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Genetic diversity analysis using microsatellite marker in *Labeo gonius* (Hamilton, 1822) from two reservoirs of Uttarakhand

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Abstract

The present study deals with the assessment of genetic diversity using microsatellite marker in the fish *Labeo gonius* from Nanak Sagar and Dhaura reservoirs of Uttarakhand having different morpho-edaphic features and self-recruiting populations of this fish. These reservoirs are distantly located and distinctly separated without any connection having negligible possibility of gene exchange with each other. Total 20 microsatellite primers were designed by using software Primer-BLAST and Primer-3 and all the designed microsatellite primers were screened in all 100 DNA samples of fish collected from both the reservoirs. Out of 20 microsatellite primers selected and screened 12 microsatellite loci were successfully amplified. After PCR amplification of microsatellite loci and performing native PAGE using amplified DNA samples as above, POP GENE Version 1.32 was used to calculate Nei's observed heterozygosity, expected heterozygosity, Nei's genetic diversity, Fixation index (Fis) and Shannon's information index (SI) and genetic variability indices viz. Gene flow(Nm), the coefficients of genetic differentiation (Fst & Gst) and Nei's genetic distance. Overall Gst value (0.1601) recorded for *L. gonius* suggested the possibility of less gene exchange among the two stocks and indicated that 16.01% variation was attributable to interstock divergence, while 83.99% to individual differences within the stocks. Nei's genetic diversity, calculated from the banding pattern of every primer, ranged from 0.4368 to 0.6922 with mean value of 0.5732 for specimens from Dhaura reservoir whereas it ranged from 0.5136 to 0.8243 with mean value of 0.6770 in specimens from Nanak Sagar reservoir. The observed and expected heterozygosities ranged from 0.4769 to 0.4981 (mean value- 0.4901) and 0.5014 to 0.5267 (mean value- 0.5169) respectively in *L. gonius* from Dhaura reservoir. The observed and expected heterozygosities ranged from 0.4641 to 0.5314 (mean value- 0.5046) and 0.4768 to 0.5682 (mean value- 0.5225) respectively in Nanak Sagar reservoir stock. Slightly better level of observed heterozygosity observed in fish from Nanak Sagar reservoir than Dhaura reservoir might be due to more differentiated stock of Nanak Sagar. Lesser value of observed heterozygosity compared to expected heterozygosity in fish from Dhaura reservoir might be possibly due to increased inbreeding in successive generations owing to small population size restricting desired germplasm exchange of appropriate genetic diversity. These observations indicated that the *L. gonius* stock of Nanak Sagar reservoir is genetically more diverse and differentiated as compared to its stock from Dhaura reservoir.

Keywords: Genetic characterization, heterozygosity microsatellites, primers, *Labeo gonius*

Introduction

Labeo gonius (Hamilton, 1822), a common Cyprinid species, is widely distributed in water bodies of North India, Asom, Odisha and along the east coast up to the Krishna river in India. It is a dominant fish species in different reservoirs of Uttarakhand and has a larger scope as potential candidate species for aquaculture. It spawns during the south-west monsoon during July to August. In Uttarakhand state a major part of aquatic resources is covered by reservoir (18,931 ha) followed by rivers (2700 km) and ponds (600 ha). Two major reservoirs of Uttarakhand viz. Dhaura (1200 ha) and Nanak Sagar (4262 ha) are greatly subjected to environmental aberrations like reduction in water volume, increased sedimentation and water abstraction, catchment area degradation due to siltation, drying of water during summer season, conversion of water area into marshy land due to soil erosion caused by deforestation which makes status of different fishes highly vulnerable and subject to unpredictable genetic changes in these reservoirs. The fish fauna of these reservoirs mainly comprises of various species of catfishes, major carps and minor carps. *L. gonius*, a minor carp contributes substantially in catches of these reservoirs and enjoys a good market value as a food fish in the state.

