



E-ISSN: 2320-7078

P-ISSN: 2349-6800

JEZS 2017; 5(3): 1351-1356

© 2017 JEZS

Received: 17-03-2017

Accepted: 18-04-2017

Abdur Rauf

Pest Warning and Quality
Control of Pesticide, Agriculture
Department, Punjab, Pakistan.

Mahmood Ayyaz

Faculty of Agricultural Sciences
and Technologies, Omer
Halisdemir University, Nigde,
Turkey.

Farukh Baig

Faculty of Science and
Engineering, Queensland
University of Technology,
Brisbane, Australia.

Muhammad Nadir Naqqash

Faculty of Agricultural Sciences
and Technologies, Omer
Halisdemir University, Nigde,
Turkey.

Muhammad Jalal Arif

Department of Entomology,
Faculty of Agriculture,
University of Agriculture
Faisalabad, Pakistan.

Correspondence**Mahmood Ayyaz**

Faculty of Agricultural Sciences
and Technologies, Omer
Halisdemir University, Nigde,
Turkey.

Response of *Chilo partellus* (Swinhoe) and entomophagous arthropods to some granular and new chemistry formulations in *Zea mays* L.

Abdur Rauf, Mahmood Ayyaz, Farukh Baig, Muhammad Nadir Naqqash and Muhammad Jalal Arif

Abstract

Comparative efficacy of granular (Carbofuran, cartap and monomehypo) and new chemistry foliar insecticides (Chlorantraniliprole, fipronil, spinosad and flubendiamide) was evaluated against *C. partellus*. Effect of insecticides on entomophagous arthropods (coccinellids and spiders species) was also focused in maize (cultivar: AAS-9732). Maximum mortality was observed in carbofuran and fipronil i.e. 89.3 ± 11.13 and 80.9 ± 7.98 adults of *C. partellus* per five plants due to carbofuran and fipronil respectively. Minimum loss was recorded in carbofuran and fipronil treatments viz. minimum number of dead hearts i.e. 0.1 ± 0.12 and 0.20 ± 0.00 were found in carbofuran and fipronil treated plots respectively. Systemic nature of these broad-spectrum insecticides (carbofuran and fipronil) has significantly enhanced their toxicity. However the problem with these insecticides was significantly higher mortality of natural enemies in these treatments. Concluding, relatively safer and targeted insecticides like spinosad should be included in integrated pest management program to allow natural enemies to flourish well. Additionally, research should be conducted to explore targeted safer insecticides.

Keywords: *Chilo partellus*, insecticidal efficacy, maize, natural enemies

Introduction

Maize (*Zea mays* L.), member of family Poaceae, is the highest yielding and genetically diverse cereal crop in the world [1, 2]. It has great significance especially in developing countries like Pakistan. This crop accounts for 4.8 % of the total cropped area and 3.5 % of the value towards agricultural output in Pakistan [3, 1]. Its area under cultivation is around 0.9 million hectares while the annual production is nearly 1.3 million tons in Pakistan. The lesser yield is primarily due to its susceptibility to a wide range of insect pests and diseases in different ecological zones in Pakistan. The crop is infested by more than one hundred insect species. Among insect-pests, stem borers are most serious pests of maize [4, 5]. The spotted stem borer (*Chilo partellus* Swinhoe) is considered as the most notorious pests for its damage to this crop [6].

Infestation of *C. partellus* at seedling stage may cause total failure of crop [7, 8]. Its larvae bore into the midrib and stalk, feed inside the plant stems. Feeding behavior of this pest protects it from natural parasitoids and insecticides [9]. The *C. partellus* not only causes mechanical damage (lodging of plant due to weakened stem) but also causes characteristic perforation in leaves called fenestrations [10]. This kind of damage results in reduction of photosynthetic area of the leaves resulting in poor cereal yield. Estimated losses vary greatly in different regions and agro-ecological zones. Severe damage of this pest may lead to grain reduction upto 81 % [11, 12].

Higher infestation of *C. partellus* is also due to unavailability of resistant cultivars which play an important role in management of this pest. The cultural practices viz., removal and destruction of infested plants residues can be helpful in controlling this pest [5]. Although, biological control agents like ants, spiders, earwigs etc., play a key role in insect-pest management but to a limited extent [13, 14].

Chemical control is the mostly used method for control of insect-pests due to its quicker and immediate results [15, 16]. The use of contact and systemic insecticides is considered most effective [17, 18] at initial stage of infestation to get rid of the *C. partellus* larvae before burrowing into the stems [5, 12]. Insecticides particularly granular formulations are recommended as soil and/or whorl application after 25 and 45 days of sowing [19, 20].

Furthermore, the foliar application of insecticide instead of granular formulation is recommended for effective control [21]. This pest can also be managed through whorl application of insecticides (granular and foliar) in liquid formulations [22]. However, the great problem with the use of insecticides is their lethal or sub-lethal effect on non-target organisms especially bio-control agents [23].

