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Insect-plant biochemical interactions for plant defense against spotted stem borer, *Chilo partellus*: A research summation

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Abstract

Maize is one of the most important cereal crop which has been reported to be attacked by 139 different insect pests in different phenological stages. Host plant resistance, which is economically viable and ecologically sound can be effectively utilized for managing various insect pests. Various morphological and biochemical characters governing the host plant resistance include a number of trichomes, trichome density, surface wax, leaf toughness and amino acids, proteins and various lipophilic compounds. Thus depending on the capacity to fulfill the nutritional requirement of an insect, a host can be resistant or susceptible. At times, the genotypes identified as resistant/tolerant to *C. partellus* under one agro-ecological region have been found to be susceptible at another region, might be because of genotype x environment interactions. Several biochemical factors, protein and lipids are the major determinants for quality of a host. The nutritional value of protein is determined by its amino acid contents. Apart from protein synthesis, high levels of free amino acids are reported to have some additional functions in neural transmission, detoxification, and synthesis of phospholipids, energy production, and morphogenetic processes that have important biological roles in insects' growth and development. Fatty acids are compounds of basic significance associated with biology of insects including storage of metabolic energy, cell and bio-membrane structure, and in regulatory physiology of insects. The requirement of amino acids and lipophilic compounds for insects can be studied in different agroecologies and this could have implications in developing resistant/tolerant varieties against a particular insect.

Keywords: Maize, amino acids, lipophilic compounds, host plant resistance

Introduction

Maize (*Zea mays* L.) is considered as one of the most important cereal crops after rice and wheat, which plays a vital role in the food economy of the world [199]. Maize is a multipurpose crop, providing food and fuel for human beings, feed for animals, poultry and livestock [75]. It is grown on 8.3 million ha with a production of 21 Mt [36]. Various constraints limit the production of maize, among them insect pests are considered as major yield reducing factors [65]. A total of 139 insect pests have been recorded attacking maize at different phenological stages of plant growth [28]. Among the insect pests attacking maize; spotted stem borer, *C. partellus* (Swinhoe) (Lepidoptera: Crambidae) attain the status of key pest causing 18 to 25% yield loss in Asia [26]. In India, 26.7 to 80.4% crop losses have been recorded across different agro-climatic regions [69, 76]. *C. partellus* is not only the major pest of cereals in Asian countries, it has also attained the status of serious pest worldwide under maize and sorghum cropping systems [6, 92, 91]. Nevertheless, it has also been noted as a pest of pearl millet [43], sugarcane [7] and rice [59]. Due to internal nature of damage, this pest is very difficult to control by conventional insecticides and biological control agents. Thus, there is a need to develop alternative management strategies through production of elite resistant/tolerant variety. The use of insect resistant cultivar has been designated as an essential component of IPM which offers an economically stable and ecologically viable mean to minimize the damage caused by the spotted stem borer.

Importance of host plant resistance to insects

Resistance to stem borer damage is expressed in terms of antixenosis, antibiosis and tolerance. Resistance to *Chilo partellus* is generally governed by additive gene action [86]. Furthermore the resistance expressed by a particular genotype is governed by various environmental

factors, so potential genotypes are usually evaluated by exposing them to different environments before desirable ones are selected. Genotype \times environment interaction is one of the important attributes associated with the differential performance of genotypes tested at different locations [12]. Both genetic and environmental effects are reported to be the contributing factor for expression of phenotypic variation [5]. At times, the genotypes identified as resistant/tolerant to *C. partellus* under one agro-ecological region have been found to be susceptible at another region, might be because of genotype \times environment interactions; however, existence of genetically different populations of *C. partellus* in different agro-climatic zones of India also can not be ignored. The differential response of a genotype for a particular trait across diverse ecologies is defined as the genotype (G) \times environment (E) interaction. The G \times E interaction impose difficulty in selecting the best performing and most stable genotypes. Thus it is important to consider G \times E interaction in plant breeding programs because it is one of the major constraints in selection of a particular genotype in any given environment.

Plant physical condition is influenced by different environmental conditions in diverse agroclimatic conditions which ultimately influence the spatial and temporal pest dynamics leading to differential yield losses. The ability of a plant to provide holistic nutrition to the insects determines the quality of a host. Thus a plant can be either resistant or susceptible based on its inherent ability to impart nutrition to the insect which is also influenced by environmental factors. Different agroecological regions act as modifier of the nutritional quality of the host thus regulate the insect-plant interaction which ultimately regulate the differential insect reaction (resistance or susceptibility) in insects towards different plant genotypes in diverse agroecological conditions. *C. partellus*, attained the status of key pest of maize, because of its adequate nutritional balance out of which different amino acids play a crucial role in determining the nutritional quality of maize. Resistance, a relative character which is determined by the amino acid composition of the plant has been reported to vary in different agroclimatic conditions [71], thus making the maize- stem borer interaction variable in different agroecological conditions.

