colonic bacteria and are not used effectively by pathogenic bacterial species^[38].

4. Synbiotics

Synbiotic defines as nutritional supplements combining probiotics and prebiotics in the form of synergism therefore improving beneficial effects of individual probiotics^[42]. According to Gibson and Roberfroid^[43] synbiotics was stated to 'characterise some colonic foods with interesting nutritional properties that make these compounds candidates for classification as health-enhancing functional ingredients'. Synbiotic positively affects the host by improving the survival and inserting of live microbial dietary supplements in the digestive tract by selectively stimulating the growth and/or by triggering the metabolism of one or a limited number of health-promoting bacteria, hence promoting the host "welfare"^[42].

4.1 Significance of Synbiotics in Aquaculture

Synbiotics can be applied through supplementation or external bathing to develop growth performance, proper feed utilization, disease resistance, digestibility and stimulation of the immune system of aquatic organisms^[44]. Application of synbiotics on different aquatic organisms in aquaculture system are given in table 3.

4.1.1 Application of Synbiotics on Fish Survival Rate, Growth Parameters and Feed Utilization

In rainbow trout, administration of *Enterococcus faecalis* and mannanoligosaccharides/polyhydroxybutyrate acid for 12 weeks not affects the survival rate of fish as well as the experimental fish was in good condition and there was no mortality during the feeding trial^[42]. According to Ai *et al.*,^[45] and Geng *et al.*,^[46] administration of *Bacillus subtilis*/fructooligosaccharides in yellow croaker or *Bacillus subtilis*/chitosan in cobia, did not affect the survival rate, showing no changes among different dietary treatments. In case of Japanese flounder nourishing with *Bacillus clausii* and mannanoligosaccharides/fructooligosaccharides, in which fish retained active ingestion, showed proper growth and survival for all time^[47].

In case of growth parameters, almost all references stated a positive effect of synbiotics application. In case of rainbow trout, dietary mannanoligosaccharides combined with *Enterococcus faecalis* significantly enhanced growth performance and nutrient utilization of fish, on the other hand, combination of *Enterococcus faecalis* and polyhydroxybutyrate acid developed growth performance of rainbow trout to some extent^[42].

4.1.2 Uses of Synbiotics on Fish Digestive Enzyme Activity

Administration of synbiotics may effect on enzymes of fish digestive system. The increase in digestive enzyme activities would permit the host degrades more nutrients, enhancing digestion and promoting a probable increase in the weight gain rate and/or feed efficiency^[42].

4.1.3 Uses of Synbiotics on Fish Immune System

As per Cerezeula *et al.*, [42] synbiotics significantly effect on fish immune system considering a number of activities such as lysozyme activity, phagocytic activity, alternative complement pathway, respiratory burst, mucus production and superoxide dismutase activity. Lysozyme is the vital bactericidal enzymes of innate immunity, and shows acrucialdefense mechanism against various pathogens in fish [48].

4.1.4 Application of Synbiotics on Fish Disease Resistance

The efficiency of synbiotic therapy in terms of protection against infectious agents could be assessed by a challenge test because of its controlling power on pathogens and disease resistance capability [42].

Table 1: Uses of Probiotic in aquaculture system (Cruz *et al.* [49])

