

E-ISSN: 2320-7078 P-ISSN: 2349-6800 JEZS 2018; 6(5): 720-726 © 2018 JEZS Received: 11-07-2018 Accepted: 13-08-2018

PB Solanke

Department of Veterinary Parasitology, College of Veterinary and Animal Sciences, Maharashtra Animal and Fishery Sciences University, Parbhani, Maharashtra, India

BW Narladkar

Department of Veterinary Parasitology, College of Veterinary and Animal Sciences, Maharashtra Animal and Fishery Sciences University, Parbhani, Maharashtra, India

Correspondence

PB Solanke Department of Veterinary Parasitology, College of Veterinary and Animal Sciences, Maharashtra Animal and Fishery Sciences University, Parbhani, Maharashtra, India

Journal of Entomology and Zoology Studies

Available online at www.entomoljournal.com



Status of bacterial biocontrol agents against cattle tick *Rhipicephalus* (B.) *microplus* (Acarina: Ixodidiae): Review

PB Solanke and **BW** Narladkar

Abstract

Currently, the control methods against ticks are focusing on the use of insecticides that lead to high cost and adverse effect to the environment. Thus user-friendly and eco-friendly approach of biological control by using entomopathogens *viz*. fungi, bacteria, viruses, and nematodes, provides an alternative way. Within the bacterial group, the microorganisms most widely used worldwide in the control of several insect pests with the highest success, are the bacterium, *Bacillus thuringiensis* Berliner (Bacillales: Bacillaceae). The use of *B.thuringiensis* is increasing rapidly because it is highly specific, significantly lowering the damage to other organisms compared to use of chemical insecticides, and because it is selfperpetuating and is therefore accepted as an environmentally friendly alternative. However very scanty studies are available on bacterial pathogens against ticks. Thus an attempt is made through this review article to highlight the work undertaken on possible ways and role of bacteria in the control of ticks. The sole purpose of the present review is to compile the pertaining literature and to create the interest of researchers in this area.

Keywords: Bacterial bio-control, Bacillus thuringiensis, Rhipicephalus (B.) microplus

Introduction

Concept of Integrated Tick Management: In the current era where a large section of the human population is showing their interest towards organic food and willing to have rid-off form food containing traces of hazardous chemicals. On the other hand the requisite quantity of production of food grains cannot be achieved without control of pests. In such situation the solution which will maintain equilibrium without much damaging to pests and will minimize the use of hazardous chemical pesticides but will not affect on the quantity of production is necessary. Such solution is nothing but the use of bio-control agents and biological control. Same thing is true for control of animal pests and for which necessary thing is the development of bio-control agents. Worldwide research on biological control, bio-control agents and integrated management is in full swing against the crop pests particularly belonging to lepidoptera. However the pace of development of bio-control agents against animal pests is slow and needs to be accelerated. Therefore to highlight the current research undertaken globally, it is presented in the form of present review, which will help to encourage young researchers from India to make efforts in this area of immense importance. The bio-control agents developed against crop pests mainly belongs to bacteria, viruses, fungus, helminthes etc. All these are easy to be used on the lepidospterous pests. However use of bio-control agents against dipteran pests and acarina pests is quite difficult because they need to be applied either on animal body, animal shed or their breeding places. Till then with all difficulties fungal, bacterial and herbal biopesticides and biocontrol agents are used with efficiency and many research article and review article can be quoted as a reference ^[1]. Present study also witnessed the effect of bacteria belonging to *Bacillus* genus and toxins of *B. thuringiensis* var kurstaki on the Rhipiciphalus microplus ticks and these bio-control agents exercised their effect by quadruple mode i.e., a) acaricidal b) oviposition deterrent, c) ovicidal, and d) ovicidal effect perpetuated from parent female tick to eggs and resulted in ovicidal action.

In the literature a very scanty work on use of bacteria against acarina pests i. e. ticks and mites of livestock is being conducted or if conducted, not used on the field. Hence such work is needed to be geared up and one study has been conducted by the author. The said study succeeded in highlighting the importance of five bacteria as bio-control agents against adult

and egg stage of tropical cattle tick Rhipicephalus (Boophilus) microplus (Acari: Ixodidae). Incorporating the results of this study present review paper has been prepared with specific objective to boost the research on this topic. According to Samish et al. [2]. biological control is becoming an increasingly attractive approach to tick management because of: (1) increasing concerns about environmental safety and human health (e.g. the gradual decrease in use of chemical insecticides in several countries is stimulating the growing market of 'organic' food); (2) the increasing costs of chemical control; and (3) the increasing resistance of ticks to pesticides. Classical biological control includes the recognition, evaluation and importation of a natural enemy from elsewhere, the conservation of local natural enemies and the augmentation of the bio-control agents. Further possibility of IPM against ticks was explored by Martinez et al. (2013)^[3] and Singh et al. (2016)^[4].

