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Bio-Management of Root-Knot Nematode, *Meloidogyne incognita* on Cowpea (*Vigna unguiculata* L.)

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Abstract

A pot culture experiment was conducted to study the effect of fungal and bacterial bio-agents viz. *Trichoderma harzianum*, *T. viride* and *Pseudomonas fluorescens* @ 5 g, 7.5 g and 10 g/kg seed against root-knot nematode, *M. incognita* infecting cowpea. Chemical check (carbosulfan 3% w/w) and untreated check were also maintained for comparison. The results of the present study revealed that the maximum shoot length, root length, shoot weight and root weight and minimum number of galls/plant, egg masses/plant, eggs/egg mass, nematode population/100cc and total nematode population were recorded with *T. harzianum* @ 10 g/kg seed followed by *T. viride* @ 10 g/kg seed and *P. fluorescens* @ 10 g/kg seed *T. harzianum* @ 10 g/kg seed over the control.

Keywords: Cowpea, *Trichoderma harzianum*, *T. viride*, *Pseudomonas fluorescens*, *M. incognita*

1. Introduction

The pulses form an integral part of the cropping system of farmers all over the country because these crops fit well in the crop rotation and occupy an important position in the human dietary, being a good source of vegetable protein (17-43%) and supplement to cereal based diet [10]. In addition to this, these crops play a vital role in increasing soil fertility by fixing atmospheric nitrogen through root nodules [15].

Cowpea (*Vigna unguiculata* L.) is one of the important *kharif* pulse grown in India [12]. Pulse crops have been the backbone of agricultural economy in India having a reasonably higher annual output. In India pulses are being cultivated in 25.51 million ha with the production of 17.29 million tonnes and the average production being 630 kg/ha during 2010-11 [2]. In Rajasthan cowpea is extensively grown in Ajmer, Bhilwara, Churu, Jaipur, Jhunjhunu, Nagaur, Pali, Sikar, Sirohi and Tonk district, with an area of about 1.05 lakh ha with 73.39 thousand tonnes production and average production being 697 kg/ha during 2011-12 [1].

The production of pulse crops unfortunately suffers from several constraints of which pest and disease are the most important one. Among pests, phytoparasitic nematodes have been recognized as one of the major constraint in pulse production [5]. The extent of losses due to nematodes especially in pulse crops is yet to be estimated properly but in cowpea production is estimated to cause annual yield losses of nearly 15% worldwide [14]. In India average loss caused by root-knot nematode on pulses may be 14.6% which could go as high as 50-80% in some crops [5]. In another study estimated 28.60 per cent losses due to root-knot nematode, *M. incognita* in cowpea [13].

There are number of practices for management of plant parasitic nematodes in which chemical nematicides is used against nematodes by farmer because it is effective, easy to apply, and show rapid effects. But on the other hand it may cause degradation in soil fertility, environmental pollution and also hazardous for animals and human being. That's why biological control are more promising management practice and also economically & ecofriendly. The present experiment was conducted to study the effect of fungal and bacterial bio-agents viz. *Trichoderma harzianum*, *T. viride* and *Pseudomonas fluorescens* @ 5 g, 7.5 g and 10 g/kg seed against root-knot nematode, *M. incognita* infecting cowpea.

2. Material and methods

The present experiment was laid out in 6" earthen clay pots which contained 1kg infested soil with an initial inoculum level of 2 larvae per g of soil and replicated three times.

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Talc-based formulations of *T. viride*, *T. harzianum* and *P. fluorescens* were used as seed dressing each @ 5 g, 7.5 g and 10 g per kg seed with given sufficient moisture. Untreated and chemical checks (Carbosulfan 3% w/w) were also maintained for comparison. After 10 days of germination one healthy plant in each pot was maintained and others were uprooted carefully. The pots were then watered regularly almost every day. At the expiry of 45 days of sowing observations were recorded. While harvesting, the care was taken to avoid losses of both roots and nematodes in adhering soil. Observation on various growth parameters viz. fresh root and shoot weights, shoot and root lengths were recorded without delay whereas for studying nematode infestation, the plant tissues were stained in 0.1% acid fuchsin in lacto phenol at 80 °C for 2-3 minutes [11]. Then after gentle wash, roots were kept in clear lacto phenol for 24 hours. These roots were then examined under the stereoscopic binocular microscope for counting the number of galls, number of egg masses per plant and number of eggs per egg mass. The soil, after removing the plant from each pot was mixed well and 100cc soil from each pot was processed by Cobb's sieving and decanting technique [7] followed by modified Baermann's funnel technique [6] for the estimation of nematode population in soil.