Sources of spotted stem borer resistance

Host plant resistance is one of the most economical and environmental friendly methods and recognized as a long-term control measure against insect pests [60]. It is one of the effective means of minimizing losses due to insect pests. However, most of the maize varieties and hybrids released for cultivation are susceptible to *C. partellus* [54]. Screening of maize germplasm for resistance to spotted stem borer, *C. partellus* has been reported to impart low to moderate levels of resistance to this pest [20, 72, 84, 47], and several new sources with high levels of stem borer resistance have also been reported and supplemented to the existing resistance sources [27].

Mechanisms of resistance to spotted stem borer

In resistant varieties of maize, all the three mechanisms of resistance, viz., non-preference, antibiosis and tolerance have been reported to be functional [82, 83, 57]. In Asia, various experiments have been carried out to elucidate the mechanism of resistance/ susceptibility in the two maize genotypes, Antigua Group 1 (Resistant) and Basi Local (Susceptible),

against *C. Partellus* [54]. The most notable sources of resistance to *C. partellus* are Antigua Group 1, Population 590, and Population 390 of CIMMYT. Little information is available on sources of resistance to second generation *C. partellus*. Kumar [55] studied the larval establishment and damage by *C. partellus* on plants at anthesis. Severe yield losses can occur at anthesis because *C. partellus* attacks maize directly in the growing ear. Kumar [56] reported a few sources of resistance to second-generation *C. partellus*. The third aspect, tolerance has not been studied adequately well in maize resistance to *C. partellus* although this is the most desirable type of resistance in plants. With tolerance as a mechanism of resistance to insects, the insects are relieved of the strong selection pressure evident in the case of strong antibiosis in plants to insects [3, 54-56].

Basis of resistance to spotted stem borer

Plant characteristics including both morphological and biochemical are responsible in determining the host plant quality [93, 8, 67, 70, 1]. Trichomes on the upper leaf surfaces of resistant genotypes were found related to oviposition non-preference by *C. partellus* [35, 2]. Plant chemicals influence the resistance/susceptibility of the plants either by determining the orientation, feeding and oviposition behaviour of the insects, or by determining the metabolism of insects serving as toxins interfering with the metabolic processes of insects [58, 54, 55, 56]. Plant resistance to insects is complex and depends on interaction of constituent characters, leading to expression of resistance to insect pests [29].

Morphological and anatomical characters imparting resistance to maize stem borer

Several morphological and anatomical plant characters have been reported to be responsible for resistance to insect pests in maize [34, 58, 57, 73, 74, 27]. The role of trichomes in inhibiting oviposition by *C. partellus* has also been experimentally demonstrated earlier [58, 95, 64]. Trichome densities, surface waxes and leaf toughness are considered to have negative effect on the oviposition and development of *C. partellus* [68, 81].

Biochemical factors

Biochemical mechanism of insect defense in crop plants is key to insect-plant interactions, and mainly governed by constitutive and/or induced plant metabolic compounds. The amino acids, phospholipids, fatty acids, steroids, and ascorbic acid have been found to serve as phagostimulants for various insect species [24]. Numerous biochemical factors in maize have also been reported to be associated with resistance/susceptibility to insect pests [49, 50, 57, 73, 74, 86]. Biochemical factors such as phenols and sugars also play an important role in plant defense mechanism to *C. Partellus* [75]. Lignin and other phenolics can strengthen cell walls against digestion and therefore can be anti-nutritional for spotted stem borer [14]. Fiber composed of cellulose, hemicellulose and lignin are the primary plant cell wall components and shows resistance to stem borers [80]. Different concentrations of nitrogen, phosphorus, potassium, iron and silicon in stem tissues are responsible for resistance and susceptibility to *C. partellus* in maize in CM 137 and HY 464 [48]. Distinctly low leaf chlorophyll, carotenoid, nitrogen, crude protein and moisture content were noticed in resistant varieties as compared to susceptible ones [74].