Therefore, present study was conducted to investigate the efficacy of some granular and foliar insecticides to *C. partellus* along-with the reduction in post-treatment crop losses. Furthermore, the effect of these insecticides on entomophagous insects was also investigated.

Materials and Methods

The maize cultivar (AAS-9732) was sown at experimental area of the University of Agriculture Faisalabad, Pakistan (Latitude: 31.432852°; Longitude: 73.069406°) during spring season of the year 2011.

Insecticides

The granular insecticides used in experiment were carbofuran (Carbofuran® 3G; Arrow International Lahore, Pakistan), cartap (Reject® 3G; Pakistan Agrochemicals Karachi, Pakistan) and monomehypo (C-Moon® 5G; Capricon Associated Karachi, Pakistan). While the tested new-chemistry foliar insecticides were chlorantraniliprole (Coragen® 20 SC; Bayer Crop Sciences, Karachi, Pakistan), fipronil (Regent® 80 WG; Bayer Crop Sciences, Karachi, Pakistan), spinosad (Tracer® 240 SC; Arysta Life Sciences Karachi, Pakistan) and flubindiamide (Belt® 48 SC; Bayer Crop Sciences, Karachi, Pakistan). These 8 insecticides were randomized

Treatments

The recommended Agronomic practices were done before sowing the crop. The crop was sown during the normal season of sowing in Randomized Complete Block Design (RCBD), maintaining row to row and plant to plant distance of 1 sq. ft and 0.9 sq. ft, respectively. The plot size was 1100m (3612sq.ft). Insecticides were applied at peak of vegetative stage when there was enormous population of test insect-pests. The experiment consisted of 8 insecticidal treatments which were randomized thrice so the total experimental units were 24 and these were separated from each other by a path as shown in the layout.

Data collection

Different parameters viz., larval density, mortality of *C. partellus* and natural enemies were measured in relation to stem tunneling, dead hearts and leaf holes. To determine the larval population five plants were randomly selected and dissected longitudinally, in each replication. While factors like population of entomophagous arthropods viz., coccinellids and spiders spp. were estimated by visual observation by selecting five randomly plants per treatment. Tunneling was assessed by dissecting the randomly selected stems while number of infested stems, dead hearts and leaf holes were observed visually in five randomly selected plants per replication. The data regarding infestation, larvae/plant, tunnel length inside stem, leaf holes, dead hearts and entomophagous arthropods were recorded at 3, 7 and 14 days post treatment period respectively [24, 25].

Statistical Analysis

The data were statistically analyzed using analysis of variance

(ANOVA) and level of significance among treatments was determined using SAS software. The level of significance between treatments means were analyzed by LSD (Least Significance Difference) test using MSTAT-C program [47].

Results

Impact of insecticides on population density of insects

Chilo partellus

Significant variations were observed in population density of *C. partellus* after 3, 7 and 14 days of treatment. Among the granular insecticides, carbofuran was proved to be most effective while among foliar formulations, fipronil was the most effective insecticide in controlling *C. partellus* larvae. After carbofuran treatment larval density was 0.7 ± 0.12 , 0.40 ± 0.20 and 0.10 ± 0.12 larvae per 5 plants after 3, 7 and 14 days of treatment respectively. While, average larval density in plots treated with fipronil was 0.90 ± 0.20 , 0.50 ± 0.12 and 0.20 ± 0.12 larvae/5 plants after 3, 7 and 14 days of treatment respectively (Table 1).

Coccinellids

Among granular insecticides, minimum numbers of coccinellids were observed in carbofuran while among foliar formulations maximum mortality was found in fipronil. Population density of coccinellids in the plots treated with carbofuran was 0.7 ± 0.12 , 0.3 ± 0.11 and 0.3 ± 0.11 coccinellids/5 plants respectively. Whereas, population density of coccinellids spp. was 0.7 ± 0.12 , 0.5 ± 0.23 and 0.4 ± 0.2 coccinellids/5 plants after 3, 7 and 14 days in fipronil respectively (Table1).

Spider species

Minimum population of spiders was found in carbofuran (among granular insecticides) and spinosad (among new chemistry insecticides) treated plots. Average density of spider spp., in plots treated with carbofuran was 0.1 ± 0.12 , 0.4 ± 0.31 and 0.26 ± 0.12 spiders/5 plants after 3, 7 and 14 days of treatment. Whereas, in case of spinosad, average larval density was 0.5 ± 0.42 , 0.2 ± 0.20 and 0.4 ± 0.20 spiders/5 plants after 3, 7 and 14 days of treatment (Table I).

