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Potential of *Mucuna pruriens* (L.) DC. for insecticidal activity, insect repellency and brine shrimp lethality tests under the laboratory conditions

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Abstract

Aerial part, fruit and root extracts of *Mucuna pruriens* (L.) DC. were obtained with petroleum ether, chloroform and methanol and evaluated for their insecticidal and insect repellent activity against the adult beetles of *Tribolium castaneum* (Hbst.) and cytotoxic activity against *Artemia salina* L. nauplii. The chloroform extract of fruit found most effective with LD₅₀ 0.534mg cm⁻² followed by the petroleum ether and methanol extracts of root with LD₅₀ 0.814 and 1.056mg cm⁻² after 48 hours of exposure. Petroleum ether and chloroform extracts of the fruit and methanol extract of the aerial part showed repellent activity against the adult beetles at 5% level of significance ($P < 0.05$). The same extracts responded through (Cytotoxicity) brine shrimp lethality assay while the methanol extract of aerial part found most effective (LC₅₀ 12.793ppm) followed by the petroleum ether extract of root and chloroform extract of fruit with LC₅₀ 24.414 and 25.208ppm after 30h of exposure.

Keywords: *Mucuna pruriens*, dose mortality, repellency, *Tribolium castaneum*, brine shrimp lethality, *Artemia salina*

Introduction

Mucuna pruriens (L.) DC. of the family Fabaceae is the native in the tropical Africa, Asia and West Indies. It is a climbing annual legume known as Cowhage, velvet-bean or Bengal velvet-bean that can be up to 6-18m long and contains a taproot with numerous 7-10m long, lateral roots. The stems are slender and slightly pubescent and the leaves are generally slightly pubescent, alternate, trifoliate with rhomboid ovate, 5-15cm long × 3-12cm broad leaflets [1-2]. It can grow on a wide range of soils, from sands to clays but thrives on well-drained, light textured soils of appreciable acidity [3-4]. *M. pruriens* finds importance as food, feed, cover crop and fodder and is extensively cultivated worldwide [5-8]. It is considered a viable source of dietary proteins [9-10] due to its high protein concentration (23-35%) in addition to its digestibility, which is comparable to that of other pulses such as soybean, rice bean, and lima bean [11]. *M. pruriens* is a popular Indian medicinal plant, which has long been used in traditional Ayurvedic Indian medicine, for diseases including Parkinsonism [12]. The present investigation was carried out to find out the potential of its insecticidal and insect repellent activity against the red flour beetle, *Tribolium castaneum* (Hbst.), and lethality against the brine shrimp, *Artemia salina* L. nauplii. The red flour beetle is reddish-brown in color and its antennae end in a three-segmented club [13]. Although small beetles, about ¼ of an inch long, the adults are long-lived and may live for more than three years [14], and thus became a suitable lab insect. The *A. salina* belongs to a genus of very primordial crustacean (crawfish-crayfish) the *Anostraca* (Fairy Shrimps). Crawfish of this genus just has a divided exoskeleton made of chitin enhanced protein, no usual crust of chitin (escutcheon) as the crawfish has. There are many species within the genus of *Anostraca*, but the *A. salina* is very nice to grow, and since the rate of successful hatches is very high this species is used as a test agent in the toxicology laboratory.

Materials and Methods

Collection and preparation of test materials: *M. pruriens* was collected from the University of Rajshahi Campus and identified by the Department of Botany, University of Rajshahi,

where a voucher specimen is kept in the herbarium. The plants were chopped into small pieces, dried under shade and powdered using a hand grinder, weighed and placed in separate conical flasks to add Pet. ether, CHCl_3 and MeOH (Merck, Germany) (100gm \times 300ml \times 2times) for 48h. Filtration was done by Whatman filter paper (made in USA) at the 24h interval in the respective flasks followed by evaporation until the extracts were left. The extracts were then removed to glass vials and preserved in a refrigerator at 4°C with proper labeling.