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At present, its fishery is flourishing well in these reservoirs but for insuring good future prospects for *L. gonius* fishery early attention is required before its decline as practically no attention has been paid for stock assessment, sustainable utilization and genetic management of this species till now. Status of genetic structure of fish is considered essential for their controlled propagation, stock improvement and developing conservation plans. To conserve intra-specific genetic diversity of the fish the status of genetic diversity of the concerned species is a pre-requisite for developing management strategies. The data on genetic diversity can be effectively used to provide scientific basis for developing measures for conservation and management of natural population of the targeted species. Heterozygosity is an important measurement of genetic diversity level and has got much attention from ecologists and aqua-culturists [1]. The best estimate of genetic variation in natural population is the mean observed heterozygosity (H_o) per locus which varies non-randomly between loci, populations and species [2]. Use of molecular markers, especially microsatellites, has been recognized to have great potential in revolutionizing the genetic management of fishery stocks for controlling the level of inbreeding and loss of genetic diversity through its ability to detect genetic uniqueness of individuals, populations or species [3]. Microsatellites or simple sequence repeats (SSRs) are short tandem repeat motifs (1-6 bases) with high level of allelic polymorphism and co-dominant inheritance [4]. Microsatellites tend to be evenly distributed in the genome on all chromosomes and all regions of the chromosome [5]. Microsatellites have been found inside gene coding regions, introns and in the non-gene sequences [6]. Microsatellites markers are preferable because they are co-dominant and highly polymorphic as compared to other genetic markers [7] and have been proved to be very useful in revealing information about allele frequency, heterozygosity, population differentiation, inbreeding co-efficient, gene flow, linkage disequilibrium, stock identification and other parameters that are crucial measures of genetic diversity and population genetics. Recently, microsatellite markers have been developed for some Indian fishes such as *Labeo rohita* [8, 9] and *Catla catla* [10] *Chitala chitala* [11] and *Cirrhinus mrigala* [12]. Previously genetic diversity studies in *L. gonius* from three reservoirs of Uttarakhand has been done through allozyme and RAPD markers [13, 14] but accuracy of predictions based on these markers has always been a matter of concern. Present study is the first attempt to assess the genetic variability status of *L. gonius* from two reservoirs-Dhaura and Nanak Sagar located in Tarai region of Uttarakhand through microsatellite marker with the aim of devising desirable management practices to conserve the available genetic resources of this fish for its sustainable production in these reservoirs.

Materials and Methods

Hundred live specimens (50 from each reservoir) of the fish, *L. gonius* were collected for present study from commercial catches of Nanak Sagar and Dhaura reservoirs. Kidney tissue were dissected out by using sterilized scissors and forceps and stored at -86°C in deep freezer for further analysis. DNA was isolated from the dissected kidney tissue using DNA isolation kit purchased from Genei (Ltd.), Bangalore, India. Total twenty microsatellite primers were selected for cross amplification and to amplify the repeat regions, primers were then modified accordingly using the web based tool (<http://primer3.sourceforge.net>) Primer 3 [15] with an optimum

annealing temperature of 55°C and a minimum GC content of 40-70%.

Amplification of microsatellite loci and analysis of microsatellite data

All the twenty microsatellite primers were initially screened in 2-2 DNA samples of *L. gonius* collected from Nanak Sagar and Dhaura reservoirs. A total of 12 microsatellite loci were successfully amplified which produced clear and polymorphic bands. Eight microsatellite primers produced stutter and unclear bands with low polymorphic information content value (less than 0.50) and hence not considered for further study. PCR amplification of microsatellite loci were performed in a 25 µl reaction mixture, which included 1X PCR buffer (10 mM Tris-HCl pH 9.0, 50 mM KCl), 0.2 mM of each dNTP, 2.0 mM of MgCl₂, 5 p mol of each primer, 1.5 U Taq DNA polymerase and 25-50 ng of template DNA using PCR (Eppendorf, USA). Initial denaturation was performed at 94°C for 3 minutes followed by 30 cycles of 94°C for 30 seconds, locus specific annealing temperatures (52-64°C) for 60 seconds and 72 °C for 90 seconds and a final elongation of 1 cycle at 72 °C for 8 min followed by storing it at 4 °C. Amplified products were mixed with 2 µl of gel loading dye and then separated on 6% denaturing polyacrylamide gel with 1x TBE on PAGE Gel along with standard marker Φ X 174/ Hinf I marker at constant power supply of 25 volts for 2 hrs. Polymorphic information content (PIC) of individual primer was estimated using the formula:

$$\sum_{i=1}^n P_{ij}$$

PIC = $1 - \frac{1}{n} \sum_{i=1}^n P_{ij}$ Where P_{ij} is the frequency of j th allele. After performing native PAGE, POP GENE Version 1.32 [16] was used to calculate Nei's observed heterozygosity (H_o), expected heterozygosity (H_e), Fixation index (F_{is}) and genetic variability indices viz. Gene flow (N_m), the coefficients of genetic differentiation (F_{st} & G_{st}) and Nei's genetic distance. Nei's genetic diversity (H_i) was calculated from the banding pattern of every screened primer across all the loci. Individual genotypes were scored using the Gene Mapper (version 4.0; Applied Biosystems) with a size standard and an internal control for allele calling. Each allele was coded according to its size in nucleotide base pairs (bp). Possible null alleles and genotyping errors caused by stuttering and/or large-allele dropout were tested using MICRO-CHECKER [17]. Scoring and human error were estimated by duplicate analyses. The polymorphic information content (PIC) was calculated by using the CERVUS version 3.03 [18].