Results

Impact of insecticides on percent mortality of insects

Chilo partellus

Maximum percentage mortality of *C. partellus* larval was recorded in carbofuran (group of granular insecticides) and fipronil (group of new chemistry insecticides). Among the granular insecticides treated plots, minimum larval density of *C. partellus* was observed i.e. 40.00 ± 20.00 , 48.01 ± 31.7 and 38.70 ± 36.43 per 5 plants in case of cartap after 3, 7 and 14 days respectively. Maximum larval mortality viz., 63.33 ± 5.77 , 75.20 ± 7.71 and 89.26 ± 11.13 per 5 plants was observed in the carbofuran treated plots after 3, 7 and 14 days of treatment respectively. Whereas, minimum larval mortality recorded was 30.00 ± 10.00 , 43.56 ± 8.96 and 46.90 ± 19.05 per 5 plants in chlorantraniliprole treated plots. While the maximum mortality i.e. 56.67 ± 5.77 , 66.14 ± 15.13 and 80.92 ± 7.98 per 5 plants was found in the case of fipronil after 3, 7 and 14 days of treatment respectively (Table 2).

Coccinellids species

Percent mortality of coccinellids varied significantly among the treatments. Among the granular insecticides treated plots, minimum percent mortality of coccinellids species was

30.07±15.25, 30.00±12.58 and 27.80±25.46 per 5 plants in plots treated with monomehypo after 3, 7 and 14 days respectively. Whereas, maximum mortality i.e. 54.23±3.75, 60.00±17.32 and 58.33±22.05 per 5 plants was found in case of carbofuran after 3, 7 and 14 days of treatment respectively. Minimum percent mortality was found in spinosad treatment i.e. 25.13±16.77, 23.33±16.07 and 27.80±25.46 per 5 plants after 3, 7 and 14 days of treatment respectively. Maximum percent mortality was recorded in fipronil i.e. 47.61±17.17 per 5 plants after 3 days of treatment, while highest mortality after 7 and 14 days of mortality was observed in flubendiamide viz., 53.33±5.77 and 47.22±24.06 per 5 plants respectively (Table 2).

Spider species

Among the granular insecticide treated plots, minimum observed percent mortality of spiders was 33.33±30.55 and 25.00±25.00 per 5 plants in monomehypo treated plot after 3 and 7 days respectively. Maximum percent mortality i.e. 71.66±10.40 and 55.60±9.60 after 3 and 7 days of treatment respectively was recorded in carbofuran treated plots. While, after 14 days there was no significant difference among the plots treated with granular insecticides.

Among the plots treated with new chemistry foliar formulations, minimum average percent mortality viz., 35.00±21.79, 52.77±21.00 was recorded in flubindiamide after 3 and 7 days of treatment. While minimum mortality 27.77±16.70 per 5 plants was recorded in spinosad treated plot after 14 days of treatment. Maximum percent mortality i.e. 63.30±15.28, 63.90±12.70 per 5 plants recorded in chlorantraniliprole after 3 and 7 days of treatment. While maximum mortality i.e. 61.10±25.50 per 5 plants was recorded in flubindiamide treated plot after 14 days of treatment respectively (Table 2).

Impact of insecticides on reduced losses

Reduction in stem tunneling

The results showed a non-significant variation among the treatments, on stem tunneling by *C. partellus*. Among the granular insecticides treated plots, minimum average stem tunneling done by *C. partellus* was 2.7±0.56 cm in carbofuran treated plots after 3 days of treatment. While after 7 and 14 days of treatment 1.90±0.35 and 1.70±0.55 cm tunnel was recorded in monomehypo treated plots. Whereas, maximum stem tunneling among granular insecticide treated plot was recorded in cartap treated plot i.e. 4.30±1.26 cm after 3 days, while after 7 days 3.70±1.12 cm tunnel was found in carbofuran and after 14 days of treatment 2.40±0.51 cm tunnel was recorded in cartap treated plots. Among the plots

treated with new chemistry foliar formulation minimum stem tunneling was 2.70±1.01 cm in fipronil after 3 days, while 1.70±0.70 cm and 0.90±0.81 cm stem tunneling was recorded in chlorantraniliprole after 7 and 14 days of treatment. Whereas maximum stem tunneling 3.10±1.12 and 2.50±0.64 cm in spinosad after 3 and 7 days of treatment, while 2.10±0.82 cm stem tunneling was observed in fipronil treated plot after 14 days of treatment (Table 3).

Reduction in dead hearts

Among the granular insecticides treated plots minimum number of dead heart caused by *C. partellus* were 0.30±0.12, 0.20±0.00 and 0.10±0.12 dead hearts/5 plants after 3, 7 and 14 days of treatment respectively in carbofuran. Maximum dead hearts 0.50±0.00, 0.40±0.12 and 0.30±0.12 dead heart/5 plants were found in monomehypo after 3, 7 and 14 days of treatment respectively. Whereas in plots treated with foliar formulations minimum dead heart 0.20±0.00/5 plants were observed in spinosad after 3 days, while 0.20±0.00 dead hearts/5 plants were observed in flubendiamide treated plots after 7 days post treatment period. After 14 days, 0.20±0.00 dead hearts/5 plants were found in fipronil treated plots. Maximum dead hearts 0.40±0.12 dead heart/5 plants were found in chlorantraniliprole treated plots after 3 and 7 days of treatment. While 0.40±0.12 dead hearts/5 plants were observed in flubindiamide after 14 days of treatment (Table 3).