Amino acids for insect nutrition and associated with insect resistance

Apart from protein synthesis, high levels of free amino acids are reported to have some additional functions in neural transmission, detoxification, and synthesis of phospholipids, energy production, and morphogenetic processes that have important biological roles in insects' growth and development. 22 standard amino acids were reported till date, out of which selenocysteine and pyrrolysine are incorporated into proteins by distinctive biosynthetic mechanisms, whereas remaining 20 are directly encoded by the universal genetic code. By using Rose's deletion method the amino acid requirement for insects for 20 different amino acids were detected. Some insects have identical requirement for amino acids, whereas some differ significantly in their requirement for life processes. *Delia antique* (Meigen), *Pectinophora gossypiella* (Saunders), and *Trogoderma granarium* Everts have similar amino acid requirements [46]. This has resulted that the L-forms of Arginine, Histidine, Isoleucine, Leucine, Lysine, Methionine, Phenylalanine, Threonine, Tryptophan and Valine are essential, while the L-forms of Alanine, Aspartic Acid, Cysteine, Cystine, Glutamic Acid, Glutamine, Glycine, Proline, Serine and Tyrosine are reported to be non-essential for majority of the insects studied [79, 17]. The essentiality and non-essentiality of a particular amino acid also differs from organism to organism. Non-essential amino acids found for rat (Proline, Serine, Cystine, and Glycine) were observed to be essential for some insects [42, 45, 46, 25, 88, 41]. Essential amino acids are reported to contribute to synthesis of protein having a carbon skeleton and cannot be synthesized *de novo* by the insects [23], whereas non-essential amino acids play a distinct role in insect defense which are synthesized by insect itself and the amino acids, that affect insect development are dose and species-dependent [45, 40, 94, 37, 4, 52, 16, 100, 19]. Insects are unable to complete its particular life process if it is deprived of any essential nutrient [40, 100]. Moreover the amino acid deficiency in larval stages also affects the performance of an adult. Cangussu and Zucoloto [16] reported that protein deficiency during the immature phase was cause of reduce adult emergence, adult female size, and oocyte maturation and also increase the duration of the larval phase of *Ceratitidis capitata*. High concentrations of glycine and serine were observed to be extremely toxic and had an inhibitory effect on the growth of *D. melanogaster* [45].

The leaf-eating by insects depends on their ability to acquire essential amino acids from dietary protein for their optimal growth. Amino acids being the major source of nitrogen their content in sap act as limiting factor in determining the survival of sap feeding insects. Amino acids, aspartic acid and glutamic acid are reported to cause toxicity to aphids and whiteflies when provided at higher concentration in artificial diet. The low amino acid content of plant tissue, poses a major nutritional challenge to phytophagous insects. Protein as both the major macronutrient and the most commonly limiting nutrient for insect growth is essential for life processes of insects [61]. The essentiality of amino acids is indicated as insect diets containing nutritionally unbalanced amino acids affect herbivory and may also influence host plant utilization pattern among various insect herbivores [63, 51, 44, 11]. For aphid, *Myzus persicae* sucrose and amino acid content were observed to play important role in probing response and feeding rate [62]. Moreover less asparagine content in rice variety impart resistance to brown plant hopper, as this particular amino acid content is less in Mudgo variety of rice,

it is said to be resistant [90]. Brown plant hopper population is stimulated to feed on rice by the presence of two dicarboxylic amino acids such as aspartic acid and glutamic acid [89]. The free amino acid content of watermelon had a significant positive correlation with fruit fly infestation, whereas ascorbic acid contents had a significant negative correlation with percentage fruit infestation and larval density per fruit. Amino acids are major resistance factor for aphids as plants containing low level of amino acids impart resistance to aphids in soybean [21]. The soybean aphid *Aphis glycene* were reported to prefer plants having higher amino acid content. Concentrations of asparagine, aspartic acid, and glutamic acid have also been reported to be responsible for antibiosis against *Myzus persicae* (Sulzer) and white fly, *Bemisia tabaci* (Gennadius) [30]. A number of amino acids especially lysine has been reported to impart resistance to sorghum shoot fly [85]. Moreover resistant lines of oat and barley contain higher amount of glutamic acid and higher quantity of asparagine [98].

Lipophilic compounds for insect nutrition and associated with insect resistance

Fatty acids are compounds of basic significance associated with biology of insects. They play significant role in storage of metabolic energy, cell and bio-membrane structure, and in regulatory physiology of insects. The essentiality of lipid nutrients have been demonstrated by the deletion method, which measures the effect by eliminating one specific component from a chemically defined diet, substitution of an essential nutrient by analogues and the use of radio-labelled precursors to measure endogenous biosynthesis [24]. The insects possessing symbionts in their body live on nutritionally poor or unbalanced diets throughout their life, e.g. phloem sap, vertebrate blood, and wood, and the symbionts (microorganisms) are believed to be a source of essential nutrients, primarily essential amino acids, vitamins, and lipids. Symbiotic bacteria in case of aphid impart it with various essential amino acids, lipids and sterols [32]. Nutritional factors are important determinants for plant utilization by the phloem feeding insects [32]. The nutritional barriers mainly constitute sugars and amino acids, which are regarded as the two most abundant classes of phloem-mobile nutrients [31], and there is a scanty information about the sterol nutrition in phloem-feeding insects. A minority of phloem-feeding insects, including some planthoppers have been reported to derive sterols from fungal symbionts [66], but in case of the great majority of phloem-feeders, bacterial symbionts cannot synthesize sterols *in vivo*. Therefore, most aphids, whitefly, psyllids, mealybugs and leafhoppers have been reported to meet their sterol requirements exclusively from phloem sap [9, 31]. The sterol profile of phloem sap can differ markedly from the sterol profile of bulk plant tissues [87, 10]. Aphids are reported to have limited capacity to convert phytosterols to cholesterol by dealkylation [15], but they differ in their capacity to utilize different phytosterols. In addition to sterols, most insects require polyunsaturated fatty acids, and many studies have shown that either linoleic or linolenic acids adequately fulfil this nutritional requirements [53]. The insects differ in their requirements for different fatty acids. Thelinoleic acid content is positively associated with adult emergence in *Homona coffearea*, when reared on mericid diet. Rock *et al.* [78] in their study concluded that a diet deprived of linseed oil will adversely affect the adult emergence of red-banded leaf roller *Argyrotaenia velutinana*, as the linseed oil contain linoleic acid and linolenic acid.