Collection and culture of test insect: Adults of *T. castaneum* were reared in glass beakers (500ml) in a standard mixture of whole-wheat flour with powdered dry yeast (19:1) in an incubator at 30 \pm 0.5°C without light and humidity control for a continuous supply of adults during experimentation.

Dose-mortality test: The dose-mortality responses of *M. Pruriens* were observed by surface film method. The concentrations used were 2.038, 1.529, 1.019, 0.510 and 0.254mg cm⁻² for Pet. ether extract of fruit followed by 2.548, 2.038, 1.529, 1.019, 0.510 and 0.254mg cm⁻² and 3.567, 3.057, 2.548, 2.038, 1.529, and 1.019mg cm⁻² for CHCl_3 and CH_3OH extracts; 4.080, 3.567, 3.057, 2.548, and 2.038mg cm⁻² for Pet. ether extracts of aerial part as well as 2.548, 2.038, 1.529, 1.019, 0.510 and 0.250mg cm⁻²; and 2.548, 2.038, 1.529, 1.019, 0.510 and 0.254mg cm⁻² for Pet. ether and CH_3OH extracts of root against *T. castaneum* in the dose mortality experiments. Each of the doses were diluted in 1ml of solvent, poured into Petri dishes and allowed to dry out. Ten adult beetles were released in each Petri dish, and the experiment of all the doses for each of the extracts were replicated in a thrice. The mortality was assessed for 12h, 24h, 36h, and 48h of exposure.

Statistical Analysis: The mortality (%) was corrected using Abbott's formula [15]:

$$P_r = P_o - P_c / 100 - P_c \times 100$$

Where, P_r = Corrected mortality (%), P_o = Observed mortality (%), P_c = Control mortality (%). The data were then subjected to probit analysis according to Finney [16] and Busvine [17] using a software developed at the University of Newcastle upon Tyne, UK.

Repellent Activity: The repellent activity test was adopted from the method (No. 3) of McDonald *et al.* [18] with some

modifications. Half filter paper discs (Whatman No. 40, 9cm diam.) were treated with the selected doses of 0.629, 0.314, 0.157, 0.0786, 0.0393mg cm⁻² for all extracts and were then attached lengthwise, edge-to-edge, to a control half-disc with adhesive tape and placed in the Petri dishes. The orientation was changed in the two remaining replicates to avoid the effects of any external directional stimulus affecting the distribution of the test insects. Ten adult insects were released in the middle of each of the filter paper circles.

Each concentration against of the solvents was tested for five times. Insects that settled on each of the non-treated half of the filter paper discs were counted after 1h and then observed repeatedly at hourly intervals for five hours. The average of the counts was converted to percent repulsion (*PR*) using the formula of Talukder and Howse^[19-20]: $PR = (N_c - 5) \times 20$, where, N_c is the percentage of insects on the untreated half of the disc.

Brine shrimp nauplii lethality test: Brine shrimp cycts were purchased from Kalabagan, Dhaka and kept in aerated seawater at room (25-30°C) temperature and took 30-48h to give nauplii. The series of concentration were for Pet. ether extracts of the fruit were: 400, 200, 100, 50 and 25ppm; followed by 100, 50, 25, 12.5 and 6.3ppm; and 100, 50, 25, 12.5 and 6.3ppm for CH_3OH and CHCl_3 extracts as well as 800,400, 200,100 and 50ppm; 400,100 and 25ppm; and 200, 100, 50, 25 and 12.5ppm for Pet. ether, CHCl_3 and CH_3OH extracts of aerial parts and for Pet. ether extracts of root were 50, 25, 12.5 and 6.3ppm; followed by 200, 100 and 50ppm; and 50, 25 and 12.5ppm for CHCl_3 and CH_3OH extracts were selected respectively. Ten freshly hatched nauplii were added to each of the test tubes with different concentrations mentioned earlier and observed mortality after 6, 12, 18, 24 and 30h of exposures. The data were then subjected to probit analysis.

