



E-ISSN: 2320-7078

P-ISSN: 2349-6800

JEZS 2017; 5(5): 876-878

© 2017 JEZS

Received: 24-07-2017

Accepted: 25-08-2017

Jayveer RajpootVeterinary College Mhow,
Indore Madhya Pradesh, India**Supriya Shukla**Veterinary College Mhow,
Indore Madhya Pradesh, India**Gaya Prasad Jatav**Veterinary College Mhow,
Indore Madhya Pradesh, India**UK Garg**Veterinary College Mhow,
Indore Madhya Pradesh, India**Vivek Agrawal**Veterinary College Mhow,
Indore Madhya Pradesh, India

Coproculture study of strongyle infection of goats from Malwa region of Madhya Pradesh

Jayveer Rajpoot, Supriya Shukla, Gaya Prasad Jatav, UK Garg and Vivek Agrawal

Abstract

The coproculture study on strongyle infection of goats was carried out from slaughter house of Mhow, Madhya Pradesh during the period from Nov. 2015 to March 2016. Out of a total of 200 samples, 130 were found positive for strongyle with an overall prevalence rate of 65%. Strongyle positive faecal samples were separately pooled and subjected to coproculture. Thus, larvae obtained were identified up to the level of genera. Upon coproculture of those faecal samples found positive for strongyle type, seven genera of parasites were recognized. *Haemonchus sp.* (32.33%) was the predominant gastrointestinal nematode followed by *Oesophagostomum sp.* (22.51%), *Trichostrongylus sp.* (18.67%), *Cooperia sp.* (15.03%), *Nematodirus sp.* (12.65%), *Ostertagia sp.* (3.34%) and *Bunostomum sp.* (3.12%). *Haemonchus* was the most common type of strongyle encountered in 32.33% goats, while *Bunostomum* was the least recovered from 3.12% goats. The finding of this examination shows that strongyle disease is one of the real issues that could hamper wellbeing and efficiency and there is requirement for outline a program to limit and control strongyle contamination in goats in the study area.

Keywords: Strongyle, Coproculture, Goat, Slaughter house

Introduction

Goat contributes basically to the agrarian economy, particularly in spaces where crop and dairy creating are not spared. India has 140.5 million goats ^[1] which account 16.53 per cent of the total global population. India positions second in goat population in the world. The cost of goat production in the present system of goat rearing is indeed low and the monetary gains through the sale of its products like meat, fiber and skin are high enough to prompt the goat owners to switch over the practices followed over the ages ^[2]. Moreover, goat production adds to about Rs. 2443.3 crore per annum to the national income ^[3]. However, various helminthic diseases are responsible for causing heavy losses due to reduced production, morbidity and mortality. Goats are vulnerable to various parasitic diseases that not only undermine their health but also play a role in lowering the overall production ^[4]. Gastro-intestinal (GI) parasitism in animals is common and one of the noteworthy issues in India. It causes anemia, debilitating, feebleness, oedema, deficiency, and death ^[5]. Since, the work of direct fecal examination alone may not be adequate to offer a correct picture of the prevalence of GI parasites, gives a record of coprological culture has likewise been widely surveyed ^[6, 7, 8]. The Overall prevalence of GI helminth in sheep and goat was 82% from Patiala and its adjoining area and recorded genera were *Trichostrongylus sp.* (78.57%), *Haemonchus sp.* (64.29%), *Strongyloides sp.* (57.14%) and *Oesophagostomum sp.* (42.86%) ^[9]. Prevalence of *Hamonchus* (64.19%), *Trichuris* (35.48%), *Nematodirus* (13.00%), *Trichostrongylus* (4.51%) and *Strongyloides* (3.22%) in goat from Rawalpindi and Islamabad in Pakistan was reported by ^[10]. However, scanty information is available on the prevalence of strongyle in goats in Mhow of M.P. Therefore, the present work has been designed to study various genera of strongyle of goat for developing worm management strategies for control of parasitic diseases.

Materials and methods

Faecal samples were collected randomly from slaughter house Mhow where animals are brought to slaughter from different region of Malwa. Freshly laid or rectal faecal samples were collected in an individually labeled polythene bags from the selected animals ^[11]. The faecal samples were collected daily from slaughter house of Mhow over a period of five months (Nov. 2015-March 2016).

Correspondence**Jayveer Rajpoot**Veterinary College Mhow,
Indore Madhya Pradesh, India

These faecal samples were taken to the laboratory at the earliest for further examination.

The faecal samples positive for strongyle infection were pooled and culture in the laboratory by glass tumbler (300 ml capacity) method. About 75-100 grams of pooled strongyle positive faecal samples was thoroughly mixed with activated charcoal in 3:1 ratio and water sufficient to get a pasty consistency, using pestle mortar and placed in a glass tumbler of 300 ml capacity. After cleaning the inner margin of the tumbler, its mouth was covered with aluminium foil and then it was incubated at 25 to 28°C for 7 days. The culture tumbler was checked once daily in the morning for optimum wetness in the sample. On the 8th day, the tumbler was taken out of the incubator, its aluminium foil was removed and lukewarm water was then filled up till a convex surface was formed at the brim. A glass petridish of 4 inches size was placed on to the mouth of the tumbler with precaution that no air bubbles should be trapped within. The whole preparation was then so inverted that the tumbler stood in the petridish and kept in little slanting position under artificial light. After 4-6 hrs, the water in the petridish was withdrawn and centrifuged at 1000 rpm for 2 minutes. After discarding the supernatant, 10% hot formaldehyde was added to the sediment containing larvae so as to preserve them stretched as per the procedure of [12].