Deficiency of these two fatty acids caused emergence of adults with their wings partly or entirely lacking in scales. A butterfly species *Morpho peleides* contains a large amount of the polyunsaturated fatty acids linoleic and linolenic acids^[97]. Essential fatty acid deficiency leads to scale wing syndrome in Lepidoptera and Hymenoptera. Various plant sources have been reported to contain several metabolite functional groups such as fatty acids, fatty alcohols, hydrocarbons, sterols and terpenoids, vitamin derivatives. Polyunsaturated fatty acids are considered as one of the most important dietary components of Lepidopteran insects^[38, 39, 53]. The effect of wheat germ oil and various vegetable oils on scale condition and emergence of adults was positively correlated with the linoleic and linolenic acid content of the oils^[78]. The ascorbic acid content is found to be positively associated with larval survival in codling moth, *Carpocapsa pomonella*^[77]. Vanderzant^[96] observed that linoleic acid was more active compound than linolenic acid in *Pectinophora gossypiella* in promoting the adult emergence. Linoleic acid deficiency symptoms were recorded both in *Schistocerca* and *Locusta* during final moult and wing formation^[22, 23]. Similarly, adult emergence was observed to be less for *Anthonomus grandis* (Bohemian) when reared on fat-free diets than on diets containing fat. This was due to the difficulty of the adults to emerge from the pupal cases^[96]. Defatted wheat and alfalfa diet fortified with Linoleic acid were observed to reduce the wing syndrome (Crumpled wings) in *Mamestra brassica* Walker^[13].

Conclusion

Study of various amino acids and lipophilic compounds in maize associated with resistance to various insect pests specially *C. partellus* could have implications for nutritional physiology, development and survival, and play important role in developing resistant varieties against *C. partellus* in maize. Furthermore, the metabolic pathways of diverse amino acids and lipophilic compounds can be elucidated and metabolic intermediates can be upregulated and downregulated as per nutritional requirements of the insects and resistant varieties can be developed. Molecular markers can also be developed for various biochemical constituents such as amino acids and lipophilic compounds to deploy in insect resistant breeding program.

References

1. Agrawal AA. Current trends in the evolutionary ecology of plant defence. *Functional Ecology*. 2011; 25:420-432.
2. Ampofo JKO. *Chilo partellus* (Swinhoe) oviposition on susceptible and resistant genotypes. *Insect Science and its Application*. 1985; 6:323-330.
3. Ampofo JKO. Maize Stalk Borer (Lepidoptera: Pyralidae) damage and plant resistance. *Environmental Entomology*. 1986; 15:1124-1129.
4. Anand R, Anand M. Dietary effect of isomers of essential amino acids on the maggots of *Dacus cucurbitae* (Coquillett). *Journal of Entomological Research*. 1994; 18:109-114.
5. Andrew RL, Wallis IR, Harwood CE, Foley WJ. Genetic and environmental contributions to variation and population divergence in a broad-spectrum foliar defence of *Eucalyptus tricarpa*. *Annals of Botany*. 2010; 105:707-717.
6. Arabjafari KH, Jalali SK. Identification and analysis of host plant resistance in leading maize genotypes against