Results and Discussion

Dose mortality effects: The dose mortality assay of Pet.E., CHCl_3 and CH_3OH extracts are represented in Table 1. All the extracts found effective against *T. Castaneum* adults. The CHCl_3 extracts of fruit found most effective showing LD₅₀ 0.933,0.702,0.579 and 0.534 mg cm⁻² followed by the Pet. ether extract of root with LD₅₀ 2.845,1.820,1.048 and 0.814mg cm⁻² and methanol extract of root with LD₅₀ 3.275, 2.259, 1.844 and 1.056 mg cm⁻² for 12, 24, 36 and 48h of exposures respectively.

Table 1: LD₅₀ values of the test extracts of *M. pruriens* established through residual film assay against *T. castaneum* adults.

Test plant	Plant parts used	Solvents	LD ₅₀ value (mg cm ⁻²)			
			Duration of exposures			
			12h	24h	36h	48h
<i>M. pruriens</i>	Fruit	Pet.E.	8.896	2.428	1.709	1.031
		CHCl_3	0.933*	0.702	0.579	0.534*
		CH_3OH	-	28.982	3.997	2.869
	Aerial part	Pet. E.	4.069	4.842*	3.436	3.210*
		Pet.E.	2.845*	1.820	1.048	0.814
	Root	CH_3OH	3.275	2.259	1.844	1.056*

* Variance has been adjusted for heterogeneity

Repellent effects: The extracts of fruit obtained in Pet. ether; and CHCl_3 and CH_3OH extract of aerial part offered promising repellent activity at 5% and 1% level of

significance ($P < 0.05$ and $P < 0.01$) respectively while the other extracts didn't show any significant repellency (Tables 2 and 3).

Table 2: ANOVA results of the repellency against *T. castaneum* by the Pet. ether, CHCl₃ and CH₃OH extracts of *M. pruriens* (fruit, aerial part and root).

<i>M. pruriens</i> plant parts	Solvents used for extraction	Sources of variation			F-ratio with level of significance		P- value	
		Between doses	Between time interval	Error	Between doses	Between time interval	Between doses	Between time interval
Fruit	Pet. ether	4	4	16	19.358*	1.477	5.6E-06	0.255
	CHCl ₃	4	4	16	8.972*	0.160	0.0005	0.955
	CH ₃ OH	4	4	16	2.819	2.593	0.060	0.076
Aerial part	Pet. ether	4	4	16	1.107	1.303	0.387	0.311
	CHCl ₃	4	4	16	6.201	2.340	0.003	0.099
	CH ₃ OH	4	4	16	10.751*	2.461	0.0002	0.087
Root	Pet. ether	4	4	16	3.281	3.075	0.038	0.047
	CHCl ₃	4	4	16	1.839	1.429	0.1707	0.270
	CH ₃ OH	4	4	16	2.928	4.586	0.054	0.012

* = Significant at 5% level ($P < 0.05$)**Table 3:** Repellent effect of Pet. ether and CHCl₃ extract of fruit and CH₃OH extract of aerial part of *M. Pruriens* against *T. castaneum*.

Solvents	Between doses (df=4)		Between time interval	
	F- values	Level of significance	F- values	Level of significance
Pet. ether (Fruit)	19.358*	$P < 0.05$	0.865	-
CHCl ₃ (Fruit)	8.972*	$P < 0.05$	0.705	-
CH ₃ OH (Aerial part)	10.751*	$P < 0.05$	2.093	-

** = Significant at 1% level ($P < 0.01$) * = Significant at 5% level ($P < 0.05$); (-) = Not significant at any level.

Brine shrimp lethality effect: The brine shrimp lethality for Pet.E., CHCl₃ and MeOH extracts of *M. pruriens* represented in Table 4. The highest lethality was observed in the CH₃OH

extract of aerial part with LC₅₀ value 12.793ppm followed by the Pet. ether extract of root with LC₅₀ 24.414ppm both after 30h of exposure against brine shrimp nauplii.

Table 4: LC₅₀ values of the test extracts of *M. pruriens* established through brine shrimp lethality assay against *A. salina* nauplii.