A drop of preserved sediment containing larvae was placed on a glass slide, mixed with a drop of Lugol's iodine or aqueous Safranin and then examined under dry magnifications of the compound microscope after applying a cover slip over the preparation. 100 L3 parasites were counted and identification of strongyle larvae was done with the help of the key and plates provided by [13].

Results and discussion

The larvae of strongyle nematodes recovered from coprocultural examination, were identified up to the level of genera to which they belonged. Out of 200 faecal samples collected from goats, 130 samples (65%) were found positive in strongyle eggs. Higher prevalence of strongyle was also recorded earlier from India and abroad [14, 15]. This may be due to their direct life cycle and the typical grazing habit of small ruminants facilitating the transmission of the parasite.

The percentage of each strongyle species were determined by identifying at least 20 strongyle larvae harvested from the pooled faecal cultures from a slaughter house. Upon coproculture of those faecal samples found positive for strongyle type, seven genera of parasites were recognized. *Haemonchus* sp. (32.33%) was the predominant gastrointestinal nematode followed by *Oesophagostomum* sp. (22.51%), *Trichostrongylus* sp. (18.67%), *Cooperia* sp. (15.03%), *Nematodirus* sp. (12.65%), *Ostertagia* sp. (3.34%) and *Bunostomum* sp. (3.12%) (Table- 1). The contribution of agro ecology and climatic parameters proposed to play an important role in the development and survivability of infective stages of strongyle nematodes on pasture. The collective predominance of *Haemonchus* sp. to be found on coproculture in the present study and well agreement with those reported by [16, 17, 18]. However, highest prevalence of *Haemonchus* sp. might be due to high biotic potential to acquire faster resistant than other nematodes. It's pathogenecity is known to be more important than other nematode. The prevalence of *Bunostomum* sp. was the lowest in our present study. Similar findings were also reported by [19]. However, the present finding is in contrast to those of [20] from Parbhani and [21], from Kenya who reported the predominance of *Trichostrongylus* during rainy season. The pre-parasitic stage preferred cooler months for its development and survival [22]. The above two areas have only to extreme climate i.e. summer and rainy season and therefore, the larvae of *Trichostrongylus* spp. got favorable environment only around rainy season.

The Study of coproculture examination showed the existence of polyparasitism which indicate animals had more than one type of parasitic eggs corroborates with the finding of [23]. Morbidity and loss of production in goat in the study area might be due to interactions and compromization of the immune system of the host by polyparasitism. This can be supported by the fact because most of goats brought to slaughter house suffering from debilitated body conditions. Premature slaughter and rejection of some parts of meat inspection due to parasitic infection responsible for direct losses and indirect losses include the reduction in production [24].

Table 1: Mean generic composition (%) and characteristic features of strongyle nematode larvae recovered from copro-culture.

Genera	Overall mean (%)	Characteristic features of strongyle nematode larvae
<i>Haemonchus</i>	32.33	Slender larva, tail of seath of medium length tapering to a point and often kinked. Tail of seath very short, conical
<i>Oesophagostomum</i>	22.51	Larva of medium size, 32 pentagonal gut cells, lumen of gut wavy
<i>Trichostrongylus</i>	18.67	Small larva bearing one or two tuberosities or indistinctly rounded
<i>Cooperia</i>	15.03	Oval bodies at anterior end of larva. Tail of larva rounded
<i>Nematodirus</i>	12.65	Tail of larva is forked
<i>Ostertagia</i>	3.34	Long, conical, "finger like" tail sheath
<i>Bunostomum</i>	3.12	Very small larva with 16 guts cells

Acknowledgement

The authors are thankful to the Dean, College of Veterinary Science and A.H., Mhow for providing all facilities to carry out the present investigation.

Conclusion

The overall composition of the coprocultural larvae revealed that *Haemonchus* was the predominant nematode, followed by *Oesophagostomum*, *Trichostrongylus*, *Cooperia*, *Nematodirus* *Oestertagia* and *Bunostomum*. These parasites are responsible for causing heavy losses due to reduced production, morbidity and mortality. Since polyparasitism is a common problem therefore strategic deworming of animals using broad

spectrum anti-helminthics are required and for this professional input of veterinarians is desired. Therefore it is concluded that the proper deworming schedule of animals, when conditions are more favourable for development and survival of strongyle larvae on the pasture. Rotational grazing pattern is used at interval and avoid the infected herd with healthy herd. Hence, proper pasture and animal management could improve the control of gastrointestinal nematode infections in goat in small holder farmer.