Chronology of Development of Bt: Singh and Mathew (2015)^[5] in a research article on "The Effect of *Bacillus thuringiensis* and Bt Transgenics on Parasitoids during Biological Control" narrated the global status of *Bacillus thuringiensis which* is presented here in the chronological order. The purpose behind mentioning the chronology of development of *Bacillus thuringiensisis* is to understand how from 1915 to 2015 it acquired the global status. Chronology is also highlighting the importance of these bacteria in biological control:

- 1. 1915: These parasporal inclusions are formed by different insecticidal crystal proteins (ICP) or δ -endotoxins. Though, the existence of parasporal inclusions in Bt was first noted in 1915^[6].
- 2. 1950: Protein composition was not delineated until the 1950s ^[7].
- 3. 1959: Discovery of Bacillus thuringiensis crystal toxicity
- 4. 1977: Bt subspecies can synthesize more than one inclusion, which may contain different ICPs. These crystals have variously shaped depending on their ICP composition. A partial correlation between crystal morphology, ICP composition, and bioactivity against target insects has been established ^[8, 9].
- 5. 1981-1987: Bt is a member of the Bc (*Bacillus cereus*) group of Gram positive, spore-forming soil bacteria. During the sporulation process, it produces one or more characteristic crystalline proteinaceous inclusions adjacent to the endospore, which have been found to be toxic for invertebrates, primarily insect species in the orders *Coleoptera*, *Diptera* and *Lepidoptera*, distinguishing it from *Bacillus cereus*^[10]
- 6. 1984-1991: Once ingested by the target larva, the parasporal crystalline ICP is dissociated to the protoxin form in the midgut, and the protoxin is then activated to a biologically active holotoxin by the proteolytic enzymes and specifically the alkaline environment of the gut ^[11, 12]. Shortly afterwards, the gut becomes paralysed and the larva ceases to feed. Pore or ion channel formation occurs after the binding of the toxin to the receptor and the subsequent failure of trans-membrane electric potential. This results in colloid-osmotic lysis of the cells ^[13], which causes vegetative cells of Bt and the pre-existing microorganisms in the gut to proliferate in the haemocoel causing septicaemia, and may thus contribute to the mortality of the insect larva.

- 7. 1992-1993: This is the leading biopesticide used in commercial agriculture, forest management and mosquito control. *Bacillus thuringiensis* is also a key source of genes for transgenic expressions to provide pest resistance in plants ^[14, 15]
- 8. 1997: proved as one of the most widely used entomopathogenic microorganism among many
- 1997: Bt has attained a wide commercial use against major lepidopteran pests and has emerged as the most successful microbial pesticide having great potential in IPM programmes ^[16].
- 10. 1998: These δ -endotoxins, encoded by the *Cry* and *Cyt* genes, have molecular weights between 14-160 kDa and can be visualized under light microscopy as inclusion bodies ^[17].
- 11. 1998: Bt crops offer great promise in controlling lepidopteran pests. A decrease in synthetic insecticide use in Bt transgenic crops could increase beneficial arthropod diversity and abundance. Among the spray formulations, Bt var. *kurstaki* (Btk) HD-1-based products are widely used in many crop ecosystems against over 100 insect species worldwide including ^[18].
- 12. 1998: The first *Bt* microbial product registration in the U.S. was in 1961 and by 1998, there were approximately 180 products registered in the U.S. Environmental Protection Agency ^[19].
- 13. 1999: The success and extensive use of *Bt* microbial pesticides worldwide can be attributed to their high specificity against target insect species while greatly limiting the negative impacts to beneficial and non-target organisms, and lack of environmental persistence of Bt toxins ^[20-22]. (WHO/IPCS, 1999; Betz *et al*, 2000; Federici and Siegel, 2008)
- 14. 2000: Microbial Bt formulations applied orally or to the host are generally non-toxic against parasitoids, because most hymenopterans lack receptors in their midgut necessary for binding of Cry toxins. However, some laboratory studies using Bt sprays have reported adverse effects ^[23].
- 15. 2006: The Bt toxins in formulations and those expressed by transgenic plants that are commercially grown have a narrow range of activity, and no direct negative effects have been reported on natural enemies belonging to other orders than the one targeted by a specific Bt toxin ^[24].
- 16. 2007: Approximately 276 registered *Bt* microbial formulations in China ^[25].
- 17. 2012: have reported at least 120 microbial products in the European Union
- 18. 2014: Developing suitable methods of pest control in accordance of the philosophy and methodology of modern integrated pest management (IPM) programme is a daunting task in an increasingly environmentally conscious world of ours ^[26].
- 19. 2014: The use of microorganisms has assumed a prominent position among the options that seek to control insect pests without the use of chemicals and with high specific toxicity applied in agro ecosystems
- 20. 2015: For the biological control of insect pests, *Bacillus thuringiensis* (Bt) has emerged as the oldest and one of the most widely used entomopathogenic microorganism ^[27].