| Uses of Probiotic | Probiotic Species | Gram Positive/negative Bacteria | Target aquatic species | Reference |
|--------------------------------|-----------------------------------|---------------------------------|----------------------------------|-----------|
| Water quality | <i>Bacillus</i> sp. | +ve | <i>Penaeus monodon</i> | [50] |
| | <i>Vibrio</i> sp. NE 17 | -ve | <i>Macrobrachium rosenbergii</i> | [51] |
| | <i>Lactobacillus acidophilus</i> | +ve | <i>Clarias gariepinus</i> | [52] |
| Control of diseases | <i>Enterococcus faecium</i> SF 68 | +ve | <i>Anguilla Anguilla</i> | [53] |
| | <i>Pseudomonas fluorescens</i> | -ve | <i>Oncorhynchus mykiss</i> | [54] |
| | <i>Lactococcus lactis</i> | +ve | <i>Epinephelus coioides</i> | [55] |
| | <i>Pseudomonas</i> sp. | -ve | <i>Oncorhynchus mykiss</i> | [56] |
| | <i>Bacillus</i> sp. | +ve | Penaeids | [14] |
| | <i>Vibrio alginolyticus</i> | -ve | Salmonids | [57] |
| Growth promoter | <i>Lactobacillus lactis</i> AR21 | +ve | <i>Brachionus plicatilis</i> | [58] |
| | <i>Bacillus</i> sp. | +ve | Catfish | [59] |
| | <i>Streptococcus thermophiles</i> | +ve | <i>Scophthalmus maximus</i> | [60] |
| | <i>Bacillus coagulans</i> | +ve | <i>Cyprinus carpio koi</i> | [61] |
| | <i>Bacillus</i> NL 110 | +ve | <i>M. rosenbergii</i> | [62] |
| Digestion | <i>Lactobacillus acidophilus</i> | +ve | <i>Clarias gariepinus</i> | [52] |
| | <i>Vibrio</i> NE 17 | +ve | <i>M. rosenbergii</i> | [51] |
| | <i>Lactobacillus helveticus</i> | +ve | <i>Scophthalmus maximus</i> | [60] |
| Improvement of immune response | <i>Clostridium butyricum</i> | +ve | Rainbow trout | [32] |
| | <i>L. casei</i> | +ve | <i>Poecilopsis gracilis</i> | [63] |
| | <i>L. acidophilus</i> | +ve | <i>Paralichthys olivaceus</i> | [64] |

Table 2: Different Prebiotic substances in aquaculture (Song *et al.* [65])

| Prebiotic substances | Subtype | Aquatic Organisms | Reference |
|----------------------|---|-------------------------------|-----------|
| Oligosaccharide | Fructooligosaccharides (FOS) Mannanoligosaccharide (MOS) Galactooligosaccharide (GOS) Arabinoxylan-oligosaccharide | <i>Salmo salar</i> L. | [66] |
| | | <i>Megalobrama terminalis</i> | [67] |
| | | <i>Paralichthys olivaceus</i> | [47] |
| | | <i>Acipenser stellatus</i> | [68] |
| | | <i>Oreochromis niloticus</i> | [69] |
| | | <i>Dicentrarchus labrax</i> | [70] |
| | | <i>Panulirus ornatus</i> | [71] |
| | | <i>Sciaenops ocellatus</i> | [72] |
| | | Atlantic Salmon | [66] |
| | | Siberian sturgeon | [73] |
| Polyoligosaccharide | Inulin | Nile tilapia | [74] |
| | | <i>Huso huso</i> | [75] |
| | | <i>Pseudoplatystoma</i> sp. | [76] |

Table 3: Application of synbiotic on different aquatic organisms (Cerezeula *et al.* [42])

| Synbiotic (Probiotic/Prebiotic) | Aquatic Organisms | Reference |
|--|-------------------------------|-----------|
| <i>Enterococcus faecalis</i> /MOS, PHB | <i>Oncorhynchus mykiss</i> | [77] |
| <i>Bacillus clausii</i> /MOS, FOS | <i>Paralichthys olivaceus</i> | [47] |
| <i>Bacillus subtilis</i> / Chitosan | <i>Rachycentron canadum</i> | [46] |
| <i>Bacillus subtilis</i> / FOS | <i>Larimichthys crocea</i> | [45] |

[MOS-Manna oligosaccharide, FOS- fructooligosaccharide, PHB- polyhydroxybutyrate acid]