References

1. Livestock census. 18th All India Livestock Census Dept. of Animal Husbandry & Dairying Ministry of

- Agriculture, GOI, 2007.
2. Bhattacharya NK. Goat Rearing. CIRG, Vijay Printing Press, Mathura (U.P.), 1989.
 3. Prasad J. Goat, Sheep and Pig Production and Management, 2nd Edn. Kalyani Publishers, New Delhi, 2002, 3-11.
 4. Sanyal PK. Gastro-intestinal parasites and small ruminant production in India. In: Sustainable Parasite Control in Small Ruminants. (Editors) L. F. Lejambre and M.R. Knox. ACIAR Proceeding, 1996; 74:109-112.
 5. Lutu WZ. Internal parasitism in milk goats in Kenya. Tropical Animal Health Production. 1983; 16:153-157.
 6. Dubey M, Chaudhry RK. Epidemiology of gastrointestinal nematodes of sheep and goats in and around Jabalpur. X National Congress of Veterinary Parasitology, Jabalpur, 4-6th December, 1998, 75-76.
 7. Singh RK, Sisodia RS, Shukla PC, Pillay AGR. A note on incidence of gastrointestinal parasites in sheep and goats in Madhya Pradesh. Indian Journal of Veterinary Medicine. 1999; 23:130-133.
 8. Kumari S, Sinha SRP, Sinha S, Hoda MZ, Mandal KG, Sharma SK. Incidence of gastrointestinal helminthosis in sheep and goats in Patna (Bihar). Journal of Veterinary Parasitology. 2010; 24(1):97-99.
 9. Kaur H, Kaur D. Prevalence of gastrointestinal parasites in domestic animals of Patiala and its adjoining areas. Journal of Veterinary Parasitology. 2008; 22(2):25-28.
 10. Gadahi JA, Arshed M, Ali JQ, Javaid SB, Shah SI. Prevalence of Gastrointestinal Parasites of Sheep and Goat in and around Rawalpindi and Islamabad, Pakistan Veterinary World. 2009; 2(2):51-53.
 11. Sloss MW, Kemp RL, Zajac AM. Veterinary Clinical Parasitology. 6th Edn. International Book Distributing Co., Lucknow, India, 1994.
 12. Roberts FHS, Sullivan PJO. Methods for egg counts and larval cultures for strongyles infesting the gastrointestinal tract of cattle. Australian Journal of Agricultural Research. 1949; 1:99-103.
 13. Ministry of Agriculture, Fisheries and Food (1971-75) Household food consumption and expenditure 1969-73. Annual Report. National Food Survey Committee. London: HMSO.
 14. Haque M, Jyoti Singh NK, Juyal PD, Singh H, Singh R, Rath SS. Incidence of gastrointestinal parasites in dairy animals of western plains of Punjab. Journal of Veterinary Parasitology. 2011; 25:168-170.
 15. Kumsa B, Tadesse T, Sori T, Dugum R, Hussien B. Helminths of sheep and goats in Central Oromia (Ethiopia) during the dry season. Journal of Animal and Veterinary Advances. 2011; 10:1845-1849.
 16. Parihar MG, Manohar GS, Pathak KML, Kumar D. Prevalence of gastrointestinal parasitosis in goats in and around Ramsar (Ajmer), Rajasthan. VIII National Congress of Veterinary Parasitology, Hissar, 9-11th October, 1996.
 17. Faizal ACM, Rajapakse RPV, Jayasinghe SR, Rupasinghe V. Prevalence of *Eimeria* spp. and gastrointestinal nematodes versus weight gain in treated goats raised in the dry area of Sri Lanka. Small Ruminant Research, 1999; 34:21-25.
 18. Githigia SM, Thamsborg SM, Maingi N, Munyua WK. The epidemiology of gastrointestinal nematodes in Goats in the low potential areas of Thika District, Kenya. Bulletin of Animal Health and Production in Africa. 2005; 53(1):5-12.
 19. Thilakan JN, Sathianesan V. Survey in prevalence of common nematode parasites of domestic ruminants in Kerala. X National Congress of Veterinary Parasitology, Jabalpur, 4-6th, 1998, 69-70.
 20. Kandhara LM, Deshpande PD, Narladkar BW. Round worm infections of ruminants in Marathwada region: Faecal culture and faecal egg counts studies. XI National Congress of Veterinary Parasitology, Bhubaneswar, 4-6th, 2000, 52-53.
 21. Nginyi JM, Duncan JL, Mellor DJ, Stear MJ, Wanyangu SW, Bair RK *et al.* Epidemiology of parasitic gastrointestinal nematode infections of ruminants on small holder farms in Central Kenya. Research in Veterinary Science. 2001; 70:33-39.
 22. Yadav CL. Agro-climatic influence on parasitic diseases of sheep and goats. Pashudhan, 2000; 15:16.
 23. Haileleul N. Study on prevalence of GIT helminth of small ruminants in and around Woilyata Soddo, Southern Ethiopia. DVM thesis, FVM, AAU, Debre Zeit. Ethiopia, 2002.
 24. Gonzalez R, Gonzalez A. Alternative for the Control of Gastrointestinal Nematode in Sheep. Conf. EEPF Matanzas, Cuba. 2004.