Reduction in leaf holes

Among the granular tested insecticide plots minimum leaf holes i.e. 0.33±0.12 leaf holes/5 plants in plots treated with cartap after 3 days of treatment, while 0.33±0.12 leaf holes/5 plants were observed in case of monomehypo after 7 days of treatment. After 14 days of treatment 0.20±0.00 leaf holes/5 plants were found in carbofuran treatment. Maximum 0.50±0.12 leaf holes/5 plants in monomehypo and cartap treated plots after 3 and 7 days of treatment, while 0.40±0.12 leaf holes/5 plants were found in monomehypo treated plots after 14 days of treatments. Among foliar formulation minimum number of leaf holes 0.26±0.12, 0.30±0.10 and 0.10±0.12 leaf holes/5 plants were found in spinosad treated plots after 3, 7 and 14 days of treatment respectively. Maximum number of leaf holes 0.30±0.10 and 0.40±0.23 were found in chlorantraniliprole after 3 and 7 days of treatment. While 0.30±0.12 leaf holes/5 plants were found in flubindiamide treated plots after 14 days of treatments (Table 3).

Table 1: Impact of insecticides on larval density of *Chilo partellus*, Coccinellids and spider species during 2011.

| Treatments | | Larval Density of <i>C. partellus</i> | | | Coccinellids spp. density | | | Spiders spp. | | |
|-----------------------|---------------------|---------------------------------------|--------------|---------------|---------------------------|--------------|---------------|--------------|--------------|---------------|
| | | After 3 days | After 7 days | After 14 days | After 3 days | After 7 days | After 14 days | After 3 days | After 7 days | After 14 days |
| Granular Insecticides | Carbofuran | 0.7±0.12e | 0.4±0.2c | 0.1±0.12b | 0.7±0.12d | 0.3±0.11c | 0.3±0.11b | 0.1±0.12b | 0.4±0.31bc | 0.26±0.12ab |
| | Cartap | 1.2±0.4bcd | 0.8±0.35bc | 0.6±0.2b | 0.8 ±0.2bcd | 0.4±0.2bc | 0.3 ±0.11b | 0.3±0.12ab | 0.5±0.12ab | 0.3±0.12ab |
| | Monomehypo | 1.1±0.12bcd | 0.8±0.2bc | 0.3±0.12b | 1.0 ±0.2bc | 0.5±0.11bc | 0.5±0.11ab | 0.6±0.20a | 0.4±0.20bc | 0.26±0.12ab |
| Foliar Insecticides | Chlorantraniliprole | 1.4±0.2b | 0.9±0.12b | 0.6±0.2b | 1.1±0.12b | 0.6 ±0.2b | 0.5±0.11ab | 0.3±0.12ab | 0.3±0.12bc | 0.26±0.12ab |
| | Fipronil | 0.9±0.2de | 0.5±0.12bc | 0.2±0.12b | 0.7 ±0.12cd | 0.5±0.23bc | 0.4 ±0.2b | 0.6±0.00a | 0.3±0.12bc | 0.26±0.23ab |
| | Spinosad | 1.3±0.23bc | 0.9±0.12b | 0.5±0.23b | 1.1±0.12b | 0.6 ±0.2b | 0.5±0.11ab | 0.5±0.42ab | 0.2±0.20c | 0.4±0.20ab |
| | Flubindiamide | 0.9±0.23cde | 0.6±0.2bc | 0.3±0.12b | 0.9±0.12bcd | 0.4 ±0.2bc | 0.3 ±0.11b | 0.6±0.20a | 0.4±0.12bc | 0.2±0.20b |
| Control | | 2.0±0a | 1.7±0.44a | 1.2±0.67a | 1.5±0.31a | 0.9±0.11a | 0.7±0.11a | 0.7±0.42a | 0.7±0.12a | 0.5±0.12a |