- spotted stem borer, *Chilo partellus* (Swinhoe) (Lepidoptera: Pyralidae). *Pakistan Journal of Biological Science*. 2007; 10(11):1885-95.
7. Assefa Y, Conlong DE, Van den Berg J, Mitchell A. Distribution of sugarcane stem borers and their natural enemies in small-scale farmers' fields, adjacent margins and wetlands of Ethiopia. *International Journal of Pest Management*. 2010; 56:233-241.
8. Beck SD. Resistance of plants to insects. *Annual Review of Entomology*. 1965; 1:207-232.
9. Behmer ST, Nes WD. Insect sterol nutrition and physiology: A global overview. *Advances in Insect Physiology*. 2003; 31:1-72.
10. Behmer ST, Grebenok RJ, Douglas AE. Plant sterols and host plant suitability for a phloem-feeding insect. *Functional Ecology*. 2011; 25:484-491.
11. Berenbaum MR. Turn about is fair play: Secondary roles for primary compounds. *Journal of Chemical Ecology*. 1995; 21:925-940.
12. Beyene Y, Mugo S, Mutinda C, Tefera T, Karaya H, Ajanga K *et al.* Genotype by environment interactions and yield stability of stem borer resistant maize hybrids in Kenya. *African Journal of Biotechnology*. 2011; 10(23):4752-4758.
13. Braken GK. The Bartha armyworm, *Mamesta configurata* (Lepidoptera: Noctuidae), Effects of dietary linolenic acid on pupal syndrome, wing syndrome, survival, and pupal fat composition. *Canadian Entomologists*. 1982; 114:567-573.
14. Brodeur-Campbell SE, Vucetich JA, Richter DL, Waite TA, Rosemier JN, Tsai CJ. Insect herbivory on low-lignin transgenic aspen. *Environmental Entomology*. 2006; 35:1696-1701.
15. Campbell BC, Nes WD. A reappraisal of sterol biosynthesis and metabolism in aphids. *Journal of Insect Physiology*. 1983; 29:149-156.
16. Cangussu JA, Zucoloto FS. Effect of protein sources on fecundity, food acceptance and sexual choice by *Ceratitis capitata* (Diptera: Tephritidae). *Revista Brasileira de Biologia*. 1997; 57:611-618.
17. Chang CL. Effect of amino acids on larvae and adults of *Ceratitis capitata* (Diptera: Tephritidae). *Annals of the Entomological Society of America*. 2004; 97:529-535.
18. Chang CL. Effect of amino acids on larvae and adults of *Ceratitis capitata* (Diptera: Tephritidae). *Annals of the Entomological Society of America*. 2004; 97:529-535.
19. Chang CL, Albrecht CP, Kurashima R. Adult reproductive capacity of *Ceratitis capitata* (Diptera: Tephritidae) on a chemically defined diet. *Annals of the Entomological Society of America*. 2001; 94:702-706.
20. Chavan BP, Khot RB, Harer PN. Reaction of maize germplasms to maize stem borer, *Chilo partellus* Swinhoe. *Journal of Entomological Research*. 2007; 31:187-190.
21. Chiozza MV, O'Neal ME, MacIntosh GC. Constitutive and induced differential accumulation of amino acid in leaves of susceptible and resistant soybean plants in response to the soybean aphid (Hemiptera: Aphididae). *Environmental Entomology*. 2010; 39:856-864.
22. Dadd RH. The nutritional requirements of locusts-I Development of synthetic diets and lipid requirements. *Journal of Insect Physiology*. 1960; 4:319-348.
23. Dadd RH. The nutritional requirements of locusts. Observations on essential fatty acids, chlorophyll,