Results

Screening of microsatellite primers in *L. gonius*

Microsatellite primers selected for further study based on amplifying successfully and exhibiting desired level of polymorphism have been summarized in Table-1. Out of 20 Primer-BLAST designed microsatellite primers for *L. gonius*, twelve primers exhibited polymorphism.

Microsatellite primers amplification in *L. gonius* from Nanak Sagar reservoir

Microsatellite primers amplification values across all loci in *L. gonius* from Nanak Sagar reservoir are recorded in Table-2. Number of alleles per locus ranged from 5-9 with mean value of 7.08 per locus. A total of 8 SSR loci were scored by the primer PL-01. The product size ranged from 0.11Kb to 0.30 Kb and the average expected genetic diversity and PIC value

of the primer were 0.635 and 0.71 respectively. A total number of 7 SSR loci were scored by the primer PL-02. The product size ranged from 0.14 Kb to 0.35 Kb and the average expected genetic diversity and PIC value of the primer were 0.783 and 0.68 respectively. The totals of 6 SSR loci were scored for the primer PL-03 with product size ranged from 0.25 to 0.37 Kb and the average expected genetic diversity and PIC value of the primer were 0.635 and 0.63 respectively. A total of 8 SSR polymorphic loci were scored for the primer PL-08. The product size ranged from 0.22 Kb to 0.48 Kb and the average expected genetic diversity and PIC value of the primer were 0.680 and 0.69 respectively. Total numbers of 6 SSR loci were scored by the primer PL-10. The product size ranged from 0.18 Kb to 0.70 Kb and the average expected genetic diversity and PIC value of the primer were 0.765 and 0.60 respectively. 7 SSR loci were scored by the primer PL-11 and the product size was 0.25 Kb to 0.65 Kb and the average expected genetic diversity and PIC value of the primer were 0.737 and 0.64. A total of 8 SSR loci with product size ranged 0.20 Kb to 0.70 Kb were scored for the primer PL-13. The average expected genetic diversity and PIC value were 0.824 and 0.68 respectively. Total 5 SSR loci were scored by the primer PL-14 and the average expected genetic diversity and PIC value of the primer were 0.796 and 0.63 respectively and product size ranged from 0.10 to 0.25 kb. 6 SSR loci were scored by the primer PL-15 and the average expected genetic diversity and PIC value of the primer were 0.513 and 0.61 respectively and product size ranged from 0.13 to 0.40 kb. 7 SSR loci were scored by the primer PL-16 and the average expected genetic diversity and PIC value of the primer were 0.576 and 0.65 respectively. Product size ranged from 0.17 to 0.40 kb. 9 SSR loci were scored by the primer PL-17 and the average expected genetic diversity and PIC value of the primer were 0.650 and 0.74 respectively and product size ranged from 0.15 to 0.30 kb. 8 SSR loci were scored by the primer PL-20 and the average expected genetic diversity and PIC value of the primer were 0.630 and 0.67 respectively and product size ranged from 0.20 to 0.40 kb.

Microsatellite primers amplification in *L. gonius* from Dhaura reservoir

Microsatellite primers amplification values across all loci in *L. gonius* from Dhaura reservoir are recorded in Table-2. Number of alleles per locus ranged from 5-9 with mean value of 6.91 per locus. A total of 7 SSR loci were scored by the primer PL-01. The product size ranged from 0.10 Kb to 0.25 Kb and the average expected genetic diversity and PIC value of the primer were 0.510 and 0.67 respectively. A total number of 6 SSR loci were scored by the primer PL-02. The product size ranged from 0.20 Kb to 0.40 Kb and the average expected genetic diversity and PIC value of the primer were 0.634 and 0.60 respectively. The total of 7 SSR loci were scored for the primer PL-03 with product size ranged from 0.20 to 0.42 Kb and the average expected genetic diversity and PIC value of the primer were 0.540 and 0.65 respectively. The total of 9 SSR loci was scored for the primer PL-08. The product size ranged from 0.18 Kb to 0.53 Kb and the average expected genetic diversity and PIC value of the primer were 0.436 and 0.73 respectively. Total numbers of 6 SSR loci were scored by the primer PL-10 and the product size ranged from 0.38 Kb to 0.65 Kb and the average expected genetic diversity and PIC value of the primer were 0.595 and 0.61 respectively. 7 SSR loci was scored by the primer PL-11 and the product size was 0.32 Kb to 0.68 Kb and the expected

