

Journal of Entomology and Zoology Studies

Journal of Entomology and Zoology Studies

Available online at www.entomoljournal.com

E-ISSN: 2320-7078 P-ISSN: 2349-6800

Accepted: 25-10-2017

JEZS 2017; 5(6): 2408-2410 © 2017 JEZS Received: 22-09-2017

Mark Cooper

University of South Africa, Pretoria, South Africa

Re-assessment of rensch's rule in Centrobolus

Mark Cooper

Abstract

Sexual size dimorphism (SSD) was investigated in the helminthomorph genus *Centrobolus* (=Chersastus). Width, length and mass were used to compare the interspecific variation for 20 species. Interspecific variation in volume was calculated in these species and an allometric coefficient of 0.7 found. The allometric equation generated for the genus also the inverse of Rensch's rule as there was a positive correlation between SSD and body size (R=0.70485; P=0.00109; n=18 spp) in this diplopod genus. This SSD is thought to have evolved through intersexual competition driven by sexual selection.

Keywords: allometry, centrobolus, rensch's, spirobolida

1. Introduction

Diplopoda are underrepresented in allometric analyses of SSD, although sexual differences are known in body mass, length, width and leg dimensions of over half the taxa studied [1-3]. Size differences occur with factors such as color, sexes, species, urbanisation and water relations [4-8]. Diplopoda resemble the majority of invertebrates where SSD is reversed [9]. SSD has consequences for the outcome of sexual encounters in diplopod mating [10-14]. The detection of a relationship between body size and SSD is known by Rensch's rule [15-16]. Rensch's rule may be explained by sexual selection [17-21]. The macro-evolutionary pattern is unresolved in Diplopoda [22].

SSD has been studied in the Spirobolida genus *Centrobolus* and the relationship between the log of male and female volumes against each other used to Corroborate Rensch's rule [15-16]. In the present study, SSD in the genus *Centrobolus* is re-investigated and re-tested. The hypothesis was SSD is negatively correlated with mean body sizes ^[23].

2. Material and Methods

Three factors were measured from the sample of Centrobolus: (1) body length (mm) by placing individuals alongside a plastic rule (calibrated in mm); (2) width (mm) with Vernier calipers; and (3) mass (accurate to 0.01 g). C. inscriptus (Mtunzini), and C. ruber (Anerley, Port Shepstone) were collected in South Africa (1996) while more data was obtained from Attems [24]. Their helminthomorph size was calculated as shape volume based on the formula for a cylinder (l, π, r^2) where l was body length and r half of the width i. e. radius. SSD was estimated as mean female volume divided by mean male volume and converted into a SSD [25] **SSD** index subtracting 1 was plotted versus (http://www.socscistatistics.com). Allometry for SSD was based on a general allometric model where male size = α (female size) β [26]. A linear regression was calculated at http://www.socscistatistics.com/tests/regression/Default.aspx and Spearman's Rho coefficients calculated at http://www.socscistatistics.com/tests/spearman/Default2.aspx. Male and female width and volume have already been compared by Cooper [23].

3. Results

The allometric equation for *Centrobolus* was (1) $\hat{y}=0.00051X$ - 0.01071. SSD ranged from 0.63 – 2.89 (1.55 ± 0.63; n = 20) and was significantly positively correlated (R = 0.70485; P = 0.00109; n = 18 spp.) with volume ranging from 284 – 2683 mm³ (1097.89 ± 638.06; 18) (Table 1).

Correspondence
Mark Cooper
University of South Africa,
Pretoria. South Africa

Table 1: Body size, sexual size dimorphism (SSD) and SSD index for *Centrobolus* Cook, 1897.