Table 2: Impact of insecticides on percent mortality of *Chilo partellus*, Coccinellids and spider species

| Treatments | Larval mortality of <i>C. partellus</i> | | | Mortality of Coccinellids spp. | | | Mortality of Spiders spp. | | |
|-------------------------------|---|---------------|---------------|--------------------------------|---------------|---------------|---------------------------|--------------|---------------|
| | After 3 days | After 7 days | After 14 days | After 3 days | After 7 days | After 14 days | After 3 days | After 7 days | After 14 days |
| Carbofuran | 63.3±5.77a | 75.2±7.41a | 89.3±11.13a | 54.2±3.75a | 60.0±17.32a | 58.3±22.05a | 71.7±10.4a | 55.6±9.6ab | 50.0±16.7 a |
| Cartap | 40.0±20.0bcd | 48.0±31.7bc | 38.7±36.43d | 44.7±14.25ab | 53.3±25.66ab | 50.0±16.67a | 65.0±8.66ab | 25.0±25.0bc | 38.9±9.7a |
| Monomehypo | 43.3±5.77bcd | 48.4±27.77bc | 69.3±16.72abc | 30.7±15.25bc | 30.0±12.58bc | 27.8±25.46ab | 33.3±30.55b | 41.7±38.0ab | 50.0±16.7a |
| Chlorantraniliprole | 30.0±10.0d | 43.6±8.96c | 46.9±19.05cd | 26.1±8.58c | 31.7±16.07bc | 30.6±4.81ab | 63.3±15.28ab | 63.9±12.7a | 50.0±16.7a |
| Fipronil | 56.7±5.77ab | 66.1±15.13ab | 80.9±7.98a | 47.6±17.17a | 46.7±25.66abc | 38.9±34.69a | 58.3±17.56ab | 63.9±12.7a | 50.0±16.7a |
| Spinosid | 36.7±11.55cd | 46.9±13.34bc | 49.4±35.37bcd | 25.1±16.77c | 23.3±16.07cd | 27.8±25.46ab | 51.7±20.21ab | 63.9±12.7a | 27.8±16.7ab |
| Flubindiamide | 53.3±11.55abc | 62.6±15.55abc | 77.6±2.51ab | 40.2±6.01abc | 53.3±5.77ab | 47.2±24.06a | 35.0±21.79b | 52.8±21.0ab | 61.1±25.5a |
| Control | 0.00 e | 0.0±0.0d | 0.0±0.0 e | 0.0d | 0.00±0d | 0.0±0sb | 00.0±c | 0.0±0c | 00.0±34.7b |
| Standard error for comparison | 8.93 | 9.5 | 14.038 | 7.5471 | 12.404 | 15.35 | 15.401 | 15.92 | 16.00 |

Table 3: Impact of insecticides on stem tunneling dead heart and leaf holes produced by *C. partellus*

| Treatments | Stem tunneling | | | Dead heart | | | Leaf holes | | |
|---------------------|----------------|--------------|---------------|--------------|--------------|---------------|--------------|--------------|---------------|
| | After 3 days | After 7 days | After 14 days | After 3 days | After 7 days | After 14 days | After 3 days | After 7 days | After 14 days |
| Carbofuran | 2.7±0.56b | 2.8±0.08bc | 2.1±0.14bc | 0.3±0.12bc | 0.2±0.00b | 0.1±0.12b | 0.4±0.00ab | 0.5±0.12ab | 0.2±0.00b |
| Cartap | 4.3±1.26ab | 3.7±1.12ab | 2.4±0.51ab | 0.5±0.00ab | 0.3±0.12b | 0.2±0.20b | 0.33±0.12ab | 0.33±0.23b | 0.3±0.12ab |
| Monomehypo | 3.8±0.30ab | 1.9±0.35c | 1.7±0.55bcd | 0.5±0.00ab | 0.4±0.12b | 0.3±0.12ab | 0.5±0.12a | 0.33±0.12b | 0.4±0.12ab |
| Chlorantraniliprole | 2.8±0.94b | 1.7±0.70c | 0.9±0.81d | 0.4±0.12abc | 0.4±0.12b | 0.3±0.12ab | 0.3±0.10ab | 0.4±0.12b | 0.1±0.12b |
| Fipronil | 2.7±1.01b | 2.1±0.65c | 2.1±0.82bc | 0.3±0.12abc | 0.4±0.12b | 0.2±0.00b | 0.27±0.12b | 0.33±0.12b | 0.2±0.00b |
| Spinosid | 3.1±1.12b | 2.5±0.64bc | 1.5±0.61bcd | 0.2±0.00c | 0.3±0.12b | 0.3±0.12ab | 0.26±0.12b | 0.3±0.10b | 0.1±0.12b |
| Flubindiamide | 2.8±1.49b | 2.3±0.60c | 1.3±0.72cd | 0.3±0.12bc | 0.2±0.00b | 0.4±0.12ab | 0.3±0.10ab | 0.4±0.23b | 0.3±0.12ab |
| Control | 5.5±1.3a | 4.8±0.85a | 3.3±0.96a | 0.6±0.12a | 0.5±0.12a | 0.5±0.12a | 0.6±0.12a | 0.6±0.00a | 0.5±0.15a |

Discussion

Over-reliance on chemicals has created resistance in insect-pests, leading towards collapse of pest management programs [47, 48]. Additionally, various problems are also linked with excessive and uncontrolled usage of insecticides such as non-target effects, human health concern, adverse effects on environmental surroundings and many others. Eco-friendly compatible control methods and approaches must be adopted in order to manage the insect-pests such as integrated pest management. Concluding, chemical control is the main tool of controlling *C. partellus* but this tool can be synchronized with biological control for better management [48, 49].