Test plant	Plant part Used	Solvents	LC ₅₀ value (ppm)				
			Duration of exposure				
			6h	12h	18h	24h	30h
<i>M. pruriens</i>	Fruit	Pet. ether	217.120	120.414	69.093	56.085	44.139
		CHCl ₃	80.511	59.947	40.234	31.767	25.208
		CH ₃ OH	-	17117.130	218.850	144.431	77.675
	Aerial Part	Pet. ether	3297.292	2110.736	2107.780	1355.386	321.291
		CHCl ₃	-	247.889	165.053	58.738	34.273
		CH ₃ OH	60.486	33.064	25.936	17.256	12.793
	Root	Pet. ether	175.297	72.111	51.943	33.436	24.414
		CHCl ₃	108.639	45.254	43.905	43.910	34.111
		CH ₃ OH	66.647	40.977	35.020	23.738	19.527

Discussion

These findings receive support from previous researchers' achievements. All parts of *Mucuna* plant are known to possess high medicinal value [21]. Its extracts have been long used in tribal communities as a toxin antagonist for various snakebites. Aqueous extracts of *M. pruriens* seeds possess compounds, which inhibit the activity of cobra and krait venoms. About 0.16mg and 0.19mg of *M. pruriens* seed extracts were able to completely neutralize the lethal activity of cobra and krait venom respectively [19]. *Mucuna* pod hairs are blended with honey and are used as vermifuge. The paste prepared from pod hairs are also used as a stimulant and mild [20]. *M. pruriens* has been found to contain L-DOPA, that converts into dopamine which is a potent neurotransmitter precursor that is believed, in part, to be responsible for the toxicity of the *Mucuna* seeds [22]. A study was conducted to develop suitable method(s) for extraction of L-DOPA from the powdered seeds of *M. pruriens* using different solvents and conditions [23]. Rajeshwar *et al.* [24] demonstrated that the methanol extract of *M. pruriens* seeds has significant *in vitro* anti-oxidant activity, and there are also indications that methanol extracts of *M. pruriens* may be a potential source of natural anti-oxidants and anti-microbial agents. Its anti-venom activities have been investigated [25] and its anti-helminthic

activity has also been demonstrated by Jalalpure [26]. *M. pruriens* has also been shown to be neuroprotective [27] and has demonstrated as analgesic and anti-inflammatory agents [28]. The Antiparkinsonian and Anti dyskinetic mechanisms of *M. pruriens* have also been reported [29]. In the Ayurvedic system of medicine, *M. pruriens* was used for the management of male infertility, nervous disorders and also as an aphrodisiac [30-31]. Fruit, aerial part and root extracts of *M. pruriens* of Pet. ether, CHCl₃ and CH₃OH were found effective against the brine shrimp nauplii in different hour of exposure. Patoary *et al.* [32] conducted a research where he reported the CHCl₃ and CH₃OH extracts of the seed coat and seed kernel of *M. pruriens* were tested against the brine shrimp, *Artemia franciscana* nauplii for mortality at 24h post exposure. All the test extracts were found to be effective. A review study conducted by Yadav *et al.* [33] established the plant *M. pruriens* as a herbal drug. It has been shown that its seeds are potentially of substantial medicinal importance. In the ancient Indian medical system, Ayurveda, *M. pruriens* used traditionally to treat Parkinson's disease. *M. pruriens* has also been shown to have antiparkinson and neuroprotective effects, which may be related to its antioxidant activity and used for the management of male infertility, nervous disorders, and also as an aphrodisiac. Some authors reported

the repellent effects of some well-known plants such as neem, turmeric, etc. Leaf of *Withania somnifera* repelled *Ryzopertha dominica* ^[34].

Conclusion

Dose-mortality, insect repellent activity and brine shrimp lethality tests revealed that this renowned medicinal plant *M. pruriens* has potentials to be a potent tool in the global pest control strategy. Thus, comprehensive phytochemical analyses of the test plant for its biologically active potentials, as well as the pharmacological studies of the active ingredients are very much to be solicited for their effective use in the future pest control and pharmaceutical endeavors.