| Species | Body size (mm ³) | SSD | SSD-1 | Sample size (n) |
|------------------|---------------------------------|------|-------|--------------------|
| C. albitarsus | 952 | 2.89 | 1.89 | 1 |
| C. decoratus | 557 | 0.63 | -0.37 | 1 |
| C. digrammus | 522 | 1.01 | 0.01 | 6 |
| C. dubius | 1210 | 1.35 | 0.35 | 1 |
| C. fulgidus | 1518 | 1.65 | 0.65 | 11 |
| C. fulgidus | 1.65 g | 1.25 | 0.25 | 22 |
| C. immaculatus | 1580 | 2.72 | 1.72 | 1 |
| C. inscriptus | 2.38 g | 1.22 | 0.22 | 176 |
| C. inyanganus | 775 | 1.44 | 0.44 | 1 |
| C. lawrencei | 962 | 1.57 | 0.57 | 1 |
| C. lugubris | 2046 | 2.18 | 1.18 | 1 |
| C. promontories | 284 | 0.69 | -0.31 | 1 |
| C. pusillus | 756 | 2.08 | 1.08 | 1 |
| C. ruber | 1.64 g | 1.62 | 0.62 | 36 |
| C. rugulosus | 1666 | 1.97 | 0.97 | 1 |
| C. sagatinus | 1659 | 1.27 | 0.27 | 1 |
| C. silvanus | 749 | 1.13 | 0.13 | 1 |
| C. titanophilus | 393 | 1.15 | 0.15 | 1 |
| C. transvaalicus | 669 | 1.26 | 0.26 | 1 |
| C. tricolor | 781 | 1.10 | 0.10 | 1 |
| C. vastus | 2683 | 1.81 | 0.81 | 1 |

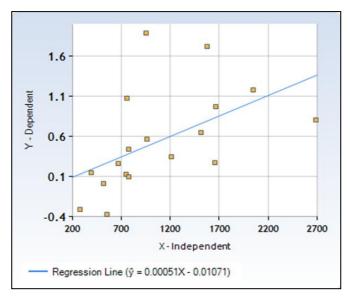


Fig 1: Regression showing the relationship between sexual size dimorphism and body mass for *Centrobolus* (N = 18)

4. Discussion

The significant differences between males and females have already been dealt with for this genus [23]. The regression of SSD on body size (Figure 1) of the data for SSD and body size (Table 1) indicates a positive regression to reject Rensch's rule in *Centrobolus*. Although the null hypothesis is accepted and the rule rejected the inverse relationship is true [26-32]. This was seen in the SSD increases with body size. The same result was obtained when the analysis was run with/without C. fulgidus suggesting there is little or no difference when mass and volume are used to calculate SSD. If Rensch's rule implies only hypo-allometric functions then it has been corroborated [23]. However, if Rensch's rule implies a negative relationship between SSD and body size then it must be rejected here. Both scenarios have been dealt with and the proximate cause of the relationship between SSD and body size may be suggested for *Centrobolus annulatus* and *C*. inscriptus in sympatry [33].

5. Conclusion

The inverse of Rensch's rule was found in *Centrobolus* based on the positive relationship between SSD and body size.

6. Acknowledgements

G. Cooper provided computer for analysis.

7. References

- 1. Hopkin SP, Read HJ. The Biology of Millipedes. Oxford University Press, U. K. 1992, 246.
- Ilić BS, Mitić BM, Makarov SE. Sexual dimorphism in Apfelbeckia insculpta (L. Koch, 1867) (Myriapoda: Diplopoda: Callipodida). Archives of Biological Sciences. 2017; 69:23-33.
- 3. Wilson HM, Anderson LI. Morphology and taxonomy of Paleozoic millipedes (Diplopoda: Chilognatha: Archipolypoda) from Scotland. Journal of Paleontology. 2004; 78(1):169-184.
- Bhakat S. Comparative water relations of some tropical millipedes. Kragujevac Journal of Science. 2014; 36:185-194.
- Bogyó D, Magura T, Simon E, Tóthmérész B. Millipede (Diplopoda) assemblages alter drastically by urbanisation. Landscape and Urban Planning. 2015; 133:118-126.
- Calligaris IB, Boccardo L, Sanches MR, Fontanetti CS. Morphometric Analysis of a Population of Diplopods of the Genus Rhinocricus Karsch, 1881. Folia Biologica (Praha). 2005; 51:40-46.
- 7. David J–F. Size criteria for the distinction between Cylindroiulus londinensis (Leach) and Cylindroiulus caeruleocinctus (Wood) (Diplopoda: Julidae). Journal of Natural History. 1995; 29:983-991.
- 8. Enghoff H. The size of a millipede. In: Meyer E, Thaler K, Schedl W (eds.) Advances in Myriapodology. Berichte des naturwissenschaftlich-medizinischen Vereins in Innsbruck, Supplement. 1992; 10:47-56.
- 9. Mori E, Mazza G, Lovari S. Sexual Dimorphism. In: Encyclopedia of Animal Cognition and Behavior (J. Vonk, and T. Shakelford, Eds). Springer International Publishing, Switzerland. 2017, 1-7.
- 10. Adolph SC, Geber MA. Mate-Guarding, Mating Success and Body Size in the Tropical Millipede 'Nyssodesmus Pythos' (Peters) (Polydesmida: Platyrhacidae). The Southwestern Naturalist. 1995; 40(1):56-61.
- Rowe M. Copulation, mating system and sexual dimorphism in an Australian millipede, Cladethosoma clarum. Australian Journal of Zoology. 2010; 58(2):127-132
- 12. Cooper MI Size matters in myriapod copulation. Journal of Entomology and Zoology Studies. 2017; 5(2):207-208.
- Tanabe T, Sota T. Complex Copulatory Behavior and the Proximate Effect of Genital and Body Size Differences on Mechanical Reproductive Isolation in the Millipede Genus Parafontaria. The American Naturalist. 2008; 171(5):692-699.
- 14. Cooper MI. The affect of female body width on copulation duration in *Centrobolus inscriptus* (Attems). Journal of Entomology and Zoology Studies. 2017; 5(1):732-733.
- 15. Rensch B. Evolution above the Species Level. Columbia, New York. 1947, 419.
- Rensch B. Die Abhängigkeit der relativen Sexualdifferenz von der Körpergrösse. Bonn Zoological Bulletin. 1950; 1:58-69.