Chronology of Bacterial use against the Dipteran pests

1. 1965: The first reported Bacillus sphericus strain active

against mosquito larvae was isolated from moribund mosquito larvae $^{\left[28\right]}$

- 2. 1973: The identification of the strain SSII-I in India and first suggested that like *B. thuringiensis* against Lepidoptera, B. *sphuericus* acts by toxemia rather than septicemia
- 3. 1977-1978: The use of microorganisms as a source of biological compounds for insect pest control started after the discovery of the highly insecticidal bacteria *Bacillus thuringiensis*. The discovery of the strain *B. thuring*iensis serovar *israelensis*^[29, 30]. (de Barjac, 1978; Goldberg and Margalit, 1977) made possible efficient microbiological control of Diptera Nematocera vectors of diseases, such as mosquitoes (Culicidae) and black flies (Simuliidae).

Chronology of Bacterial use against the tick pests

- 1. The possibility of using bacteria probably started from 1997 with the work of Hassanain *et al.* (1997) ^[31] who evaluated the activity of three subspecies of *B.thuringiensis* (*kurstaki, israeliensis,* and *thuringiensis*), spraying spore/crystal mixtures on the soft tick *Argas persicus* and the hard tick *Hyalomma dromedarii*.
- 2. Samish and Rehacek (1999) ^[32] mentioned 100% mortality using mixtures of *B. thuringiensis* spores and blood fed to *Ornithodoros erraticus* through an artificial membrane.
- 3. Zhioua *et al.* (1999) ^[33] evaluated *B. thuringiensis kurstaki* strain against engorged larvae of *Ixodes scapularis*, achieving 96% mortality with a dose of 10^8 spores/ml.
- 4. Brum *et al.* (1991) ^[34] used Enterobacteriaceae as a microbial pathogen against the hard tick *Boophillus microplus* and also found that the microbe has produced a genital infection and/or the death of engorged females.
- Ostfeld et al. (2006) [35] in a review article on 5. 'Controlling Ticks and Tick-borne Zoonoses with Biological and Chemical Agents' described the possible future of natural enemies of ticks include insectivorous birds, parasitoid wasps, nematodes, Bacillus thuringiensis bacteria, and deuteromycete fungi. According to review, although several bacterial species are pathogenic to ticks, the usefulness of bacteria as biocontrol agents is poorly studied. Further they opined that Bacillus thuringiensis, which is used as a bio-control agent for many insects, is pathogenic to ticks, but apparently must be ingested to be effective [32, 33]. Because ticks tend to ingest only host blood, inducing ticks to ingest these bacteria seems impractical, and the prospects for *B. thuringiensis* as a biocontrol agent seem poor. Recent surveys of microbes naturally infecting blacklegged ticks and American dog ticks ^[36, 37]. Reveal a rich flora including spore-forming and crystal-forming bacteria that, if found to be entomopathogenic, could be developed as potential biocontrol agents.
- 6. Fernández-Ruvalcaba *et al.* (2010) ^[42] stated that in Integrated Pest Management (IPM) the use of *B. thuringiensis* is increasing rapidly because it is highly specific, significantly lowering the damage to other organisms compared to use of chemical insecticides, and also because it is biodegradable and is therefore accepted as an environmentally friendly alternative. In addition, *B. thuringiensis* has no adverse effects on humans. *B. thuringiensis* products can be combined with other pest control techniques and it is an essential component in

Integrated Pest Management (IPM).

- 7. Martinez *et al.* (2013) ^[3] studied an Integrated Pest Management of ticks with the aim to use natural products to gradually reduce the use of conventional chemicals. They opined that use of the entomophatogenic fungi, *Metarhizum anosopliae* and *Bacillus thuringiensis*, and extracts from plants that have shown biocidal or biostatic activity that can be used for the control of livestock pests of economic importance like the *R. (Boophilus) microplus* tick.
- 8. Singh *et al.* (2016) ^[4] during the presentation in an international conference on the topic "Prospects for biological control of cattle fever ticks by natural enemies along the Texas-Mexico border" expressed that, for the control of cattle ticks, candidate methods include ants, predatory mites, chickens, parasitoid wasp, *Bacillus thuringiensis*, entomopathogenic nematodes and oxpeckers.

Acaricidal properties of the Entomopathogenic bacteria

About the Bacteria: Martinez *et al.* $(2013)^{[3]}$ in a research article entitled as "Evaluation of natural origin products for the control of *Rhipicephalus (Boophilus) microplus* (Acari: Ixodidae) on cattle artificially infested" described the nature and characteristics of *Bacillus thuringiensis* which are described here. The characteristics of *Bacillus thuringiensis* described are: is a gram-positive bacterium, aerobic strict and its life cycle has two phases: vegetative growth, when the bacteria duplicate by bipartition and the sporulation. *B. thuringiensis*, is considerated an ubiquitous bacteria since it has been isolated from several parts of the world and from diverse systems, like soil, water, plants leafs, insects bodies, spider webs, and others.