- nutritional salt mixtures, and the protein and amino acid components of synthetic diets. *Journal of Insect Physiology*. 1961; 6:126-146.
24. Dadd RH. Qualitative requirements and utilization of nutrients: insects. In: *Handbook Series in Nutrition and Food* (Rehceigl Jr. M., ed.). CRC Press, Cleveland, Ohio. 1977, 305-346.
 25. Davis GRF. Amino acid requirements of *Oryzaephilus surinamensis* (L.) (Coleoptera: Silvanidae) for pupation. *Canadian Journal of Zoology*. 1956; 34:82-85.
 26. Dhaliwal GS, Jindal V, Mohindru B. Crop losses due to insect pests: global and Indian scenario. *Indian Journal of Entomology*. 2015; 77(2):165-168.
 27. Dhillon MK, Gujar GT. Maize genotypes identified with resistance to spotted stem borer, *Chilo partellus* and favourable agronomic traits. *Annals of Plant Protection Sciences*. 2013; 21:224-228.
 28. Dhillon MK, Kalia VK, Gujar GT. Insect pests and their management: Current status and future need of research in quality maize. In: *Maize: Nutrition Dynamics and Novel Uses* (Choudhary, D.P., Kumar, S. and Langan, S. eds.). Springer, New York, USA. 2014a, 95-103.
 29. Dhillon MK, Sharma HC, Reddy BVS, Singh R, Naresh JS, Kai Z. Relative susceptibility of different male-sterile cytoplasms in sorghum to shoot fly, *Atherigona soccata*. *Euphytica*. 2005; 144:275-2833.
 30. Dixit S, Srivastava S, Upadhyay SK, Verma PC, Chandrashekar K. A method to produce insect resistance in plant by altering amino acid content in sap. *International Journal of Biotechnology and Research*. 2013; 3(2):13-20.
 31. Douglas AE. Nutritional physiology of aphid. *Advances in Insect Physiology*. 2003; 31:73-140.
 32. Douglas AE. Phloem-sap feeding by animals: problems and solutions. *Journal of Experimental Botany*. 2006; 57:747-754.
 33. Douglas AE, Price DR, Minto LB, Jones E, Pescod KV, Francois CL *et al*. Sweet problems: insect traits defining the limits to dietary sugar utilisation by the pea aphid, *Acyrtosiphon pisum*. *Journal of Experimental Biology*. 2006; 209:1395-1403.
 34. Durbey SL, Sarup P. Ovipositional responses of moths of *Chilo partellus* (Swinhoe) on different maize germplasms. *Journal of Entomological Research*. 1982; 6:1-9.
 35. Durbey SL, Sarup P. Biological parameters related to antibiosis mechanisms of resistance in maize varieties to *Chilo partellus* (Swinhoe). *Journal of Entomological Research*. 1984; 8:140-147.
 36. FAO. (Food and Agriculture Organisation). FAOSTAT 2011. Food and Agriculture Organisation of the United Nations, Viale delle Terme de Caracella 00153 Rome, Italy, 2011. <http://www.faostat@fao.org>.
 37. Ferro MIT, Zucoloto FS. Effect of the quantity of dietary amino acids on egg production and layings by *Ceratitis capitata* (Diptera, Tephritidae). *Brazilian Journal of Medical and Biological Research*. 1990; 23:525-532.
 38. Fraenkel G, Blewett M. Linoleic acid, alpha tocopherol and other fat soluble substances as nutritional factors for insects. *Nature*. 1945; 155:392-393.
 39. Fraenkel G, Blewett M. Linoleic and Vitamin E and other fat soluble substances in the nutrition of certain insects (*Ephestia kuehniella*, *E. elutella*, *E. cautella* and *Plodia interpunctella* (Lepidoptera). *Journal of Experimental Biology*. 1946; 22:172-190.
 40. Friend WG, Backs RH, Cass LM. Studies on amino acid requirements of larvae of the onion maggot, *Hylemya antiqua* under aseptic conditions. *Canadian Journal of Zoology*. 1957; 35:535-543.
 41. Gilmour D. *The Biochemistry of Insects*. 1961. Academic Press, New York.
 42. Golberg L, DeMeillon B. The nutrition of the larva of *Aedes aegypti* Linnaeus-4. Protein and amino acid requirements. *Biochemical Journal*. 1948; 43:379-387.
 43. Harris K. Keynote address: bioecology of *Chilo* species. *International Journal of Tropical Insect Science*. 1990; 11:467-477.
 44. Haukioja E, Ruohomaki K, Suomela J, Vuorisalo T. Nutritional quality as a defense against herbivores. *Forest Ecology Management*. 1991; 39:237-245.
 45. Hinton T, Noyes DT, Ellis J. Amino acids and growth factors in a chemically defined medium for *Drosophila*. *Physiological Zoology*. 1951; 24:335-353.
 46. House HL. Nutritional studies with *Pseudosarcophaga affinis* (Fall.), a Dipterous parasite of the spruce budworm *Choristoneura fumigerana* (Clem.). Three effects of 19 amino acids on growth. *Canadian Journal of Zoology*. 1954; 32:351-357.
 47. IIMR. Annual Report 2015. Indian Institute of Maize Research (IIMR), Pusa Campus, New Delhi, 2015.
 48. Jalali SK, Lalitha Y, Kamath SP, Mohan KS, Head GP. Baseline sensitivity of lepidopteran corn pests in India to Cry1Ab insecticidal protein of *Bacillus thuringiensis*. *Pest Management Science*. 2010; 66:809-815.
 49. Kabre GB, Ghorpade SA. Studies on association of some chemical constituents of maize genotypes with susceptibility of stem borer. *Journal of Maharashtra Agricultural University*. 1997; 22:301-304.
 50. Kabre GB, Ghorpade SA. Susceptibility to maize stem borer, *Chilo partellus* (Swinhoe) in relation to sugars, proteins and free amino acids content of maize germplasm and F1 hybrids. *Journal of Insect Science*. 1999; 12:37-40.
 51. Karowe DN, Martin MM. The effects of quantity and quality of diet nitrogen on the growth, efficiency of food utilization, nitrogen budget, and metabolic rate of 5th instar *Spodoptera eridania* larvae (Lepidoptera, Noctuidae). *Journal of Insect Physiology*. 1989; 35:699-708.
 52. Kaur S, Srivastava BG. Effect of amino acids on various parameters of reproductive potential of *Dacus cucurbitae* (Coquillett). *Indian Journal of Entomology*. 1994; 56:370-380.
 53. Kerkurt GA, Gilbert LI. *Comprehensive Insect Physiology, Biochemistry and Pharmacology*. 1985. Pergamon press, Oxford, UK.
 54. Kumar H. Inhibition of ovipositional responses of *Chilo partellus* (Lepidoptera: Pyralidae) by the trichomes on the lower leaf surface of a maize. *Journal of Economic Entomology*. 1992a; 85:1736-1739.
 55. Kumar H. Oviposition, larval arrest and establishment of *Chilo partellus* (Lepidoptera: Pyralidae) on maize genotypes during anthesis. *Bulletin of Entomological Research*. 1992b; 82:355-360.
 56. Kumar H. Components of resistance in maize (*Zea mays* L.) to first and second generation *Chilo partellus* (Swinhoe). *Maydica*. 1994c; 39:165-170.
 57. Kumar H. Resistance in maize to *Chilo partellus*