- 17. Andresson M, Wallander J. Animal behaviour: Relative size in the mating game. Nature. 2004; 431:139-141.
- 18. Bonduriansky R. Sexual selection and allometry: a critical reappraisal of the evidence and ideas. Evolution. 2007; 61(4):838-849.
- 19. Clutton-Brock TH, Harvey PH, Rudder B. Sexual dimorphism, socionomic sex ratio and body weight in primates. Nature. 1977; 269:797-800.
- 20. Dale J, Dunn PO, Figuerola J, Lislevand T, Székely T, Whittingham LA. Sexual selection explains Rensch's rule of allometry for sexual size dimorphism. Proceedings of the Royal Society B. 2007; 274:2971-2979.
- 21. Gaulin SJC, Sailer LD. Sexual dimorphism in weight among the Primates: the relative impact of allometry and sexual selection. International Journal of Primatology. 1984; 5(6):515-535.
- 22. Cooper MI Sexual size dimorphism and corroboration of Rensch's rule in *Chersastus* millipedes. Journal of Entomology and Zoology Studies. 2014; 2(6):264-266.
- 23. Cooper MI. Sexual size dimorphism and corroboration of Rensch's rule in *Chersastus* millipedes. Journal of Entomology and Zoology Studies. 2014; 2(6):264-266.
- 24. Attems C. The Myriapoda of South Africa. Annals of the South African Museum. 1928; 26:1-431.
- 25. Lovich JE, Gibbons JW. A review of techniques for quantifying sexual size dimorphism. Growth Development and Aging. 1992; 56:269-281.
- Leutenegger W. Scaling of sexual dimorphism in body size and breeding system in primates. Nature. 1978; 272:610-611.
- 27. Leutenegger W. Scaling of sexual dimorphism in body size and breeding system in primates. Nature. 1978; 272:610-611.
- 28. Herczeg G, Gonda A, Merilä J. Rensch's rule inverted female-driven gigantism in nine-spined stickleback *Pungitius pungitius*. Journal of Animal Ecology. 2010; 79:581-588.
- 29. Liao WB. Evolution of sexual size dimorphism in a frog obeys the inverse of Rensch's rule. Evolutionary Biology. 2013; 40:493-499.
- 30. Lu D, Zhou CQ, Liao WB. Pattern of sexual size dimorphism supports the inverse Rensch's rule in two frog species. Animal Biology. 2014; 64:87-95.
- 31. Liao WB, Liu WC, Merilä J. Andrew meets Rensch: sexual size dimorphism and the inverse of Rensch's rule in Andrew's toad (*Bufo andrewsi*). Oecologia. 2015; 177:389-399.
- 32. Cooper MI. The relative sexual size dimorphism of Centrobolus inscriptus (Attems) compared to 18 congenerics. Journal of Entomology and Zoology Studies. 2016; 4(6):504-505.
- 33. Cooper MI. Sex ratios, mating frequencies and relative abundance of sympatric millipedes in the genus Chersastus. Arthropods. 2014; 3(4):174-176.