genetic diversity and PIC value of the primer were 0.652 and 0.64 respectively. 9 SSR loci with product size ranged 0.28 Kb to 0.74 Kb was scored for the primer PL-13. The average expected genetic diversity and PIC value were 0.623 and 0.73 respectively. 6 SSR loci were scored by the primer PL-14 and the average expected genetic diversity and PIC value of the primer were 0.571 and 0.61 respectively and product size ranged from 0.10 to 0.20 kb. 8 SSR loci were scored by the primer PL-15 and the average expected genetic diversity and PIC value of the primer were 0.692 and 0.66 respectively and product size ranged from 0.25 to 0.35 kb. 5 SSR loci were scored by the primer PL-16 and the average expected genetic diversity and PIC value of the primer were 0.497 and 0.58 respectively. Product size ranged from 0.25 to 0.45 kb. A total of 7 SSR loci were scored by the primer PL-17 and the average expected genetic diversity and PIC value of the primer were 0.548 and 0.63 respectively and product size ranged from 0.15 to 0.30 kb. 6 SSR loci were scored by the primer PL-20 and the average expected genetic diversity and PIC value of the primer were 0.578 and 0.61 respectively and product size ranged from 0.20 to 0.45 kb.

Microsatellite variation based Genetic diversity in *L. gonius* from Nanak Sagar and Dhaura reservoirs

Microsatellite variation based genetic diversity values in *L. gonius* from Dhaura and Nanak Sagar reservoirs are recorded in Table 3 & 4 respectively. Nei's genetic diversity values ranged from 0.4368 to 0.6922 with mean value of 0.5732 for specimens from Dhaura reservoir whereas it ranged from 0.5136 to 0.8243 with mean value of 0.6770 in specimens from Nanak Sagar reservoir. The observed and expected heterozygosities ranged from 0.4769 to 0.4981 (mean value 0.4901) and 0.5014 to 0.5267 (mean value- 0.5169) respectively in *L. gonius* from Dhaura reservoir. The observed and expected heterozygosities ranged from 0.4641 to 0.5314 (mean value- 0.5046) and 0.4768 to 0.5682 (mean value- 0.5225) respectively in Nanak Sagar reservoir stock. The mean values of Fis were found to be 0.124 in Nanak Sagar reservoir and 0.145 in Dhaura reservoir. Mean values for Shannon's information index for all microsatellite loci in *L. gonius* were 1.1862 for Dhaura reservoir population and 1.2342 for Nanak Sagar population.

Genetic divergence in *L. gonius* stocks from Nanak Sagar and Dhaura reservoirs

Observations related with genetic divergence in *L. gonius* stocks from both reservoirs are presented in Table 5. Genetic differentiation (P-value) for *L. gonius* across all loci among different population pairs in Nanak Sagar and Dhaura reservoirs was found to be 0.0184. Values of Gene flow (Nm) and Nei's genetic distance among reservoir populations of *L. gonius* were found to be 1.276 and 0.2134 respectively. Values of coefficients of genetic differentiation (Fst & Gst) observed were 0.093 and 0.1632 respectively for overall population of *L. gonius*. Total genetic diversity in overall population (Ht) and within sample genetic diversity (Hs) was 0.5274 and 0.4430, respectively. The observed (na) and effective number of alleles (ne) in *L. gonius* from Nanak Sagar reservoir were found to be 4.9805 and 4.7762 respectively and for Dhaura reservoir these values were 4.8126 and 4.5531 respectively.