Possibility of Bacillus thuringiensis for control of ticks: In the beginning several workers (Samish and Rehacek, 1999; Zhioua et al. 1999) ^[32, 33] expressed their doubt about the usefulness of Bacillus thuringiensis bacteria as bio-control agents because of two reasons, one as a) ticks being hematophagous ingesting only host blood and b) to occur tikicidal activity of bacteria, like insects, bacteria must be ingested in the gut. This problem was addressed by the research of (Habeeb and El-hag, 2008)^[38] who have shown first time that B. thuringiensis toxins are lethal to hemoplast cells of *H. dromedarii* ticks. Hemocytes are the circulating cells of arthropods which are as much as 50-60% of the hemolymph, or the circulating fluid content in ticks (Sonenshine, 1993) ^[39] were functional equivalent to mammalian immune cells. Once immune system is destroyed ticks will not survive. It is a general principle which is well documented by (Giradin et al., 2002; Estrada-Pena et al., 2004) ^[40, 41]. That, innate immune system is one of the most important factors in the ability of metazoan organisms to survive when challenged by microbes. The innate immune system comprises cell-mediated and soluble components and is initiated through recognition of Pathogen Associated Molecule Patterns (PAMP). In the light of this principle, the work of Habeeb and El-hag [38] created a ray of hope and their work has also thrown the light on probable mechanism of action on hemoplast cells. They have undertaken the study by intra-hemocoelic injections of Bacillus thuringiensis serovar thuringiensis H14 -endotoxin (43-kDa Cry4Ba toxin) on the hemocytes which provided the spores access to the more favorable environment of the hemocoel, where they germinate

and reproduce. According to their work mechanism of killing was extensive cell lysis leading to septicemia and death of ticks. They also reported that tick mortality was only in injected groups while the orally exposed ticks were not affected. This may be due to inactivation of B. t. Cry4Ba 43kDa protein by midgut proteases which may not give the toxic protein any chance to interact with hemocytes, while injection of toxic proteins allows it to bind directly with hemocytes. The work of Fernández-Ruvalcaba et al. (2010) [42]. showed another ray of hope in which they concluded that some B. thuringiensis strains had a toxic effect on R. microplus using the adult immersion assay. The R.microplus acarasideresistant strain could be controlled with pathogenic B. thuringiensis Strains and indicated that immersion trials are effective to control R.microplus and mode of action as other than ingestion, probably by means of the spiracles or genital pore. Zhioua et al. (1999) [33] also reported the route of infection as spiracles or genital pore.

The many earlier studies also witness the use of bacteria for control of ticks.

- 1. Acaricidal effect of *B.thuringiensis* varies according to the dose, time of application, and insect species. The toxic effect was most marked during physiologically critical stages such as molting, pupation, or metamorphosis ^[43-49]. Even if mortality is not produced, the surviving insects may succumb at any later stage due to retarded development or failure to accomplish pupation, or emergence ^[50-54].
- 2. Use of bacterial species with high efficacy against adults, immature stages, and eggs of different species of mites [55, 47, 48].
- 3. Carlberg and Lindstrom (1987)^[56] have reported that *Bacillus* strains produce two main types of toxins: delta endotoxins, which are mainly, used for the control of various Lepidoptera, and beta exotoxins, which are mainly used for the control of various Diptera.
- 4. Brum *et al.* (1991) ^[34] used agents from Enterobacteriaceae family as a microbial pathogen against the hard tick *Boophillus microplus*. They also found that the microbe produced a genital infection and/or the death of engorged females.
- 5. Hassanain et al. (1997) ^[31] first time evaluated the activity of three subspecies of *B.thuringiensis* (kurstaki, israeliensis, and thuringiensis), spraying spore/crystal mixtures on the soft tick Argas persicus and the hard tick Hyalomma dromedarii. He reported that *B. thuringiensis kurstaki* produced 100% mortality against *A. persicus* engorged females after five days at a dose of 1 mg/ml. *B. thuringiensis israelensis* caused 100% mortality at a dose of 2.5 mg/ml, and *B.thuringiensis thuringiensis* at a 5 mg/ml dose induced 93.3% mortality. With *H. dromedarii*, none of the *B. thuringiensis* strains produced 100% mortality, even at doses as high as 10 mg/ml.
- 6. Samish and Rehacek $(1999)^{[32]}$ mentioned 100% mortality using mixtures of *B. thuringiensis* spores and blood to feed *Ornithodoros erraticus* through an artificial membrane
- Zhioua *et al.* (1999) ^[33] evaluated a *B.thuringiensis kurstaki* strain against engorged larvae of *Ixodes scapularis*, achieving 96% mortality with a dose of 10⁸ spores/ml. it was shown that *B.thuringiensis kurstaki* spores (10⁶/ml) were toxic to engorged *I. scapularis* larvae. However, an LC₅₀ has been reported with

10⁷spores.