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References

1. US Forest Service. *Mucuna pruriens* (L.) DC. Pacific Island Ecosystems at Risk (PIER), 2011; http://www.hear.org/pier/species/mucuna_pruriens.htm
2. Wulijarni-Soetjijto N, Maligalig RF. *Mucunapruriens* (L.) DC. cv. group *Utilis*. Record from Proseabase. Faridah Hanum, I; van der Maesen, L.J.G. (Editors). PROSEA (Plant Resources of South-East Asia) Foundation, Bogor, Indonesia, 1997; http://proseanet.org/prosea/e-prosea_detail.php?frt=&id=63
3. FAO. Grassland Index. A searchable catalogue of grass and forage legumes, 2011.
4. Maasdorp B, Jiri O, Temba E. Contrasting adoption, management, productivity and utilization of *Mucuna* in two different smallholder farming systems in Zimbabwe. In: Tropical Legumes for Sustainable Farming Systems in Southern Africa and Australia (eds- Whitbread, A.M. and Pengelly, B.C.). ACIAR Proceedings. 2004; 115:154-163.
5. Siddhuraju P, Vijayakumari K, Janardhanan K. Chemical composition and protein quality of the little known legume velvet bean (*Mucuna pruriens* (L.) DC.) J of Agriculture and Food Chem. 1996; 44:2636-2641.
6. Siddhuraju P, Becker K, Makkar HP. Studies on the nutritional composition and antinutritional factors of three different germplasm seed materials of an under-utilized tropical legume, *Mucuna pruriens* var. *utilis*. J of Agriculture and Food Chem. 2000; 48:6048-6060.
7. Siddhuraju P, Becker K. Rapid reversed-phase high performance liquid chromatographic method for the quantification of L-Dopa (L-3, 4-dihydroxyphenylalanine), non-methylated and methylated tetra hydro isoquinoline compounds from *Mucuna* beans. J of Agriculture and Food Chem. 2001; 72:389-394.
8. St. Laurent L, Livesey J, Arnason JT, Bruneau A. Variation in L-dopa concentration in accessions of *Mucuna pruriens* (L.) DC var. *utilis* (Wall. ex Wight) Baker ex Burck. and in *Mucuna brachycarpa* Rech. In: Flores M, Eilitta M, Myhrman R, Carew LB, Carsky RJ, (Eds.). Food and Feed from *Mucuna*: Current Uses and the Way Forward. Tegucigalpa, Honduras: CIDICCO, 2002.
9. Janardhanan K, Gurumoorthi P, Pugalenti M. Nutritional potential of five accessions of a South Indian tribal pulse, *Mucuna pruriens* var. *utilis*. Part I. The effect of processing methods on the contents of L-Dopa phytic acid, and oligosaccharides. J. of Tropical and Subtropical Agro-eco. 2003; 1:141-152.
10. Pugalenti M, Vadivel V, Siddhuraju P. Alternative food/feed perspectives of an under-utilized legume *Mucuna pruriens* Utilis-A Review. Linn. J of Plant Foods and Human Nutrition. 2005; 60:201-218.
11. Gurumoorthi P, Pugalenti M, Janardhanan K. Nutritional potential of five accessions of a south Indian tribal pulse *Mucuna pruriens* var. *utilis*; II Investigation on total free phenolics, tannins, trypsin and chymotrypsin inhibitors, phytohaemagglutinins, and *in vitro* protein digestibility. Tropical, Subtropical and Agroeco. 2003; 1:153-158.
12. Sathiyarayanan L, Arulmozhi S. *Mucuna pruriens*, A comprehensive review. Pharmacognosy Revised. 2007; 1:157-162.
13. Bousquet Y. Beetles associated with stored products in Canada. Canadian Government Publishing Centre, Ottawa, 1990, 189-192.
14. Walter VE. Stored product pest. Franzak and Foster Co., Cleveland, OH, 1990, 526-529.
15. Abbott WS. A method of computing the effectiveness of an insecticide. J of Eco Ent. 1925; 18(2):265-267.
16. Finney DJ. Probit analysis: a statistical treatment of the sigmoid response curve. Cambridge University Press, London, 1947, 333.
17. Busvine JR. A critical review of the techniques for testing insecticides. Commonwealth Agricultural Bureaux, London, 1971, 345.
18. McDonald LL, Guy RH, Speirs RD. Preliminary evaluation of new candidate materials as toxicants, repellents and attractants against stored-product insects. Marketing Research Report No. 882. Agricultural Research Service, US Department of Agriculture, Washington, DC, 1970.
19. Talukder FA, Howse PE. Deterrent and insecticidal effects of extracts of Pithraj, *Aphanamixis polystachya* (Meliaceae), against *Tribolium castaneum* in storage. J of Chem Eco. 1993; 19(11):2463-2471.
20. Talukder FA, Howse PE. Evaluation of *Aphanamixis polystachya* as a source of repellents, antifeedants, toxicants and protectants in storage against *Tribolium castaneum* (Herbst), J of Stored Product Res. 1995; 31(1):55-61.
21. Metcalf CL, Flint WP. Destructive and useful insects. McGraw-Hill Publishing, New York, 1962, 1087.
22. Zyromska-Rudzka H. Abundance and emigrations of *Tribolium* in a laboratory model. Ekologia Polska A.
23. Meenatchisundaram S, Michael A. Antitoxin activity of *Mucuna pruriens* aqueous extract against Cobra and Krait venom by *in vivo* and *in vitro* method. Ind. J of Experimental Bio. 2010; 48:865-878.
24. Rajeshwar Y, Gupta M, Mazumder UK. *In vitro* Lipid Peroxidation and Antimicrobial Activity of *Mucuna pruriens* Seeds. Iranian J of Pharmacology and Therapeutics. 2005; 4:32-35.
25. Guerranti R, Aguiyi JC, Errico E, Pagani R, Marinello E. Effects of *Mucuna pruriens* extract on activation of