Discussion

In the fish *L. gonius* the genetic diversity value (0.6770) based on observed and expected heterozygosities (0.5046 and

0.5225) from Nanak Sagar reservoir compared to genetic diversity value (0.5732) and observed and expected heterozygosities (0.4901 and 0.5169) from Dhaura reservoir indicated that its stock from Nanak Sagar reservoir exhibited better genetic diversity. These two reservoirs are distinctly separated without any connection and distantly located hence having negligible possibility of gene exchange with each other which might have been responsible for varied sub-structuring of the stocks of *L. gonius*. The values of Nei's genetic diversity in Nanak Sagar (0.6770) and Dhaura (0.5732) reservoir in *L. gonius* assessed through microsatellite marker are found to be higher than Nei's genetic diversity values in these reservoirs (0.3980 and 0.2243) in *L. gonius* by using RAPD marker [14]. Genetic diversity values based on codominant microsatellite markers are considered more accurate and preferable as compared to allozyme and dominant RAPD markers as microsatellite markers are highly polymorphic, capable of detecting small genetic differences (even single nucleotide base change variation) which is more useful in assessing the genetic variability of the populations [19]. Nei's genetic diversity range (0.436 to 0.824) in *L. gonius* (over all loci) was found to be in similar range (0.679 to 0.874) as reported in *T. tambroides* through microsatellite marker [20]. Similar level of observed and expected heterozygosity values in Nanak Sagar population of *L. gonius* and lesser observed heterozygosity value compared to expected heterozygosity in fish from Dhaura might be possibly to some extent correlated with the level of inbreeding incidences in successive generations due to less effective population size limiting germplasm exchange required for maintaining appropriate genetic diversity. Large area of Dhaura gets dried up in summer and most fishes including *L. gonius* are extensively exploited even from deeper isolated pockets thus adversely affecting their effective population size (N_e) available for breeding in the following breeding season. The mean values of observed heterozygosity in *L. gonius* from both the reservoirs were found to be comparable with the mean value of observed heterozygosity (0.46) reported for some (596) other freshwater fishes [4]. The observed heterozygosity range (over all loci) in *L. gonius* are found to be comparable with the observed heterozygosity range (0.0000 to 0.9000) reported in *Tor putitora* using seven microsatellites loci developed from *Catla catla* and *Barbus barbus* by [21]. Similar observations on heterozygosity range were reported for other cyprinid fishes like silver carp and bighead carp by [22]; and in three wild and one farm population of *L. rohita* by Sahoo *et al.* [23]. However, the observed heterozygosity range in *L. gonius* was found to be higher than in *Cirrhinus mrigala* reported from different rivers by [24]. The mean values of observed and expected heterozygosity in *L. gonius* from Nanak Sagar population using microsatellite marker was is comparable with the mean values of observed and expected heterozygosity (0.501 and 0.539) using allozymes marker in *L. gonius* from Nanak Sagar population [13]. Small differences between observed values of population genetic diversity ($H_s=0.4430$) and total genetic

diversity ($H_t=0.5274$) indicated moderate genetic differentiation among *L. gonius* stocks. Heterozygosity results in *L. gonius* recorded in present study seem to be well in agreement with the observations made by [25] in different strains of common carp, *C. carpio* L in which five microsatellite loci were used to study the genetic diversity. Calculated observed ($n_a=4.9805$ and 4.8126) and effective ($n_e=4.7762$ and 4.5531) number of alleles in Nanak Sagar and Dhaura reservoirs indicated that significant genetic variation exists within stocks of *L. gonius* in both the reservoirs. However, relatively lower genetic variability in Dhaura stock as compared to Nanak Sagar stock might be correlated with effective population size owing to exploitation pattern in them. The problems of bottleneck, genetic drift and inbreeding depression, are correlated with small populations, effective population size to population genetic structure of fishes [26]. Overall G_{st} value (0.1601) calculated for *L. gonius* suggested the possibility of less gene exchange among the two stocks and indicated that 16.01% variation was attributable to the interstock divergence, while 83.99% to individual differences within the stock. The moderate level genetic differentiation exhibited by calculated value of coefficient of genetic differentiation across all loci ($F_{st}=0.093$) might be result of difference in effective population size as Dhaura reservoir has smaller population size than Nanak Sagar reservoir. However this pattern of moderate differentiation, is relatively higher than in other Indian fresh water fishes- in *L. dero* ($F_{st}=0.019$) and *L. dussumieri* ($F_{st}=0.041$) through microsatellites exhibiting low level of differentiation reported by [27] and by [28] respectively. Whereas [29] reported high F_{ST} values (0.501 to 0.598) in two species of *Oreochromis* indicating that there was little evidence of introgression between these species and [30] reported low F_{st} value (0.035) in *L. calbasu*. Low gene flow ($N_m=1.276$), among *L. gonius* populations is indicative of their negligible migration resulting in low gene flow level and very little gene exchange among the populations because of less number of migrants as these reservoirs are distinctly separated from each other without any apparent connection. Nei's genetic distance calculated between pairs of stock of *L. gonius* from two Nanak Sagar and Dhaura reservoirs reveal larger genetic distance value (0.2134) between their stocks. Value of genetic distance among *L. gonius* population is correlated with the value of genetic distance ($D=0.171$ to 0.199) reported by [31] in guppy fish, population and with value of genetic distance (0.1898) in *L. gonius* reported by [14]. Comparison among the separate population pairs from these two distantly located reservoirs and having no connection with each other, P -value (0.0184) showed significant genetic differentiation at $P<0.05$ among Nanak Sagar and Dhaura reservoirs populations. As genetic distance value increases with the increase in geographic distance [32] the observations on genetic distance in stocks of *L. gonius* from both the reservoirs might be correlated with the geographical distance between the stock of two distinctly and distantly separated reservoirs.