Carbofuran, a widely used carbamate insecticide, has recently been banned in many countries [26, 27]. It has systemic anticholinergic actions in cortex and striatum regions of central nervous system by forming carbofuran acetylcholinesterase complex [28]. Along with various systemic effects many other abnormalities are also caused by this insecticide viz., sterility [29, 30, 31] congenital abnormalities [28, 32] hepatotoxicity, enhanced risk of dysfunctions on gastrointestinal [33, 34] neurological [35, 46], and endocrine systems [33]. Carbofuran was the most effective treatment which not only eliminated leaf injury but also significantly reduced the pest population [24, 25]. Lethal and sub-lethal effects of carbofuran on coccinellids are well known [36, 37]. Carbofuran also significantly affect the predatory spiders' population [38].

Fipronil (a phenylpyrazole compound) was developed in the mid-1990s and became an effective insecticide especially due to its high toxicity against insects resistant to conventional insecticides and its lower toxicity to mammals [39]. Fipronil potentially blocks the insect GABA-gated chloride channels at nanomolar concentrations [40]. It is clear from previous findings that fipronil significantly affects the population of borers due to susceptibility of insects to new chemistry insecticides like fipronil. Additionally, it also reduces damage of insect-pests [41, 42]. Fipronil also significantly affects non-target organisms like spiders, coccinellids etc. [43].

Chlorantraniliprole, a new chemistry insecticide (anthranilic

diamide in nature), is effective to a number of insect-pests [44]. It is an activator of insect ryanodine receptors, causing rapid muscle dysfunction and paralysis [43, 45]. It is extensively being used for the effective management of lepidopteran pests and as a result preventing damages caused in different crops [44].

Conclusion

However, carbofuran and fipronil killed the maximum population of *C. partellus*, but they are also detrimental to the population of natural enemies. So, insecticides like spinosad should be included in integrated pest management programs owing to satisfactory pest control with minimum damage to predators' population. Additionally, research should be done to explore the potentially safer and targeted insecticides not only in maize crop but also for other crops. Insecticide rotation should be also included in pest management programs to manage resistance in insect-pests.

Acknowledgements

The authors are very grateful to the local and international pesticides companies for supplying pesticides used for the experimental trails.

Conflict of interest statement: The Author declares that there is no conflict of interest.

References

1. Ferdu AK, Demissew J, Birhane A. Major insect pests of maize and their management. Journal of Pesticide Science. 2002; 85:109-115.
2. Channappagoudar BB, Biradar NR, Patil JB, Hiremath SM. Assessment of sweet Sorghum genotype for come yield, juice characters and sugar levels. Karnataka Journal of Agricultural Sciences. 2007, 294-296.
3. Anonymous, Pakistan Agriculture Research Council, Government of Sindh, Agriculture Department, 2005. <http://www.sindhagri.gov.pk/maize-about.html> (Accessed on 4-7-2013).
4. Songa JM, Zhon G, Verhold WA. Relationship of stem