5. Constraints in the use of probiotic, prebiotic and synbiotic:

The use of probiotic, prebiotic and synbiotic receive excessive attention as a beneficial approach in aquaculture system but sometime due to insufficient information on their modes of action, the results of these application were hampered [78]. In human, probiotics may be responsible for four types of side effects in susceptible individuals: systemic infections, deleterious metabolic activities, excessive immune

stimulation and gene transfer. There are few reports of bacteremia in humans, where separation of probiotic bacteria from infections seems to be the consequence of an opportunistic infection caused by skin lesions, cancer, chronic illness, or a drug induced abnormality. These conditions lead to a decreased intestinal wall that promotes the route of the bacteria through the mucosal epithelium and ultimately that may progress to septicemia [49]. Though, no evidence has been found in the fishes. According to Verschuere *et al.*, [79] the use

of probiotics as biological control agents should be treated as a kind of risk insurance that may not provide any remarkable advantage when the culture is executed under optimal conditions and in the absence of pathogens. When culturing the aquatic species, microbial populations in the intestinal contents are much higher than those in the surrounding water^[80]. Since, there is a chance of transmission of resistant bacteria from aquaculture environments to humans. The probiotics are destroyed by any other chemical or drug which generally interferes with the establishment of useful microbes^[78]. Injecting probiotic into aquatic animals has been used to stimulate fish immune response against bacterial pathogenic infection^[81, 82]. However, it is tough to inoculate probiotics into cultured fish, especially into small animals, and to treat huge numbers of fish in this way^[83]. According to Hoseinifar *et al.*,^[84] the administration of *Lactobacillus lactis* on haematological parameters of *Acipenser persicus* demonstrated that the number of blood lymphocytes decreased whereas both the granulocytes increased. Evidences on the effect of prebiotic and synbiotics on fish immunomodulation is limited. In case of beluga juveniles (*Huso huso*), prebiotic like dietary oligofructose didn't show any significant effect on serum total protein, serum glucose, RBC counts, mean cell haemoglobin concentration, mean corpuscular volume or different enzyme activities like alkaline phosphatase, serum lactate dehydrogenase, alanine aminotransferase and aspartate aminotransferase^[85]. The effects of fructooligosaccharide on haemato-immunological parameters of stellate sturgeon juvenile were studied by Akrami *et al.*,^[68] after 11 weeks of administration, they revealed that haemato-immunological parameters (RBC, WBC, haemoglobin, haematocrit and lymphocyte levels etc.) were significantly developed, while respiratory burst activity of leucocytes was not significantly improved by the dietary supplementations. Very limited studies have been conducted on constraints in the use of probiotic, prebiotic and synbiotic in aquaculture systems.

6. Conclusion

In recent years, probiotics, prebiotics as well as synbiotics have become an essential parts of the aquaculture practices for improving the growth performance and disease resistance. Probiotics play an important role in feed conversion, growth rates, weight gain, immune response and disease resistance of fish. On the other hand, prebiotics also have various beneficial effects mainly in disease resistance and nutrient availability of fish. Synbiotics, the combined application of probiotics and prebiotics, which improve the survival and establishment of the live microbial dietary supplement in the gastrointestinal tract of the host. The use of synbiotics confirm greater benefits than the application of individual probiotics. Administration of a commercial synbiotics show an increase in growth performances, survival rate, serum protein and albumin content and enhanced feeding efficiency. These biotic applications are essential for developing the health status and production of fish, in place of more traditional immunological and disease control methods such as vaccinations, antimicrobials and immunostimulants. The significant variation in growth, feed utilization as well as health benefits with the dietary use of these biotic is possibly dependent on the fish species, duration of feeding and supplement dose.