Table 1: Primer-BLAST designed microsatellite primers for *L. gonius*

Locus	Primer Sequence (5'-3')	Annealing Temp	Annealing Time
PL-01	F-GAAAGCTGCTCGTCCTTGAA R-GAAAGCTGCTCGTCCTTGAA	52 °C	1min 30 sec
PL-02	F-GGGTGTGGGAGAGAAAGAGAG R-GGAGTCTGACAAATGCAGCAAG	64 °C	1 min
PL-03	F-TCTCAGTGGGTGTCATTACCTG R-CCCATCAAACCATCTCTCTAGC	52 °C	1min
PL-08	F-CTGACACTCTTATCTCGCTGCC	53 °C	1min 30 sec

	R-GACCTGAGCAAACAAACCTCAT		
PL-10	F-TCTCTCTTTGTCTTTCCCCTTG R-CACAAGCCACTGTTAGCTTCA	64 °C	1min
PL-11	F-CAAATCTGTGAACATGCAAGC R-CCTAGTCCCCTCTAGTCAGCA	58 °C	1 min 30 sec
PL-13	F-AGATAAGACCCTTCTTCCTCGG R-TTTATTAGGGAGCGTCGAGTG	64 °C	1min
PL-14	F-CTGTTGGTGAAGTGTAGGGTGAA R-GAGAACTCGGTTTGAACATGC	58 °C	1min 30 sec
PL-15	F-ACAGTAATCTTGTGTCTGTCTCTC R-GTCTAAACGTGTCTGAGCTGTG	57 °C	1 min 30 sec
PL-16	F-TGAATGTTTCCAGTCACCACAT R-GTAATGCAGCGGAGAATAAACC	57 °C	1min
PL-17	F-ACAATTCCTGTGTCAACTGTGC R-TACCGTCTCAGTCTCTTTTCGG	55 °C	1min
PL-20	F-ATAGTCGAAATTGGTCTCTGTC R-CAATACCATGACTGAAGTGCC	55 °C	1min 30 sec

Table 2: Microsatellite primers amplification in *L. gonius* from Nanak Sagar and Dhaura reservoir

Locus	Nanak Sagar reservoir			Dhaura reservoir		
	Amplified Product (Kb)	Number of alleles	(PIC)	Amplified Product (Kb)	Number of alleles	(PIC)
PL- 01	0.11-0.30	8	0.71	0.10-0.25	7	0.67
PL-02	0.14-0.35	7	0.68	0.20-0.40	6	0.60
PL-03	0.25-0.37	6	0.63	0.20-0.42	7	0.65
PL-08	0.22-0.48	8	0.69	0.18-0.53	9	0.73
PL-10	0.18-0.70	6	0.60	0.38-0.65	6	0.61
PL-11	0.25-0.65	7	0.64	0.32-0.68	7	0.64
PL-13	0.20-0.70	8	0.68	0.28-0.74	9	0.73
PL-14	0.10-0.25	5	0.59	0.10-0.20	6	0.61
PL-15	0.13-0.4	6	0.61	0.25-0.35	8	0.66
PL-16	0.17-0.40	7	0.65	0.25-0.45	5	0.58
PL-17	0.15-0.30	9	0.74	0.15-0.30	7	0.63
PL-20	0.2-0.40	8	0.67	0.2-0.45	6	0.61

Table 3: Genetic Diversity of *L. gonius* from Dhaura Reservoir based on Microsatellite markers.