- borer damage and plant physical condition to maize yield in a semi arid zone of Eastern Kenya. *International Journal of Tropical Insect Science*. 2001; 21:243-249.
5. Kfir R, Overholt WA, Khan ZR, Polaszek A. Biology and management of economically important Lepidopteran cereal stem borers in Africa. *Annual review of entomology*. 2002; 47:701-31.
 6. Kerns DL, Stewart SD. Sublethal effects of insecticides on the intrinsic rate of increase of cotton aphid. *Entomologia Experimentalis et Applicata*. 2000; 94:41-9.
 7. Sekhon SS, Kanta V. Effect of seed treatment and other insecticidal formulation on the maize borer (*Chilo partellus* Swin.). *Journal of Insect Science*. 1992; 5:45-47.
 8. Barrow MR. The effect of first generation maize stalk borer *Busseola fusca* (Fuller) (Lepidoptera: Noctuidae) on yield of different maize genotypes *Journal of the Entomological Society of Southern Africa*. 1987; 50:291-298.
 9. Siddalingappa, Thippeswamy C, Hosamani V, Yalavar S. Biology of maize stem borer, *Chilo partellus* (Swinhoe) (Crambidae: Lepidoptera *International Journal of Plant Protection* 2010; 3:91-93.
 10. Naerls, Recommended practices for the production of maize. Extension recommended practices No. 3. National Agricultural Extension and Research Liason Services, Ahmadu Bello University, Zaria, 1982.
 11. Panwar VPZ, Mukherjee BK, Ahuja VP. Maize inbreds tolerant to tissue borers, *Chilo partellus* and *Atherigona spp.* *Indian Journal of Genetics and Plant Breeding*. 2000; 60:71-75.
 12. Kumar H. Resistance in maize to larger grain borer, *Prostephanus truncates* (Horn) (Coleoptera: Bostrichidae *Journal of Stored Products Research*. 2002; 38:267-280.
 13. Saeed S, Saeed Q, Saeed R, Shafiq M, Jaleel W, Ishfaq M *et al.* Impact of Various Diets on Biological Parameters of *Chrysoperla Carnea Stephen* (Neuroptera: Chrysopidae) Adults under Controlled Conditions. *Applied Sciences and Business Economics*. 2014; 01:2-9.
 14. Mailafiya DM, Le-Ru BP, Kairu EW, Calatayud PA, Dupas S. Species diversity of lepidopteran stem borer parasitoids in cultivated and natural habitats in Kenya. *Journal of Applied Entomology*. 2009; 133:416-429.
 15. Afzal, MBS, Shad SA, Abbas N, Ayyaz M, Walker WB. Cross-resistance, the stability of acetamiprid resistance and its effect on the biological parameters of cotton mealybug, *Phenacoccus solenopsis* (Homoptera: Pseudococcidae), in Pakistan. *Pest management Sciences*. 2015; 71(1):151-158.
 16. Malik MU, Javed H, Ayyaz M. Evaluation of Different Groundnut *Arachis hypogea L.* Cultivars Against Termites, *Odontotermes obesus* (Rambur) in Rawalpindi, Pakistan. *Turkish Journal of Agriculture-Food Science and Technology*. 2015; 3(6).
 17. Sétamou M, Schulthess F. The influence of egg parasitoids belonging to *Telenomus busseolae* (Hym.: Scelionidae) species complex on *Sesamia calamistis* (Lepidoptera: Noctuidae) population in maize fields in southern Benin. *Biocontrol Science and Technology*. 1995; 5:69-81.
 18. Ndemah R, Gounou S, Schulthess F. The role of wild grasses in the management of new class of insecticides with a novel mode of action, ryanodine receptor activation. *Bulletin of entomological research*. 2002; 84:196-214.
 19. Halimie MA, Mughal MS, Mehdi SA, Rana ZA. Response of different timing of carbofuran (Furadan 3G) on the maize borer incidence and yield of the crop. *Journal of Agricultural Research*. 1989; 27:337-340.
 20. Khan MA, Waqarullah Y, Shah GS, Ishtiaq A, Liaquatullah M. Effect of different insecticide formulations and doses against maize stem borer in corn field. *Sarhad Journal of Agriculture*. 2004; 20(4):609-912.
 21. Shams MR, Afzal M. Effect of synthetic pyrethroids organophosphates on maize shoot fly: *Atherigona soccata* (Rond.) and maize borer: *Chilo partellus* (Swin). *Journal of Agriculture Research*. 1989; 27:65-70.
 22. Gunewardena KNC, Madugalla SRK. Efficacy of selected granular insecticides for the control of maize-stem borer (*Chilo partellus*) (Lepidoptera: Pyralidae). *Tropical Agricultural Research and Extension*. 2011; 14(1):12-15.
 23. Biondi A, Desneux N, Siscaro G, Zappalà L. Using organic-certified rather than synthetic pesticides may not be safer for biological control agents: selectivity and side effects of 14 pesticides on the predator *Orius laevigatus*. *Chemosphere*. 2012; 87:803-812.
 24. Ganguli RN, Chaudhary RN, Ganguli J. Effect of time of application of chemicals on management of maize stem borer, *Chilo partellus* (Swinhoe). *International Journal of Pest Management*. 1997; 43:253-259.
 25. Bhat ZH, Baba ZA. Efficacy of different insecticides against maize stem borer *Chilo partellus* (Swinhoe) and maize aphid *Rhopalosiphum maidis* (Fitch) infesting maize. *Pakistan. Entomology*. 2007; 29:73-76.
 26. Jemutai-Kimosop S, Orata FO, K'owino IO, Getenga ZM. The dissipation of carbofuran in two soils with different pesticide application histories within Nzoia River Drainage Basin, Kenya. *Bulletin of environmental contamination and toxicology*. 2014; 92:616-620.
 27. Otieno PO, Lalah JO, Virani M, Jondiko IO, Schramm KW. Carbofuran and its toxic metabolites provide forensic evidence for Furadan exposure in vultures (*Gyps africanus*) in Kenya. *Bulletin of environmental contamination and toxicology*. 2010; 84(5):536-544.
 28. Gupta RC. Carbofuran toxicity. *Journal of Toxicology and Environmental Health, Part A Current Issues*. 1994; 43(4):383-418.
 29. Baligar PN, Kaliwal BB. Reproductive toxicity of carbofuran to the female mice: effects on estrous cycle and follicles. *Industrial health*. 2002; 40:345-352.
 30. Pant N, Prasad AK, Srivastava SC, Shankar R, Srivastava SP. Effect of oral administration of carbofuran on male reproductive system of rat. *Human & experimental toxicology*. 1995; 14:889-894.
 31. Yousef MI, Salem MH, Ibrahim HZ, Helmi S, Seehy MA, Bertheussen K. Toxic effects of carbofuran and glyphosate on semen characteristics in rabbits. *Journal of Environmental Science & Health Part B*. 1995; 30:513-534.
 32. Maroni M, Colosio C, Ferioli A, Fait A. Biological monitoring of pesticide exposure: A review. *Toxicology*. 2000; 143:1-118.
 33. Ram RN. Carbofuran-induced histophysiological changes in thyroid of the teleost fish, *Channa punctatus* (Bloch). *Ecotoxicology and environmental safety*. 1988; 16:106-113.
 34. Rizos E, Liberopoulos E, Kosta P, Efremidis S, Elisaf M.