2. Result and discussion

Data presented in Table 1 revealed that all fungal and bacterial bio-agents applied significantly increased the plant

growth and reduced nematode reproduction as compared to untreated check. The maximum shoot length (31.20 cm), root length (23.20 cm), shoot weight (19.60 g) and root weight (8.11 g) were recorded with *T. harzianum* @ 10 g/kg seed followed by *T. viride* @ 10 g/kg seed (30.50 cm, 22.90 cm, 19.20 g and 7.83 g) and *P. fluorescens* @ 10 g/kg seed (29.20 cm, 22.50 cm, 18.90 g and 7.61 g). However, maximum shoot length (34.80 cm), root length (25.02 cm), shoot weight (20.80 g) and root weight (8.33 g) were observed with seed dressing of carbosulfan 3% w/w while minimum shoot length (19.50 cm), root length (13.20 cm), shoot weight (12.13 g) and root weight (3.42 g) were observed with untreated check. Data presented in Table 2 showed that the minimum number of galls/plant (29.32), egg masses /plant (16.33), eggs/egg mass (100.67), nematode population/100cc (114) and total nematode population (1787) observed with *T. harzianum* @ 10 g/kg seed followed by *T. viride* @ 10 g/kg seed (30.91, 18.00, 108.00, 142 and 2116) and *P. fluorescens* @ 10 g/kg seed (32.03, 20.33, 112.67, 150 and 2473). However, minimum number of galls (20.43), egg masses/plant (11.67 eggs/egg mass (81.00), nematode population/100cc (84) and total nematode population (1050) observed with seed dressing of carbosulfan 3% w/w while maximum galls/plant, egg masses/plant, eggs/egg mass, nematode population/100cc and total nematode population were observed with untreated check 49.08, 35.33, 205.33, 360 and 7664 respectively.

Table 1: Effect of bio-agents at three different doses as seed treatment on plant growth characters cowpea

Treatments		Plant growth characters			
		Shoot length (cm)	Root length (cm)	Shoot weight (g)	Root weight (g)
<i>T. viride</i> 5g/kg seed	T ₁	25.90	21.50	17.00	6.50
<i>T. viride</i> 7.5g/kg seed	T ₂	28.30	22.10	18.20	6.82
<i>T. viride</i> 10g/kg seed	T ₃	30.50	22.90	19.20	7.83
<i>T. harzianum</i> 5g/kg seed	T ₄	26.40	21.70	17.30	6.63
<i>T. harzianum</i> 7.5g/kg seed	T ₅	28.90	22.40	18.50	7.32
<i>T. harzianum</i> 10g/kg seed	T ₆	31.20	23.20	19.60	8.11
<i>P. fluorescens</i> 5g/kg seed	T ₇	25.10	21.20	16.80	6.41
<i>P. fluorescens</i> 7.5g/kg seed	T ₈	27.50	21.80	17.70	6.72
<i>P. fluorescens</i> 10g/kg seed	T ₉	29.20	22.50	18.90	7.61
Chemical check (Carbosulfan 3% w/w)	T ₁₀	34.80	25.02	20.80	8.33
Untreated check	T ₁₁	19.50	13.20	12.13	3.42
SEm±		1.036	0.760	0.631	0.215
CD (P=0.05)		3.038	2.228	1.851	0.632

Note: (i) Data are average value of three replications
(ii) Initial inoculums level 2 juvenile / g soil

Table 2: Effect of bio-agents against root-knot nematode, *M. incognita* at three different doses as seed treatment on cowpea

Treatments		Nematode reproduction				
		No. of galls/plant	No. of egg masses/plant	No. of eggs/ egg mass	Nematode population/ 100cc soil	Total population
<i>T. viride</i> 5g/kg seed	T ₁	37.12	29.33	130.67	230	4100
<i>T. viride</i> 7.5g/kg seed	T ₂	34.68	24.00	119.00	183	3074
<i>T. viride</i> 10g/kg seed	T ₃	30.91	18.00	108.00	142	2116
<i>T. harzianum</i> 5g/kg seed	T ₄	35.93	27.67	127.33	206	3765
<i>T. harzianum</i> 7.5g/kg seed	T ₅	31.51	22.67	116.33	153	2822
<i>T. harzianum</i> 10g/kg seed	T ₆	29.32	16.33	100.67	114	1787
<i>P. fluorescens</i> 5g/kg seed	T ₇	40.71	31.00	133.33	243	4417
<i>P. fluorescens</i> 7.5g/kg seed	T ₈	36.53	26.00	124.00	199	3459
<i>P. fluorescens</i> 10g/kg seed	T ₉	32.03	20.33	112.67	150	2473
Chemical check (Carbosulfan 3% w/w)	T ₁₀	20.43	11.67	81.00	84	1050
Untreated check	T ₁₁	49.08	35.33	205.33	360	7664
SEm±		1.366	0.893	3.159	8.297	101.557
CD (P=0.05)		4.006	2.618	9.266	24.334	297.856