Locus	Observed Heterozygosity (Ho)	Expected Heterozygosity (He)	Nei's genetic Diversity (Hi)	Shannon's Information Index	Fixation Index Fis (-ve)
PL- 01	0.4769	0.5014	0.510	1.2596	0.123
PL-02	0.4961	0.5042	0.634	1.0764	0.133
PL-03	0.4883	0.5286	0.540	1.1889	0.148
PL-08	0.4961	0.5143	0.436	1.1743	0.185
PL-10	0.4865	0.5221	0.595	1.2573	0.132
PL-11	0.4947	0.5129	0.652	1.2589	0.110
PL-13	0.4838	0.5224	0.623	1.1320	0.105
PL-14	0.4919	0.5231	0.571	1.1250	0.103
PL-15	0.4861	0.5127	0.692	1.1124	0.111
PL-16	0.4959	0.5228	0.497	1.4021	0.109
PL-17	0.4872	0.5125	0.548	1.2269	0.099
PL-20	0.4981	0.5267	0.578	1.0214	0.092
Mean	0.4901	0.5169	0.5732	1.1862	0.145

Table 4: Genetic Diversity of *L. gonius* from Nanak Sagar based on Microsatellite markers.

Locus	Observed Heterozygosity (Ho)	Expected Heterozygosity (He)	Nei's genetic Diversity (Hi)	Shannon's Information Index	Fixation Index Fis (-ve)
PL- 01	0.5171	0.5289	0.635	1.3675	0.103
PL-02	0.5282	0.5475	0.783	1.3354	0.120
PL-03	0.4737	0.4958	0.635	1.2519	0.118
PL-08	0.5314	0.5682	0.680	1.4005	0.155
PL-10	0.5287	0.5494	0.765	1.4299	0.105
PL-11	0.4721	0.4986	0.737	1.0563	0.090
PL-13	0.5135	0.5327	0.824	1.3321	0.085
PL-14	0.4641	0.4768	0.796	1.1042	0.098
PL-15	0.4859	0.4981	0.513	1.0368	0.091
PL-16	0.5278	0.5379	0.576	1.1525	0.081
PL-17	0.5282	0.5380	0.650	1.1103	0.115
PL-20	0.4853	0.4988	0.630	1.2335	0.081
Mean	0.5046	0.5225	0.6770	1.2342	0.124

Table 5: Genetic variability indices in *L. gonius* stocks from Nanak Sagar and Dhaura reservoirs

Parameters	Values	
Coefficient of genetic differentiation (Fst)	0.093	
Estimation of Gene flow (Nm)	1.276	
Total genetic diversity in population (Ht)	0.5274	
Within sample genetic diversity (Hs)	0.4430	
Coefficient of genetic differentiation (Gst)	0.1601	
Nei's genetic distance	0.2134	
P-Value	0.0184*	
	Nanak Sagar Reservoir	Dhaura Reservoir
Observed number of alleles (na)	4.9805	4.8126
Effective number of alleles (ne)	4.7762	4.5531

*Significant at P < 0.05

Conclusion

On the basis of observations on heterozygosity (Ho and He), Nei's genetic diversity (Hi), Fixation Index (Fis), Shannon's Information Index (SI) and Genetic variability indices it can be inferred that *L. gonius* stock of Nanak Sagar reservoir is genetically more diverse and moderately differentiated compared to Dhaura reservoir stock.