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8. References

- Pandiyar P, Balaraman D, Thirunavukkarasu R, George EGJ, Subaramaniyam K, Manikkam S *et al.* Probiotics in aquaculture. *Drug Invent Today*. 2013; 5:55-59.
- Bondad-Reantaso MG, Subasinghe RP, Arthur JR. Disease and health management in Asian aquaculture. *Vet Parasitol*. 2005; 132:249-272.
- Denev SA. Ecological alternatives of antibiotic growth promoters in animal husbandry and aquaculture. DSc. Thesis. Starajagora, Bulgaria. Department of Biochemistry Microbiology, Trakia University, 2008; 294.
- Parker RB. Probiotics:the other half of the antibiotics story. *Anim Nutr Health*. 1974; 29:4-8.
- Fuller R. Probiotic in man and animals. *J Appl Bacteriol*. 1989; 66:365-378.
- Silva EF, Soares MA, Calazans NF, Vogeley JL, doValle BC, Soares R *et al.* Effect of probiotic (*Bacillus* spp.) addition during larvae and post larvae culture of white shrimp *Litopenaeus vannamei*. *Aquac Res*. 2013; 44:13-21.
- Zhou X, Wang Y, Li W. Effect of probiotic the larvae shrimp (*Penaeus vannamei*) based on water quality, survival rate and digestive enzyme activities. *Aquacult*. 2009; 287:349-353.
- Irianto A, Austin B. Use of probiotics to control furunculosis in rainbow trout, *Oncorhynchus mykiss* (Walbaum). *J Fish Dis*. 2002; 25:1-10.
- Nayak SK, Swain P, Mukherjee SC. Effect of dietary supplementation and vitamin c on the immune response of Indian major carp, *Labeo rohita* (Ham.). *Fish Shellfish Immunol*. 2007; 23:892-896.
- Mukherjee A, Dutta D, Banerjee S, Ringo E, Breines EM, Hareide E *et al.* Potential probiotics from Indian major carp, *Cirrhinus mrigala*. Characterization, pathogen inhibitory activity, partial characterization of bacteriocin and production of coenzymes. *Res Vet Sci*. 2016; 108:76-84.
- Prasad L, Baghel DS, Kumar V. Role and prospects of probiotics use in aquaculture. *Aquacult*. 2003; 4(2):247-251.
- Nageswara PV, Babu DE. Probiotics as an alternative therapy to minimize or avoid antibiotics use in aquaculture. *Fishing Chimes*. 2006; 26(1):112-114.
- Gildberg A, Mikkelsen H, Sandaker E, Ringo E. Probiotic effect of lactic acid bacteria in the feed on growth and survival of fry of Atlantic cod (*Gadus morhua*). *Hydrobiologia*. 1997; 352:279-285.
- Moriarty DJW. Control of luminous vibrio species in penaeid aquaculture ponds. *Aquacult*. 1998; 164(1-4):351-358.
- Lipton AP. Disease management in shrimp culture with species reference to probiotics and additives workshop on National Aquaculture Week. January-February 1997. Aquaculture Foundation of India Chennai. 1998, 20-219.
- Dawood MAO, Koshio S. Recent advances in the role of probiotics and prebiotics in carp aquaculture: a review. *Aquacult*. 2016; 454:243-251.
- Kamei Y, Yoshimizu M, Ezura Y, Kimura T. Screening of bacteria with antibacterial activity from fresh water salmonid hatcheries. *Microbiol Immunol*. 1988; 32:67-73.
- Allameh SK, Yusoff FM, Ringo E, Daud HM, Saad CR, Ideris A. Effects of dietary mono and multi probiotic strains on growth performance, gut bacteria and body