- Carbofuran-induced acute pancreatitis. *Journal of Pancreas*. 2004; 5:44-47.
35. Kamboj A, Kiran R, Sandhir R. Carbofuran-induced neurochemical and neuro-behavioral alterations in rats: attenuation by N-acetylcysteine. *Experimental Brain Research*. 2006; 170:567-575.
 36. Alexander A, Krishnamoorthy SV, Kuttalam S. Toxicity of Insecticides to the Coccinellid Predators, *Cryptolaemus Montrouzieri* Mulsant and *Scymnus coccivora* Ayyar of Papaya Mealybug, *Paracoccus marginatus* Williams and *Granara De Willink*. *Journal of Biological Control*. 2013; 27:18-23.
 37. Papachristos DP, Milonas PG. Adverse effects of soil applied insecticides on the predatory coccinellid *Hippodamia undecimnotata* (Coleoptera: Coccinellidae). *Biological Control*. 2008; 47:77-81.
 38. Tahir HM, Butt A, Mukhtar MK, Khan SY, Arshad M, Ahsan MM. Effect of carbofuran on the diversity and mean abundance of ground spiders. *African Journal of Biotechnology*. 2013; 10: 12303-12308.
 39. Tingle CC, Rother JA, Dewhurst CF, Lauer S, King WJ. Fipronil: environmental fate, ecotoxicology, and human health concerns. In *Reviews of environmental contamination and toxicology* Springer New York. 2003, 1-66.
 40. Zhao X, Salgado VL, Yeh JZ, Narahashi T. Differential actions of fipronil and dieldrin insecticides on GABA-gated chloride channels in cockroach neurons. *Journal of Pharmacology and Experimental Therapeutics*. 2003; 306:914-924.
 41. Teli VS, Chavan BP, Ankalkoppe MN, Khot RB, Harers PN. Evaluation of some insecticides for the control of maize stem borer, *Chilo partellus* (Swinhoe). *Journal of Entomological Research*. 2007; 31(4):323-326.
 42. Nault BA, Straub RW, Taylor AG. Performance of novel insecticide seed treatments for managing onion maggot (*Diptera: Anthomyiidae*) in onion fields. *Crop Protection*. 2006; 25:58-65.
 43. Pisa LW, Amaral-Rogers V, Belzunces LP, Bonmatin JM, Downs CA, Goulson D *et al.* Effects of neonicotinoids and fipronil on non-target invertebrates. *Environmental Science and Pollution Research*. 2014; 22:68-102.
 44. Hannig GT, Ziegler M, Marçon PG. Feeding cessation effects of chlorantraniliprole, a new anthranilic diamide insecticide, in comparison with several insecticides in distinct chemical classes and mode-of-action groups. *Pest Management Sciences*. 2009; 65(9):969-974.
 45. Lahm GP, Selby TP, Freudenberger JH, Stevenson TM, Myers BJ, Seburyamo G *et al.* Insecticidal anthranilic diamides: a new class of potent ryanodine receptor activators. *Bioorganic & Medicinal Chemistry Letters*. 2005; 15:4898-4906.
 46. Kim SJ, Kim JE, Ko BH, Moon IS. Carbofuran induces apoptosis of rat cortical neurons and down-regulates surface alpha7 subunit of acetylcholine receptors *Molecules & Cells* (Springer Science & Business Media BV). 2004; 17:242-247.
 47. Steel RGD, Torrie JH. *Principles and Procedures of statistics: A Biometrical Approach*. Mc-Graw Hill Book Co., Inc., New York. 1980, 232-58.
 48. Ayyaz M, Nadeem M, Begum HA. Screening of okra varieties resistance against insect pests under agro climatic conditions of Dera Ismail Khan, Pakistan. *Russian Agricultural Sciences*. 2017; 43(2):149-152.
 49. Naqqash MN, Gökçe A, Bakhsh A, Salim M. Insecticide resistance and its molecular basis in urban insect pests. *Parasitology research*. 2016; 115(4):1363-73.