- 8. Casique-Arroyo *et al.* (2007) ^[57] reported that *B. thuringiensis* strains have chitinolytic activities and hence can be used in mite control.
- 9. Ostfeld *et al.* ^[35] reported the use of *B. thuringiensis* for cattle tick control
- 10. Fernández-Ruvalcaba *et al.* (2010) ^[42] The four selected *B. thuringiensis* strains GP123, GP138, GP139, and GP140 produced 62.5, 81.25, 64.58, and 77.08% mortality, respectively, by the fifth day. These data indicated that the GP138 strain was the most pathogenic. Analysis of the effect of *B. thuringiensis* strains on *R. microplus* with the immersion aassay led us to infer that the *B. thuringiensis* strains can affect *R. microplus* through approaches other than ingestion, probably by means of the spiracles or genital pore.
- 11. Martinez *et al.* (2013) ^[3] reported high mortality of adult *R. microplus* females in the presence of *B. thuringiensis kurstaki* strains.
- 12. Dunstand-Guzmán *et al.* (2015) ^[58] reported the high efficacy of *Bacillus thuringiensis* protein extracts on the mite *Psoroptes cuniculi* and observed that acaricidal effect through histological damage.

Action Mechanism: The ultrastruture and characteristics of hemocytes of ixodidae tick, Hyalomma dromedarii, after treatment with Bacillus thuringiensis serovar thuringiensis H14 -endotoxin were studied by Habeeb and El-Hag (2008) ^[38]. To evaluate the effect of *B. thuringiensis* 43-kDa toxins elicit a toxic response in the hemocoel, intra-hemocoelic injections of 43-kDa Cry4Ba toxin on the hemocytes (ultra structure and characteristics) and survival of Hyalomma dromedarii engorged female was studied. After study author reported that, of 43-kDa Cry4Ba toxin was highly toxic within short term (48h) to Hyalomma dromedarii engorged female. The result indicated that the complete growth was arrested and death in a dose-dependent manner. On receiving 10µl of 157µg per ml soluble B. t. toxin/tick a rapid paralysis, followed by hemocytic disruption and death was occurred. This investigation revealed that a severe damage in the cells membrane and granulocytes of the hemolymph after injection with -endotoxin. Bacillus thuringiensis var. thuringiensis H14 -endotoxin, this toxin destroys the granular cell and renders it abnormal. In short toxin kills the tick by causing a malfunction of the cellular immune system of the tick. The study also suggested that Bacillus thuringiensis var. thuringiensis -endotoxins targets are not only the gut but also are haemocoel.

Summarizing the observations of many studies conducted by Hassanain *et al.* (1997), Zhioua *et al.* (1999), Fernandez *et al.*, (2006a), Martinez *et al.* (2013), Solanke (2018) ^[3, 31, 33, 59, 60] it can be concluded that *Rhipicephalus* (*Boophilus*) *microplus* can be effectively controlled through use of bacteria as biological control agents. Three sub species of *B. thuringiensis* (*kurstaki, israeliensis* and *thuringiensis*) against effectively works against tick species as *Argas persicus* and *Hyalomma dromedarii, Ixodes scapularis, Rhipicephalus* (*Boophilus*) *microplus*) species (hard ticks).

Conclusions

An experiment was conducted ^[60] to develop the effective bioacaricides which will be alternative to chemical acaricides against an important pest of cattle *Rhipicephalus* (*B*) *microplus* in which five bacteria tested, which were *Bacillus* thuringiensis var israelensis, Bacillus weihenstephanensis var WSBC, Bacillus weihenstephanensis var KBAB4, and Bacillus sphaericus which were available in the Department of veterinary Parasitology as a lyophilized powder containing 2x 10¹⁰, 1.3x 10¹⁰, 1.3x 10¹⁰ and 1.5x 10¹⁰ spores per gram of powder. One bacteria namely Bacillus thurinngiensis var kurstaki and its toxin was produced from market. For judging the effects of bacteria on Rhipicephalus (Boophilus) microplus ticks for criteria's were employed namely a) mortality of adult ticks and eggs, b) reduction in egg laying capacity, c) hatchability of eggs laid by treated female ticks and, d) hatchability of treated eggs by following standard procedure in the laboratory. These bacteria were found very efficacious and shown efficacy as adulticidal, reduced the significant egg laying capacity of female ticks, reduced the hatchability of eggs laid by treated females and reduced the hatchability of the treated eggs. While causing mortality in the adults, bacteria might have probably entered through either genital opening or cuticle. Thus the present experimentation helped to draw the conclusions that the only ingestion of bacterial toxin is not the sole mode of causing activity but, it can been other routes also. As spores of bacteria can enter through genital opening, spiracles or through cuticle and followed by their entry in haemocoel and producing damage to tick body. The effect of bacteria on adult ticks was comparatively less than the eggs. Eggs were found more susceptible to the action of bacteria. Adult females treated with different bacteria, it resulted in significant reduction in egg laying capacity. Looking into potent activity of all types of bacteria they can be very well inducted in the integrated tick management program. Hence in the integrated tick management programmes particularly against ticks bacterial bio-control agents can be very well inducted along with other managemental practices. Further research in the different geographic areas is the need of hr.