- composition of Javanese carp (*Puntius gonionotus*, Bleeker 1850). *Aquac Nutr.* 2014. <http://dx.doi.org/10.1111/anu.12265>.
19. Chi C, Jiang B, Yu XB, Liu TQ, Xia L, Wang GX *et al.* Effects of three strains of intestinal autochthonous bacteria and their extracellular products on the immune response and disease resistance of common carp, *Cyprinus carpio*. *Fish Shellfish Immunol.* 2014; 36:9-18.
 20. Faramazi M, Kiaalvandi S, Lashkarbolooki M, Iranshahi F. The investigation of *Lactobacillus acidophilus* as probiotics on growth performance and disease resistance of rainbow trout (*Oncorhynchus mykiss*). *Am-Eurasian J Sci Res* 2011; 6(1): 32-38.
 21. Harikrishnan R, Balasundaram C, Heo MS. Potential use of probiotic- and triherbal extract-enriched diets to control *Aeromonas hydrophila* infection in carp. *Dis Aquat Org.* 2010; 92:41-49.
 22. Michael ET, Amos SO, Hussaini LT. A review on probiotics application in aquaculture. *Fish Aquac J.* 2014; 5:4.
 23. Ma GW, Cho YS, Oh KH. Removal of pathogenic bacteria and nitrogen by *Lactobacillus* spp. JK-8- JK-11. *Aquacult.* 2009; 287:266-270.
 24. Stanier RY, Doudoroff M, Adelberg EA. The microbial world. Prentice- Hall Inc., Englewood Cliffs. N.J. 1963; 153-156.
 25. Dalmin G, Kathiresan K, Purushothaman A. Effect of probiotics on bacterial population and health status of shrimp in culture pond ecosystem. *Indian J Exp Biol.* 2001; 39:939-942.
 26. Ngan PTT, Phu TQ. Effects of *Bacillus* bacteria (B8, B37, B38) on water quality of black tiger shrimp (*Penaeus monodon*) cultured tanks. Proceedings of the 4th aquaculture and fisheries conference, 2011; 28-41.
 27. Verschuere, Rombaut G, Sorgeloos P, Verstraete W. Probiotic bacteria as biological control agents in aquaculture. *Microbiol Mol Biol Rev.* 2000; 64:655-671.
 28. Rengpipat S, Rukpratanporn S, Priyatitivorakul S, Menasaveta P. Immunity enhancement in black tiger shrimp (*Penaeus monodon*) by probiotic bacterium (*Bacillus* S11). *Aquacult.* 2000; 191:271-288.
 29. Yassir AL, Adel ME, Azze A. Use of probiotic bacteria as growth promoters, antibacterial and the effect on physiological parameters of *Oreochromis niloticus*. *J Fish Dis.* 2002; 25: 633-642.
 30. Noh SH, Han K, Won TH, Choi YJ. Effect of antibiotics, enzymes, yeast culture and probiotics on the growth performance of Israeli carp. *Korean J Animal Sci.* 1994; 36:480-486.
 31. Ringø E, Strøm E, Tabachek JA. Intestinal microflora of salmonids: a review. *Aquac Res.* 1995; 26:773-789.
 32. Sakai M, Yoshida T, Atsuta S, Kobayashi M. Enhancement of resistance to vibriosis in rainbow trout, *Oncorhynchus mykiss* (Walbaum), by oral administration of *Clostridium butyricum* bacterin. *J Fish Dis.* 1995; 18:187-190.
 33. Guerreiro I, Couto A, Machado M, Castro C, Pousao-Ferreira P, Oliva-Teles A *et al.* Probiotics effect on immune and hepatic oxidative status and gut morphology of white sea bream (*Diplodus sargus*). *Fish Shellfish Immunol.* 2016; 50:168-174.
 34. Gibson GR, Probert HM, Van Loo J, Rastall RA, Roberfroid MB. Dietary modulation of the human colonic microbiota: Updating the concept of prebiotics. *Nutr Res Rev.* 2004; 17:259-275.