- (Swinhoe) (Lepidoptera: Pyralidae): An overview. Crop Protection. 1997; 16:243-250.
58. Kumar H, Saxena KN. Ovipositional responses of *Chilo partellus* (Swinhoe) to certain susceptible and resistant maize genotypes. Insect Science and its Application. 1985; 6:331-335.
 59. Kumar V, Kanta U. Effectiveness of *Trichogramma chilonis* Ishii in the suppression of *Chilo partellus* (Swinhoe) in summer maize. Journal of Biological Control. 2011; 25:92-97.
 60. Luginbill P. Developing resistant plants, the ideal method of controlling insects. USDA-Agricultural Research Service Production Research Report. 1969; 111:1-14.
 61. Mattson WJ. Herbivory in relation to plant nitrogen content. Annual Review Ecological System. 1980; 11:119-161.
 62. Mittler TE, Dadd RH. Differences in the probing responses of *Myzus persicae* (Sulzer) elicited by different feeding solutions behind a parafilm membrane. Entomologia Experimentalis et Applicata. 1965; 8(2):107-122.
 63. Moran N, Hamilton WD. Low nutritive quality as defense against herbivores. Journal of Theoretical Biology. 1980; 86:247-254.
 64. Munyiri SW, Mugo SN, Otim M, Tefera T, Beyene Y, Mwololo JK *et al.* Responses of tropical maize landraces to damage by *Chilo partellus* stem borer. African Journal of Biotechnology. 2013; 12:1229-1235.
 65. Ngoko Z, Cardwell KF, Marasas WFO, Wingfield MJ, Ndemah R, Schulthess F *et al.* Biological and physical constraints on maize production in the Humid Forest and Western Highlands of Cameroon. European Journal of Plant Pathology. 2002; 108:893-902.
 66. Noda H, Koizumi Y. Sterol biosynthesis by symbiotes: cytochrome P450 sterol C-22 desaturase genes from yeast like symbiotes of rice planthoppers and anobiid beetles. Insect Biochemistry and Molecular Biology. 2003; 33:649-658.
 67. Norris DM, Kogan M. Biochemical and morphological bases of resistance. In: Breeding Plants Resistant to Insects (Maxwell, F.G. and Jennings, P.R., eds.). John Wiley and Sons, New York, 1980, 23-62.
 68. Padhi G. Biochemical basis of resistance in rice to yellow stem borer, *Scirpophaga incertulas* Wlk. Madras Agricultural Journal. 2004; 91(4-6):253-256.
 69. Panwar VPS, Mukherjee BK, Ahuja VP. Maize inbreds resistant to tissue borer, *Chilo partellus* and *Atherigona* spp. Indian Journal of Genetics and Plant Breeding. 2000; 60(1):71-75.
 70. Pathak MD, Dale AA. The biochemical basis of resistance in host plants to insects. In: Chemistry and World Food Supplies: The New Frontiers (Schemilt, L.W., ed.). CHEMRAWN II. 1983, 129-142.
 71. Pavlik M, Pavlikova D, Balik J, Neuberger M. The contents of amino acids and sterols in maize plants growing under different nitrogen conditions. Plant, Soil and Environment. 2010; 56(3):125-132.
 72. Rakshit S, Kaul J, Dass S, Singh R, Sekhar JC, Singh SB. Catalogue of Indian maize inbred lines. Technical Bulletin No. 2008/3. Directorate of Maize Research, Pusa Campus, New Delhi 110 012, India. 2008, 40.
 73. Rao CN, Panwar VPS. Morphological plant characters affecting resistance to *Chilo partellus* in maize. Annals of Plant Protection Sciences. 2000; 8:145-149.
 74. Rao CN, Panwar VPS. Anatomical plant characters affecting resistance to *Chilo partellus* (Swinhoe) in maize. Annals of Plant Protection Sciences. 2001; 9:304-306.
 75. Rasool I, Wani AR, Nisar M, Dar ZA, Nehru RK, Hussain B. Antixenosis and antibiosis as a resistance mechanism to *Chilo partellus* (Swinhoe) (Lepidoptera: Pyralidae) in some maize genotypes. Journal of Entomology and Zoology Studies. 2017; 5(2):22-27.
 76. Reddy KVS, Zehr UB. Novel strategies for overcoming pests and diseases in India. In: New Directions for a Diverse Planet, Proceedings of 4th International Crop Science Congress, 26 Sep–1 Oct., 2004 (Fisher T. *et al.* eds.), Brisbane, Australia. 2004, 1-8.
 77. Rock GC. Aseptic rearing of the codling moth on synthetic diets: Ascorbic acid and fatty acid requirements. Journal of Economic Entomology. 1967; 60(4):1002-1005.
 78. Rock GC, Patton RL, Glass EH. Studies of the fatty acid requirement of *Argyrotaenia velutinana* (Walker). Journal of Insect Physiology. 1965; 11:91-101.
 79. Rose WC. The nutritive significance of the amino acids. Physiological Reviews. 1938; 18:109-136.
 80. Santiago R, Barros-Rios J, Malvar RA. Impact of cell wall composition on maize resistance to pests and diseases. International Journal of Molecular Sciences. 2013; 14(4):6960-6980.
 81. Sarwar M. Management of rice stem borers (Lepidoptera: Pyralidae) through host plant resistance in early, medium and late plantings of rice (*Oryza sativa* L.). Journal of Cereals and Oil Seeds. 2012; 3:10-14.
 82. Saxena KN. Patterns of insect plant relationships determining susceptibility or resistance of different plant to an insect. Entomologia Experimentalis et Applicata. 1969; 17:303-318.
 83. Saxena KN. Behavioural basis of plant resistance or susceptibility to insects. Insect Science and its Application. 1985; 6:303-313.
 84. Sekhar JC, Rakshit S, Kumar P, Mehrajuddin, Anuradha M, Dass S. Differential reaction of CIMMYT maize lines and their hybrid combinations to pink stem borer, *Sesamia inferens* Walker. Annals of Plant Protection Sciences. 2008; 16:404-406.
 85. Sharma HC, Nwanze KF. Mechanisms of resistance to insects in sorghum and their usefulness in crop improvement. Information Bulletin No. 45. International Crops Research Institute for the Semi-Arid Tropics (ICRISAT), Patancheru, Andhra Pradesh, India, 1997, 56.
 86. Sharma HC, Dhillon MK, Pampapathy G, Reddy BVS. Inheritance of resistance to spotted stem borer, *Chilo partellus* in sorghum, *Sorghum bicolor*. Euphytica. 2007; 156:117-128.
 87. Shigematsu Y, Murofushi N, Ito K, Kaneda C, Kawabe S, Takahashi N. Sterols and asparagine in the rice plant, endogenous factors related to resistance against the brown planthopper (*Nilaparvata lugens*). Agricultural and Biological Chemistry. 1982; 46:2877-2879.
 88. Singh KRP, Brown AWA. Nutritional requirements of *Aedes aegypti* L. Journal of Insect Physiology. 1957; 1:199-220.
 89. Sogawa K. The rice brown planthopper: feeding physiology and host plant interactions. Annual Review of Entomology. 1982; 27:49-73.