- prothrombin by *Echiscarinatus* venom. J of Ethnopharm. 2001; 75:175-180.
26. Jalalpure SS, Alagawadi KR, Mahajanashell CS. *In vitro* antihelmintic property of various seed oils against *Pheritima posthuma*. Indian J of Pharmaceutical Sci. 2007; 69:158-160.
 27. Misra L, Wagner H. Extraction of bioactive principles from *Mucuna pruriens* seeds. Indian J of Biochem and Bioph. 2007; 44:56-60.
 28. Hishika R, Shastry S, Shinde S, Guptal SS. Preliminary phytochemical and anti-inflammatory activity of seeds of *Mucuna pruriens*. Indian J of pharm. 1981; 13(1):97-98.
 29. Lieu CA, Kunselman AR, Manyam BV, Venkiteswaran K, Subramanian T. A water extract of *Mucuna pruriens* provides long-term amelioration of Parkinsonism with reduced risk for dyskinesias. Parkinsonism and Related Disorders. 2010; 16(7):458-465.
 30. Ravishankara MN, Shrivastava N, Jayathirtha MG, Padh H, Rajani MM. A sensitive High-performance thin layer chromatographic method for the estimation of diospyrin, a tumour inhibitory agent from stem bark of *Diospyros Montana* Roxb. J of Chromatography B. 2000; 744:257-262.
 31. Mugendia JB, Njagi EM. Effects of Processing *Mucuna* Bean (*Mucuna pruriens* L.) on Protein Quality and Antinutrients Content, Conference on International Research on Food Security, Natural Resource Management and Rural Development. 2010; 9:14-16.
 32. Patoary R, Mondal OA, Islam W, Khan AR. Cytotoxicity of certain seed extracts against *Artemia fransiscana*. Bangladesh J Zool. 2014; 42(2):133-139.
 33. Yadav MK, Upadhyay P, Purohit S, Pandey BL, Shah H. Phytochemistry and pharmacological activity of *Mucuna pruriens*: A review. J of Green Pharm. 2017; 11(2):69-73.
 34. El-Lakwah FA, Khaled OM, Khattab MM, Abdel-Rahman TA. Effectiveness of some plants extract and powder against the lesser grain borer (*Rhizopertha dominica* F.). Annals of Agri. Sci. 1997; 35(1):567-578.