 35. Akhter N, Wu B, Memon AM, Mohsin M. Probiotics and prebiotics associated in aquaculture: a review. *Fish Shellfish Immunol.* 2015; 45:733-741.
 36. Mahious AS, Ollevier F. Probiotics and Prebiotics in Aquaculture. 1st Regional Workshop on Techniques for Enrichment of Live Food for Use in Larviculture-2005, AAARC, Urmia, Iran. 2005; p. 67.
 37. Fooks LJ, Fuller R, Gibson GR. Prebiotics, probiotics and human gut microbiology. *Int Dairy J.* 1999; 9:53-61.
 38. Yousefianl M, Amiri MS. A review of the use of prebiotic in aquaculture for fish and shrimp. *Afr J Biotechnol.* 2009; 8(25):7313-7318.
 39. Mahious AS, Gatesoupe FJ, Hervi M, Metailler R, Ollevier F. Effect of dietary inulin and oligosaccharides as prebiotics for weaning turbot, *Psetta maxima* (Linnaeus, C.1758). *Aquac Int.* 2006; 14(3):219-229.
 40. Bailey J, Blankenship L, Cox N. Effect of fructooligosaccharide on *Salmonella* colonization of the chicken intestine. *Poult Sci.* 1991; 70:2433-2438.
 41. Flickinger EA, Van Loo J, Fahey GC. Nutritional responses to the presence of inulin and oligofructose in the diets of domesticated animals: a review. *Crit Rev Food Sci Nutr.* 2003; 43:19-60.
 42. Cerezuela R, Meseguer J, Esteban MA. Current knowledge in Synbiotic use for fish aquaculture: a review. *J Aquac Res Development.* 2011; 1:1-7.
 43. Gibson GR, Roberfroid MB. Dietary modulation of the human colonic microbiota: introducing the concept of prebiotics. *J Nutr.* 1995; 125:1401-1412.
 44. Dehaghani PG, Baboli MJ, Moghadam AT, Ziaei-Nejad S, Pourfarhadi M. Effect of synbiotic dietary supplementation on survival, growth performance and digestive enzyme activities on common carp (*Cyprinus carpio*) fingerlings. *Czech J Anim Sci.* 2015; 60:224-232.
 45. Ai Q, Xu H, Mai K, Xu W, Wang J. Effects of dietary supplementation of *Bacillus subtilis* and fructooligosaccharide on growth performance, survival, non-specific immune response and disease resistance of juvenile large yellow croaker, *Larimichthys crocea*. *Aquacult.* 2011; 317:155-161.
 46. Geng X, Dong XH, Tan BP, Yang QH, Chi SY. Effects of dietary chitosan and *Bacillus subtilis* on the growth performance, non-specific immunity and disease resistance of cobia, *Rachycentron canadum*. *Fish Shellfish Immunol.* 2011; 31:400-406.
 47. Ye JD, Wang K, Li FD, Sun YZ. Single or combined effects of fructo- and mannan oligosaccharide supplements and *Bacillus clausii* on the growth, feed utilization, body composition, digestive enzyme activity, innate immune response and lipid metabolism of the Japanese flounder *Paralichthys olivaceus*. *Aquac Nutr.* 2011; 17:902-911.
 48. Lindsay GJH. The significance of chitinolytic enzymes and lysozyme in rainbow trout (*Salmo gairdneri*) defence. *Aquacult.* 1986; 51:169-173.
 49. Cruz PM, Ibanez AL, Hermosillo OAM, Saad HCR. Use of Probiotic in Aquaculture. *ISRN Microbiol.* 2012; 1-13. doi:10.5402/2012/916845.
 50. Shishehchian F, Yusoff FM, Shariff M. The effects of commercial bacterial products on macrobenthos community in shrimp culture ponds. *Aquac Int.* 2001; 9(5):429-436.
 51. Rahiman M, Yousuf J, Ambat T, Hatha M. Probiotic effect of *Bacillus* NL110 and *Vibrio* NE17 on the survival, growth performance and immune response of