Note: (i) Data are average value of three replications
(ii) Initial inoculums level 2 juvenile / g soil

These findings are in agreement with the results of powder formulation of *T. harzianum* as seed treatment for the management of root-knot and root rot disease complex, caused by the root-knot nematode *M. javanica* and the fungus *R. solani*, on soybean plants under greenhouse conditions. Number of galls, root galling, egg masses and disease severity were reduced sharply on plants treated with *T. harzianum* as seed treatment, compared with the non-treated plants [9]. Similarly, significantly increased the plant growth and reduced final nematode population, treated with *T. harzianum* and *P. fluorescens* over streptomycin [3]. *T. harzianum* and *P. fluorescens* as seed treatment to manage the disease complex caused by *M. incognita* and *R. solani* on. Both the bio-agents were found to be significantly effective in reducing the damage and increasing the growth parameters of okra as compared to the treatment inoculated with *M. incognita* and *R. solani* [4]. *T. viride*, *T. harzianum* and *P. fluorescens* each @ 10 g/kg seed increased the growth parameters of okra plants and root-knot nematode populations were reduced in all the treatments compared to inoculated control. Against root-knot nematode, *M. incognita* [8].

3. References

1. Anonymous. Directorate of Agriculture, Government of Rajasthan, Jaipur, 2012.
2. Anonymous. Agriculture statistics at a glance, Department of Agriculture and Cooperation, Ministry of Agriculture, Government of India, New Delhi, 2011.
3. Barua L, Bora BC. Comparative efficacy of *Trichoderma harzianum* and *Pseudomonas fluorescens* against *Meloidogyne incognita* and *Ralstonia solanacearum* complex in brinjal. Ind J Nemat. 2008; 38:86-89.
4. Bhagawati B, Chaudhary BN, Sinha AK. Management of *Meloidogyne incognita*-*Rhizoctonia solani* complex on okra through bioagents. Ind J Nemat. 2009; 39:156-161.
5. Bhatti DS. Role of nematodes in crop production futuristic approaches. In: Nematode pests of crops (Eds. D.S. Bhatti & R.K. Walia), CBS publisher & Distributors, Delhi, 1992, 344-357.
6. Christie JR, Perry VG. Removing nematode from soil. Proceedings of Heminthological Society of Washington. 1951; 18:106-108.
7. Cobb NA. Estimating the nematode population of soil. US Deptt Agr Bu Plant Ind Agr Tech Cir. 1918; 1:1-48.
8. Kumar V, Jain RK. Management of root-knot nematode, *Meloidogyne incognita* by *Trichoderma viride*, *T. harzianum* and bacterial antagonist *Pseudomonas fluorescens* as seed treatment on okra. Ind J Nemat. 2010; 40:226-228.
9. Mahdy ME, El-Shennawy RZ, Khalifa EZ. Biological control of *Meloidogyne javanica* and *Rhizoctonia solani* on soybean by formulation of *Bacillus thuringiensis* and *Trichoderma harzianum*. Arab Uni J Agril Sci. 2006; 14:411-423.
10. Mann HS. Pulse and our protein gap. Annals of Arid Zone. 1975; 14:1.
11. Mc Beth CW, Taylor AL, Smith AL. Note on staining nematodes in root tissues. Proc Helminth Soc Washington. 1941; 8:26.
12. Menon MV, Reddy DB, Pameela P, Krishnankutty J. Seed production in vegetable cowpea [*Vigna unguiculata* (L.) Walp.] Under integrated nutrient management. Legume Res, 2010; 33:299-301.
13. Parvatha Reddy P, Singh DB. Assessment of avoidable yield losses in okra, brinjal, French bean and cowpea due to root-knot nematode. III International Symposium of Plant Pathology, New Delhi, 1981, 93-94.
14. Sasser JN, Freckman DW. A World Perspective on Nematology. The role of the society. In: Vistas on Nematology. (Ed. By Veech, J.A. and Dickson, D.W.) Society of Nematologists, Hyattsville, Maryland. 1987, 7-14.
15. Sivakumar T. Review on Biofertilizers. Int. J Curr. Res. Chem. Pharma. Sci. 2014; 1:17-23