90. Sogawa K, Pathak MD. Mechanisms of brown planthopper resistance in Mudgo variety of rice (Hemiptera: Delphacidae). *Applied Entomology and Zoology*. 1970; 5:145-158.
91. Sylvain NM, Manyangarirwa W, Tuarira M, Onesime MK. Effect of the Lepidoptera stem borers, *Busseola fusca* (Fuller) and *Chilo partellus* (Swinhoe) on green mealies production. *International Journal of Innovative Research and Development*. 2015; 4(10):366-374.
92. Tamiru A, Getu E, Jembere B, Bruce T. Effect of temperature and relative humidity on the development and fecundity of *Chilo partellus* (Swinhoe) (Lepidoptera: Crambidae). *Bulletin of Entomological Research*. 2012; 102:9-15.
93. Thorsteinson AJ. Host selection in phytophagous insects. *Annual Review of Entomology*. 1960; 5:193-218.
94. Tsiropoulos GJ. Holidic diets and nutritional requirements for survival and reproduction of the walnut husk fly. *Journal of Insect Physiology*. 1978; 24:239-242.
95. Van den Berg J. Oviposition preference and larval survival of *Chilo partellus* (Lepidoptera: Pyralidae) on Napier grass (*Pennisetum purpureum*) trap crops. *International Journal of Pest Management*. 2006; 52:39-44.
96. Vanderzant ES. Nutrition of boll weevil larva. *Journal of Economic Entomology*. 1963; 56:357-362.
97. Wang Y, Lin DS, Bolewicz L, Connor WE. The predominance of polyunsaturated fatty acids in the butterfly *Morpho peleides* before and after metamorphosis. *Journal of Lipid Research*. 2006; 47:530-536.
98. Weibull JHW. Free amino acids in the phloem sap from oats and barley resistant to *Rhopalosiphum padi*. *Phytochemistry*. 1987; 27: 2069-2072.
99. Yonow T, Kriticos DJ, Ota N, Van den Berg J, Hutchison WD. The potential global distribution of *Chilo partellus*, including consideration of irrigation and cropping patterns. *Journal of Pest Science*. 2017; 90:459-477.
100. Zografou EN, Tsiropoulos GJ, Margaritis LH. Survival, fecundity and fertility of *Bactrocera oleae*, as affected by amino acid analogues. *Entomologia Experimentalis et Applicata*. 1998; 87(2):125-132.