- Macrobrachium rosenbergii* (de Man). Aquacult Res. 2010; 41(9):e120-e134.
52. Dohail A, Abdullah M, Roshada H, Aliyu M. Effects of the probiotic, *Lactobacillus acidophilus*, on the growth performance, haematology parameters and immunoglobulin concentration in African Catfish (*Clarias gariepinus*, Burchell 1822) fingerling. Aquacult Res. 2009; 40(14):1642-1652.
 53. Chang CI, Liu WY. An evaluation of two probiotic bacterial strains, *Enterococcus faecium* SF68 and *Bacillus toyoi*, for reducing edwardsiellosis in cultured European eel, *Anguilla anguilla* L. J Fish Dis. 2002; 25(5):311-315.
 54. Gram L, Melchiorson J, Spanggaard B, Huber I, Nielsen TF. Inhibition of *Vibrio anguillarum* by *Pseudomonas fluorescens* AH2, a possible probiotic treatment of fish. Appl Environ Microbiol. 1999; 65(3):969-973.
 55. Zhang S, Sing Y, Long M, Wei Z. Does dietary administration of *Lactococcus lactis* modulate the gut microbiota of grouper, *Epinephelus coioides*. J World Aquacult Soc. 2012; 43(2):198-207.
 56. Spanggaard B, Huber I, Nielsen J. The probiotic potential against vibriosis of the indigenous microflora of rainbow trout. Environ Microbiol. 2001; 3(12):755-765.
 57. Austin B, Stuckey LF, Robertson PAW, Effendi I, Griffith DRW. A probiotic strain of *Vibrio alginolyticus* effective in reducing diseases caused by *Aeromonas salmonicida*, *Vibrio anguillarum* and *Vibrio ordalii*. J Fish Dis. 1995; 18(1):93-96.
 58. Harzeveli ARS, VanDuffel H, Dhert P, Swing J, Sorgeloos P. Use of a potential probiotic *Lactococcus lactis* AR21 strain for the enhancement of growth in the rotifer *Brachionus plicatilis* (Muller). Aquacult Res. 1998; 29(6):411-417.
 59. Queiroz JF, Boyd CE. Effects of a bacterial inoculum in channel catfish ponds. J World Aquacult Soc. 1998; 29(1):67-73.
 60. Gatesoupe FJ. The use of probiotics in aquaculture. Aquaculture. 1999; 180(1-2):147-165.
 61. Lin Sh, Guan Y, Luo L, Pan Y. Effects of dietary chitosan oligosaccharides and *Bacillus coagulans* on growth, innate immunity and resistance of koi (*Cyprinus carpio koi*). Aquacult. 2012; 342-343:36-41.
 62. Rahiman M, Yousuf J, Ambat T, Hatha M. Probiotic effect of *Bacillus* NL110 and *Vibrio* NE17 on the survival, growth performance and immune response of *Macrobrachium rosenbergii* (de Man). Aquacult Res. 2010; 41(9):e120-e134.
 63. Hernandez LHH, Barrera TC, Mejia JC. Effects of the commercial probiotic *Lactobacillus casei* on the growth, protein content of skin mucus and stress resistance of juveniles of the Porthole livebearer *Poecilopsis gracilis* (Poecilidae). Aquacult Nutr. 2010; 16(4):407-411.
 64. Taoka Y, Maeda, Jo JY. Growth, stress tolerance and non-specific immune response of Japanese flounder *Paralichthys olivaceus* to probiotics in a closed recirculating system. Fisheries Sci. 2006; 72(2):310-321.
 65. Song SK, Beck BR, Kim D, Park J, Kim J, Kim HD *et al*. Probiotics as immunostimulants in aquaculture: A review. Fish Shellfish Immunol. 2014; 40:40-48.
 66. Grisdale-Helland B, Helland SJ, Gatlin III DM. The effects of dietary supplementation with Mannan oligosaccharide, fructooligosaccharide or galactooligosaccharide on the growth and feed utilization of Atlantic salmon (*Salmo salar* L.). Aquacult. 2008; 283:163-7.
 67. Zhang CN, Li XF, Xu WN, Jiang GZ, Lu KI, Wang LN *et al*. Combined effects of fructooligosaccharide and *Bacillus licheniformis* on innate immunity, antioxidant capability and disease resistance triangular bream (*Megalobrama terminalis*). Fish Shellfish Immunol. 2012; 32(2):316-21.
 68. Akrami R, Iri Y, Rostamy HK, Razeghi Mansour M. Effect of dietary supplementation of fructooligosaccharide (FOS) on growth performance, survival, lactobacillus bacteria population and hemato-immunological parameters of stellate sturgeon (*Acipenser stellatus*) juvenile. Fish Shellfish Immunol. 2013; 35:1235-9.
 69. Samrongpan C, Areechon N, Yoonpundh R, Sirsapoome P. Effect of mannan oligosaccharide on growth, survival and disease resistance of Nile tilapia (*Oreochromis niloticus* Linnaeus) fry. In: 8th international symposium on tilapia in aquaculture; Cairo, Egypt. 2008; 345-353.
 70. Torrecillas S, Makol A, Benitez-Santana T, Caballero MJ, Montero J, Sweetman J *et al*. Reduced gut bacterial translocation in European sea bass (*Dicentrarchus labrax*) fed Mannan oligosaccharides (MOS). Fish Shellfish Immunol. 2011; 30(2):674-681.
 71. Sang HM, Fotedar R. Effects of mannan oligosaccharide dietary supplementation on performances of the tropical spiny lobsters juvenile (*Panulirus ornatus*, Fabricius 1798). Fish Shellfish Immunol. 2010; 28(3):483-489.
 72. Zhou QC, Buentello JA, Galtin III DM. Effects of dietary prebiotics on growth performance, immune response and intestinal morphology of red drum (*Sciaenops ocellatus*). Aquacult. 2010; 309:253-257.
 73. Geralylou Z, Souffreau C, Rurangwa E, D'Hondt S, Calleweert L, Caurtin CM *et al*. Effect of arabinoxylan-oligosaccharide (AXOS) on juvenile Siberian sturgeon (*Acipenser baerii*) performance, immune responses and gastrointestinal microbial community. Fish Shellfish Immunol. 2012; 33:718-724.
 74. Ibrahim MD, Fathi M, Meslhy M, El-Aty AMA. Effect of dietary supplementation of inulin and vitamin C on the growth hematology, innate immunity and resistance of Nile tilapia (*Oreochromis niloticus*). Fish Shellfish Immunol. 2010; 29:241-246.
 75. Ahmdifar E, Akrami R, Ghelichi A, Zarejbad AM. Effects of different prebiotic inulin level on blood serum enzymes, hematologic and biochemical parameters of great sturgeon (*Huso huso*) juveniles. Comp Clin Path. 2011; 20:447-451.
 76. Mpurino JLP, Do Nascimento Vieira F, Jatoba AB, Da Silva BC, Jesus GFA, Seiffert WQ *et al*. Effect of dietary supplementation of inulin and W. cibaria on hemato-immunological parameters of hybrid surubim (*Pseudoplatystoma* sp). Aquacult Nutr. 2012; 18:73-80.
 77. Rodriguez-Estrada U, Satoh S, Haga Y, Fushimi H, Sweetman J. Effects of single and combined supplementation of *Enterococcus faecalis*, mannan oligosaccharide and polyhydroxybutyric acid on growth performance and immune response of rainbow trout *Oncorhynchus mykiss*. Aquacult Sci. 2009; 57:609-617.
 78. Raja S, Nandhini E, Sahana K, Dhanakkodi B. Beneficial and destructive effects of probiotics in aquaculture systems-A review. Int J Fish Aquat Stud. 2015; 2(3):153-159.
 79. Verschuere L, Rombaut G, Sorgeloos P, Verstraete W.