Acknowledgments

The authors would like to thank the Associate Dean, Veterinary and Animal Sciences Parbhani, University, for providing the facilities to carry out the research work.

References

- Greeshma UB, Narladkar BW, Rajurkar SR. *In-vitro* evaluation of the herbal acaricide product against the cattle tick *Rhipicephalus* (B.) *microplus* (Acarina: Ixodidiae) Journal of Entomology and Zoology Studies. 2018; 6(2):544-548.
- Samish M, Ginsberg H, Glazer I. Anti-tick biological control agents: assessment and future perspectives. Published by Cambridge University Press. 2008; 506:447-469.
- Martinez R, Fernadez-Ruvalcaba M, Hernandez-Velazquez VM, Pena-Chora G, Lina-Garcia P, Osorio-Miranda J. Evaluation of natural origin products for the control of *Rhipicephalus (Boophilus) microplus* (Acari: Ixodidae) on cattle artificially infested. Basic Research Journal of Agricultural Science and Review. 2013; 2(3):64-79.
- 4. Singh NK, John AG, Perez de Leon AA. Medical Parasitology and Zoology United States Department of Agriculture-Agricultural Research Service, USA Bacteriol Parasitol 2016; 7:5.
- 5. Singh D, Mathew IL. The Effect of *Bacillus thuringiensis* and Bt Transgenics on Parasitoids during Biological

Control Centre for Environmental and Applied Entomology. International. Journal. Pure Applied. Bioscience. 2015; 3(4):123-13.

- 6. Berliner E. About the sleep sickness of the *Ephestia* kühniella Zell. and its vector *Bacillus thuringiensis*, Z Angewandte Entomology. 1915; 2:29-56.
- 7. Angus TA. A bacterial toxin paralysing silkworm larvae, Nature (Lond). 1954; 173:545-546.
- Höfte H, Whiteley HR. Insecticidal Crystal Proteins of Bacillus thuringiensis. Microbiology Reviews. 1989; 53:242-255.
- 9. Bulla LA, Kramer KJ, Davidson LI. Characterization of the entomocidal parasporal crystal of *Bacillus thuringiensis*, Journal of Bacteriology. 1977; 130:375-383.
- 10. De Barjac H. Insect pathogens in the genus *Bacillus*. In: Berkley RCW & Goodfellow M ed. The aerobic endospore-forming bacteria: Classification and identification. Academic Press Inc., 1981, 241-250.
- 11. Aronson AI, Han ES, McGaughey W, Johnson D. The solubility of inclusion proteins from *Bacillus thuringiensis* is dependent upon protoxin composition and is a factor in toxicity to insects, Applied Environmental Microbiology. 1991; 57:981-986.
- 12. Honée G, Visser B. The mode of action of *Bacillus thuringiensis* crystal proteins, Entomologia Experimentalis et Applicata. 1993; 69:145-155.
- Knowles BH, Ellar DJ. Colloid-osmotic Lysis Is A General Feature of the Mechanism of Action of *Bacillus thuringiensis* d-endotoxins With Different Insect Specificities. Biochimica. et Biophysica. Acta. 1987; 924:509-518.
- McGaughey WH, Whalon ME. Managing insect resistance to *Bacillus thuringiensis* toxins, Science, Wash. 1992; 258(5087):1451-1455.
- 15. Meadows MP. *Bacillus thuringiensis* in the Environment: Ecology and Risk Assessment, in *Bacillus thuringiensis*, An Environmental Biopesticide: Theory and Practice, 1993, 193-220.
- Blumberg D, Navon A, Keren S, Goldenberg S, Ferkovich SM. Interactions among *Helicoverpa armigera* (Lepidoptera: Noctuidae), its larval endoparasitoid *Microplitis croceipes* (Hymenoptera: Braconidae), and Bacillus thuringiensis. Journal of Economic Entomology. 1997; 90(5):1181-1186.
- 17. Schnepf E, Crickmore N, van Rie J, Lereclus D, Baum J, Feitelson J *et al. Bacillus thuringiensis* and its pesticidal crystal proteins. Microbiology and Molecular Biology Reviews. 1998; 62(3):775-806.
- Lee HK, Cheng H, Gill SS. Microbial control of insects. In: Dhaliwal GS, Heinrichs EA (eds) Critical issues in insect pest management. Common wealth Publishers, 1998, 87-117.
- 19. EPA. *Bacillus thuringiensis* RED Facts, EPA-738-F-98-001, 1998.
- WHO. Microbial pest control agent *Bacillus* thuringiensis. Repor UNEP/ILO/WHO (EHC, 217). WHO, Geneva, 1999.
- 21. Betz FS, Hammond BG, Fuchs RL. Safety and advantages of *Bacillus thuringiensis*-protected plants to control insect pests. Regulatory Toxicology and Pharmacology. 2000; 32:156-177.
- 22. Federici BA, Siegel JP. Safety assessment of *Bacillus thuringiensis* and Bt crops used in insect control, Food