References

- Xu Zh. Genetic diversity of wild and cultured *Penaeus monodon* in the philippines using microsatellites. *Aquaculture*. 2001; 199:13-40.
- Allendorf FW, Utter FM. Population genetics. In W.S. Hoar and D.J.Randall (Eds.). *Fish Physiology*, 8. Academic Press, New York, 1979, 407-454.
- Lakra WS. Fish genetics and conservation research in India: status and perspectives. *Fish Physiology and Biochemistry*. 2007; 33:475-487.
- DeWoody JA, Avise JC. Microsatellite variation in marine, freshwater and anadromous fishes compared with other animals. *Journal of Fish Biology*. 2000; 56(3):461-473.
- Chistiakov DA, Hellemans B, Volckaert FAM. Microsatellites and their genomic distribution, evolution, function and applications: A review with special reference to fish genetics. *Aquaculture*. 2006; 255:1-29.
- Liu ZJ. Microsatellite containing genes from the channel catfish brain:evidence of trinucleotide repeat expansion in coding region of nucleotide excision gene RAD23B. *Biochemistry and Biophysics Research Communication*. 2001; 289:317-324.
- Liu ZJ, Cordes JF. DNA marker technologies and their applications in aquaculture genetics. *Aquaculture*. 2004; 238:1-37
- Das P. Isolation and characterisation of polymorphic microsatellites in *Labeo rohita* and their cross species amplification in related species. *Molecular Ecology Notes*. 2005; 5:231-233.
- Patel A. Development of 21 new microsatellite markers in *Labeo rohita* (rohu). *Animal Genetics*. 2009; 40:253-254.
- McConnell SK. Microsatellite markers from the Indian major carp species, *Catla catla*. *Molecular Ecology Notes*, 2001; 1:115-116.
- Punia P. Polymorphic microsatellite markers isolated from partially enriched genomic library of *Chitala chitala*. *Molecular Ecology Notes*. 2006; 6:1263-1265.
- Lal KK. Genetic variation in Hilsa shad (*Tenualosa ilisha*) population in River Ganges. *Indian Journal of Fisheries*, 2011; 51:33-42
- Tewari, Grishma. Genetic diversity analysis using allozyme marker in *Labeo gonius* (Hamilton, 1822) from two reservoirs of Uttarakhand. *African Journal of Biotechnology*. 2013a; 12(19):2532-2539.
- Tewari Grishma, Singh IJ, Barat AK. Population structure analysis of *L. gonius* from three reservoirs of Uttarakhand using RAPD marker. *Current Science*. 2013b; 105(2):237-241.
- Rozen S, Skaletsky HJ. Primer3 on the WWW for general users and for biologist programmers. *Methods in Molecular Biology*. 2000; 132:365-386.
- Raymond M, Rouseet F. An exact test for population differentiation. *Genetics and Evolution*. 1995; 49:1280-1283.
- Van Oosterhout C. Micro Checker: software for identifying and correcting genotyping errors in microsatellite data. *Molecular Ecology Notes*. 2004; 4:535-538.
- Kalinowski ST, Taper ML, Marshall TC. Revising how the computer program CERVUS accommodates genotyping error increases success in paternity assignment. *Molecular Ecology*. 2007; 16:1099-1106.
- Balloux F, Lugonae MN. The estimation of population differentiation with microsatellite markers. *Molecular Ecology*. 2002; 11:155-165.
- Nguen T. Charecterization of microsatellite DNA markers for a mahseer species, *Tor tambroides* (Cyprinidae) and cross amplification in four congeners. *Molecular Ecology Notes* 2007; 7:11-17.
- Mohindra V. Microsatellite loci to assess genetic variation in *Tor putitora*. *Journal of Applied Ichthyology*. 2004; 20:466-469.
- Tong J. Cross-species amplification in silver carp and bighead carp with microsatellite primers of common carp. *Molecular Ecology Notes*. 2002; 2:245-247.
- Sahoo L. Limited genetic differentiation in *Labeo rohita* (Hamilton 1822) populations as revealed by microsatellite markers. *Biochemical Systematics and Ecology*. 2014; 57:427-431.
- Lal KK. Identification of microsatellite DNA markers for population structure analysis in Indian major carp, *Cirrhinus mrigala*. *Journal of Applied Ichthyology*. 2004; 20:87-91.
- Singh Ekta. Microsatellite Based Genetic Diversity and Differentiation of Common Carp, *Cyprinus carpio* in Rajasthan (India). *National Academy Science Letters*. 2015; 38(3):193-196.
- Ayappan S. *Handbook of Fisheries and Aquaculture*. 2011, 49-51.
- Chaturvedi A. Population genetic structure and phylogeography of cyprinid fish, *Labeo dero* (Hamilton, 1822) inferred from allozyme and microsatellite DNA marker analysis. *Molecular Biology*. 2011; 38:3513-3529.

28. Gopalakrishnan A. Low genetic differentiation in the populations of the malabar carp *Labeo dussumieri* as revealed by allozymes, microsatellites and RAPD. Asian Fisheries Science. 2009; 22:359-391.
29. Appleyard SA, Mather PB. Genetic characterization of cultured Tilapia in Fiji using allozymes and RAPD. Asian Fisheries Science. 2002; 15:249-264.
30. Singh, Rajeev. Genetic diversity of Indian Major Carp, *Labeo calbasu* (Hamilton, 1822) populations inferred from microsatellite loci. Biochemical Systematics and Ecology. 2012; 44:307-316.
31. Khoo G. Genetic diversity within and among feral populations and domesticated strains of the guppy in Singapore. Marine Biotechnology. 2002; 4:367-378.
32. Allendorf FW, Waples RS. Conservation and genetics of salmonid fishes Conservation Genetics. Case Stories from Nature. 1996, 238-280.