- Probiotic bacteria as biological control agents in aquaculture. *Microbiol Mol Biol Rev.* 2000; 64(4):655-671.
80. Denev S, Staykov Y, Moutafchieva R, Beev G. Microbial ecology of the gastrointestinal tract of fish and the potential application of probiotics and prebiotics in finfish aquaculture. *Int Aquat Res.* 2009; 1:1-29.
 81. Anderson DP, Siwicki AK. Duration of protection against *Aeromonas salmonicida* in brook trout immunostimulated with glucan or chitosan by injection or immersion. *Prog Fish Cul.* 1994; 56(4):258-261.
 82. Sahoo PK, Mukherjee SC. Influence of the immunostimulant, chitosan on immune responses of healthy and cortisol treated rohu (*Labeo rohita*). *J Aqua Trop.* 1999; 14:209-215.
 83. Tuan TN, Duc PM, Hatai K. Overview of the use of probiotics in aquaculture. *Int J Res Fish Aquacult.* 2013; 3(3):89-97.
 84. Hoseinifar SH, Ringø E, Masouleh AS, Esteban MA. Probiotic, prebiotic and synbiotic supplements in sturgeon aquaculture: a review. *Rev Aquacul.* 2014; 6:1-14.
 85. Hoseinifar SH, Mirvaghefi A, Merrifield DL, Amiri BM, Yeighi S, Bastami KD *et al.* The study of some haematological and serum biochemical parameters of juvenile beluga (*Huso huso*) fed oligofructose. *Fish Physiol Bioch.* 2011b; 37:91-96.