Safety of Proteins in Agricultural Biotechnology, ed. B. G. Hammond (Boca Raton, FL: CRC Press), 2008, 22.

- 23. Glare TR, O'Callaghan M. *Bacillus thuringiensis*: biology, ecology and safety. Wiley, Chichester, 2000; 350:26.
- 24. Romeis J, Meissle M, Bigler F. Transgenic crops expressing *Bacillus thuringiensis* toxins and biological control. Nature Biotechnology. 2006; 24:63-71.
- 25. Huang DF, Zhang J, Song FP, Lang ZH. Microbial control and biotechnology research on *Bacillus thuringiensis* in China. Journal of Invertebrate Pathology. 2007; 95:175-180.
- 26. Mathew IL, Singh D, Singh RP, Tripathi CPM. *Bacillus thuringiensis*: The biocontrol agent in a food web perspective. Biolife. 2014; 2(1):348-362.
- De Maagd RA. *Bacillus thuringiensis*-Based Products for Insect Pest Control. In Principles of Plant- Microbe Interactions, Springer International Publishing, 2015, 185-192.
- 28. Kellen W, Clark T, Lindergren J, Ho B, Rogoff M. *Bacillus sphaericus* Neide as a pathogen of mosquitoes. Journal of Invertebrate. Pathology. 1965; 7:44248.
- 29. De Barjac H. Une nouvelle varitt de *Bacillus* thuringiensis tr5s toxique pour les moustiques: *B.* thuringiensis var. israelensis drotype H14. C. R. Acad. Sci. Pans Skr. D. 1978; 286:797-800.
- Goldberg LJ, Margalitt JA. Bacterial spore demonstrating rapid larvicidal activity against Anopheles sergentii, Uranotaenia unguiculata, Culex univitattus, Aedes aegypti and Culex pipiens. Mosquito News. 1977; 37:355-358.
- 31. Hassanain MA, El Garhy FM, Abdel-Ghaffar AF, El-Sharaby A, Abdel Megeed NK. Biological control studies of soft and hard ticks in Egypt. I. The effect of *Bacillus thuringiensis* varieties on soft and hard ticks (Ixodidade). Parasitology Research. 1997; 83:209-213.
- 32. Samish M, Rehacek J. Pathogens and predators of ticks and their potential in biological control. Annual Review of Entomology. 1999; 44:159-182.
- Zhioua E, Heyer K, Browning M. Pathogenicity of Bacillus thuringiensis variety kurstaki to Ixodes scapularis (Acari: Ixodidae). Journal of Medical Entomology. 1999; 36(6):900-2.
- Brum JGW, Teixeira MO, Silva EGD. Infection in engorged females of *Boophilus microplus* (Acari: Ixodidea). I. Etiology and seasonal incidence. Arquivo Brasileiro Medicina Veterinaria Zootecnia. 1991; 43:25-30.
- Ostfeld RS, Price A, Hornbostel LV, Benjamin AM, Keesing F. Controlling ticks and tick-borne zoonosis with biological and chemical agents. Bio Science. 2006; 56(5):383-394.
- 36. Martin PAW, Schmidtmann ET. Isolation of aerobic microbes from *Ixodes scapularis* (Acari: Ixodidae), the vector of Lyme disease in the eastern United States. Journal of Economic Entomology. 1998; 91:864-868.
- Grindle N, Tyner JJ, Clay K, Fuqua C. Identification of Arsenophonustype bacteria from the dog tick *Dermacentor variabilis*. Journal of Invertebrate Pathology. 2003; 83:264-266.
- Habeeb SM, El-hag HAA. Ultrastructural changes in hemocyte cells of hard tick (*Hyalomma dromedarii*: *Ixodidae*): A model of *Bacillus thuringiensis* var. thuringiensis H14 d-endotoxin mode of action. *American.*

Eurasian. Journal of Agriculture and Environmental. Sciences. 2008; 3:829-836.

- 39. Sonenshine DE. Biology of Ticks. Oxford University Press, 1993, 2.
- 40. Giradin SE, Sansonett PJ, Philpott DJ. Intracellular vs. extracellular recognition of pathogens-common concepts in mammals and flies. Trends in Microbiology. 2002; 10:193-199.
- Estrada-Pena A, Bouattow A, Camicas JL, Walker AR. Ticks of domestic animals in the Mediterranean region. A guide to identification of species. First Edition, 2004, 131.
- 42. Fernández-Ruvalcaba M, Peña-Chora G, Romo-Martínez A, Hernández-Velázquez V, de La Parra AB, de La Rosa DP. Evaluation of *Bacillus thuringiensis* pathogenicity for astrain of the tick, *Rhipicephalus microplus*, resistant to chemical pesticides. Journal of Insect Science. 2010; 10:1-6.
- 43. Krieg A. Effectiveness of *B. thuringiensis* exotoxin on *Tetranchys telarius* (Acari: Tetranychidae). Journal of Invertebrate Pathology. 1968; 12:478.
- 44. Angus TA. Implications of some recent studies of *Bacillus thuringiensis*. *Proc.* TV Int. Colloq. Insect Pathol, College Park, Md, U.S.A.: Soc. Invertebrate Patho, 1970, 183-189.
- 45. Hall IM, Hunter DK, Arakawa KY. The effect of the Bexotoxin fraction of *Bacillus thuringiensis* on the citrus red mite. Journal of Invertebrate Pathology. 1971; 18:359-365.
- 46. Sebasta K, Farkas J, Horska K. Thuringiensin, the beta exotoxin of *Bacillus thuringiensis*. In: Burges HD (ed) Microbial control of pests and plant diseases. Academic Press. 1981; 249-281.
- 47. Hoy MA, Ouyang YL. Toxicity of the beta exotoxin of *Bacillus thuringiensis* to *Tetranchys paci*®*cus* and *Metaseiulus occidentalis*. J Econ Entomol. 1987; 80:507-511.
- 48. Neal JW, Lindquist RK, Gott KM, Cascy ML. Activity of the thermostable B-exotoxin of *Bacillus thuringiensis berliner* on *Tetranchys urticae* and *T. cinnobarinus*. Journal of Agricultural Entomology. 1987; 4:33-40.
- 49. Chapman MH, Hoy MA. Relative toxicity of *Bacillus* thuringiensis var. tenebrionis to the two spotted spider mite Tetranchys urticae Koch and its predator Metaseiulus occidentalis. Journal of Applied Entomology. 1991; 111:147-154.
- 50. Hall IM. The use of *B. thuringiensis berliner* to control the western grape leaf skeletonizer. Journal of Economic Entomology. 1955; XLVII:656
- 51. Gricarick AA, Tanada YA. ®eld test for the control of *Trichoplusia ni* (Hbn) on celery with several insecticides and *Bacillus thuringiensis*. Journal of Economic Entomology. 1959; LII:1013-1014.
- Guthrie FE, Rabb RL, Bowery TG. Evaluation of candidate insecticides and insect pathogen for tobacco horn worm control. Journal of Economic Entomology. 1959; LII:798-804.
- 53. Genung WG. Comparision of insecticides, insect pathogens and insecticides-pathogens. Combination for control of cabbage looper *Trichoplusia ni* (Hbn). Florida Entomologist. 1960; XLIII:65-68.
- Fisher R, Rosner L. Toxicology of microbial insecticides: thuricide. Agricultural and Food Chemistry. 1969; VII:686-688.

Journal of Entomology and Zoology Studies

- 55. Miller LK, Lingg AJ, Bulla LA. Jr Bacterial, viral and fungal insecticides. Science. 1983; 219:715-72.
- 56. Carlberg G, Lindstrom R. Testing y-chromosome resistance to *thuringiensis* produced by *Bacillus thuringiensis*, serotype H.1. Journal of Invertebrate Pathology. 1987; 49:194-197.
- 57. Casique A, Dennis B, Rube'n SH, Jose EBC. Development of a recombinant strain of Bacillus thuringiensis subsp. kurstaki HD-73 that produces the endochitinase ChiA74. Antonie van Leeuwenhoek. 2007; 92:1-9.
- 58. Dunstand G, Guadalupe PC, Claudia HC, Mario PM, Víctor MHV, Jorge MM, *et al.* Acaricidal effect and histological damage induced by Bacillus thuringiensis protein extracts on the mite Psoroptes cuniculi. Parasites & Vectors. 2015; 8:285.
- 59. Fernandez RM, Pena-Chora RMA, Hernandez V, Bravo A, De la Rosa JD, Rojas MC et al. Evaluacion y Seleccion in Vitro de Cepas Patogenas de Bacillus thuringiensis contra Boophilus microplus Resistentes a Ixocidas. XXIX Congreso Nacional de Control Biologico. Manzanillo, Colima, Mexico. 2006, 507-511.
- 60. Solanke PB. In-vitro evaluation of bacterial bio-control agents against cattle tick, *Rhipicephalus microplus* (Acarina: Ixodidae), MVSC thesis, Approved by the Maharashtra Animal and Fishery Sciences University, Nagpur, 2018.