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Haemato-biochemical evaluation of midazolam propofol induction combination isoflurane anaesthesia in cattle

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Abstract

Six animals were administered Midazolam (3mg/kg) intravenously, 10 minutes later Propofol was given @3mg/kg body weight intravenously for induction of anaesthesia followed by immediate intubation; maintenance of anaesthesia was done under isoflurane. Anaesthetic combinations were compared by haematological and biochemical observations.

Haematological observations in the present study revealed that haemoglobin, packed cell volume and total erythrocyte count decreased significantly at maximum depth of anaesthesia in all the animals. Neutrophilia with relative lymphocytopenia were recorded during anaesthesia suggesting, certain amount of anaesthetic stress was produced by these combinations. In all the animals most of the biochemical parameters showed the changes, which were within the normal range, suggesting that the anaesthetic combinations used in the present study do not produce adverse effect on these parameters.

Keywords: midazolam, propofol, isoflurane, cattle

1. Introduction

Cattle being docile animals allow many of the surgical and diagnostic procedures to be performed under physical restraint in conjunction with local or regional anaesthesia blockade with or without sedation. However, the resulting conditions are sub-optimal for a number of situations for complex and prolonged surgical procedures and the use of anaesthesia is desired (Adetunji *et al.*, 1984) ^[1].

General anaesthesia may be preferred over local analgesia for many surgical interventions, as it provides complete unconsciousness, better insensitivity to pain, good muscle relaxation, and freedom from reflex responses and loss of motor ability (Thurmon *et al.*, 1996) ^[2].

Intravenous anaesthesia involves the delivery of a bolus dose or a fast loading infusion to achieve an adequate blood concentration of the drug. In veterinary practice, intravenous anaesthetic drugs are commonly used for induction of anaesthesia in order to facilitate endotracheal intubation, oxygen administration and artificial ventilation. Maintenance of anaesthesia can be obtained by infusion of intermittent boluses (IBI) or by continuous rate infusion (CRI), which causes a cumulative effect and prolonged recovery (Malik, 2014) ^[3]. Hence, inhalant anaesthetic agents, which provide predictable and rapid adjustment of anaesthetic depth and help to minimize the patient morbidity and mortality, form the foundation for maintenance of general anaesthesia (Mckenzi, 2008) ^[4]. Anatomical considerations in cattle carry the risk of complications like tympany, regurgitation and aspiration pneumonia, which could be minimized by fasting the animal before anaesthesia. Heavy body weight in adult cattle carries greater risk of developing myopathies and neuropathies following prolonged recumbency so; good positioning and protective padding must be ensured.

Currently ketamine is used as induction agent in cattle, along with several pre- anaesthetic agents, xylazine (Arai *et al.*, 2006) ^[1], diazepam (Riazuddin *et al.*, 2004a) ^[7], acepromazine (Kumar *et al.*, 2012) ^[5] and guaifenesin (Riazuddin *et al.*, 2004b) ^[6] under isoflurane anaesthesia.

2. Materials and Method**2.1. Source of Animals**

The study was conducted in 06 clinical cases presented to VCC, Veterinary College, Bidar,

with various surgical conditions to evaluate haemato-biochemical observations for midazolam-propofol induction combinations under isoflurane anaesthesia in cattle.

2.2 Grouping of animals and anaesthetic protocol

In all the animals, midazolam was administered at the dose rate of 0.4 mg per kg body weight intravenously, five minutes later the animals were restrained in lateral recumbency and anaesthesia was induced by administering propofol intravenously, at the dose rate of 3mg/kg body weight, followed by immediate intubation, the animals were maintained on 5 per cent to 1 per cent of isoflurane.

2.3 Haematological evaluation

The haematological observations *viz.*, Haemoglobin, Packed cell volume, TEC, TLC and DLC was estimated before administration of any drug, immediately after induction (0 min) and then at 30 min, 60 min and 24 hr. After induction of anaesthesia.

2.4 Biochemical evaluation

The biochemical observations *viz.*, alanine transaminase, aspartate transaminase, creatinine and serum urea nitrogen was estimated before administration of any drug, immediately after induction (0 min) and then at 30 min, 60 min and 24 hr. after induction of anaesthesia.

3. Results

Haematological observations

The haemoglobin (Mean \pm SE) in all the animals before anaesthesia, immediately after induction (0 minute), and then at 30 minutes, 60 minutes and 24hrs after induction were; 10.15 \pm 0.75, 10.48 \pm 0.62, 09.35 \pm 0.86, 09.23 \pm 0.90 and 09.20 \pm 0.62 respectively (table no1). The comparison between the groups at different intervals revealed that there was no statistically significant ($P > 0.05$) difference in the haemoglobin.

Packed cell volume (%)

The packed cell volume (Mean \pm SE) in all the animals before anaesthesia, immediately after induction (0 minute), and then at 30 minutes, 60 minutes and 24hrs after induction were; 35.48 \pm 2.83, 33.15 \pm 2.93, 30.40 \pm 2.96, 28.40 \pm 3.40 and 30.20 \pm 2.17 (table no1) respectively.

Total erythrocyte count (x10⁶ / μ l)

The total erythrocyte count (Mean \pm SE) in all the animals before anaesthesia, immediately after induction (0 minute), and then at 30 minutes, 60 minutes and 24hrs after induction were; 7.62 \pm 0.53, 7.26 \pm 0.54, 6.79 \pm 0.52, 6.49 \pm 0.56 and 7.11 \pm 0.40 (table no.1) respectively.

Total leucocyte count (x10³ / μ l)

The total leucocyte count (Mean \pm SE) in all the animals before anaesthesia, immediately after induction (0 minute), and then at 30 minutes, 60 minutes and 24hrs after induction were; 11.72 \pm 1.49, 09.63 \pm 0.94, 08.20 \pm 0.57, 08.77 \pm 0.56 and 10.12 \pm 0.84 (table no1) respectively

Neutrophils (%)

The total neutrophils (Mean \pm SE) in all the animals before anaesthesia, immediately after induction (0 minute), and then at 30 minutes, 60 minutes and 24hrs after induction were; 32.40 \pm 1.33, 34.10 \pm 1.47, 41.30 \pm 1.37, 32.43 \pm 1.47 and

40.10 \pm 2.12 (table no2) respectively.

Lymphocytes (%)

The lymphocytes (Mean \pm SE) in all the animals before anaesthesia, immediately after induction (0 minute), and then at 30 minutes, 60 minutes and 24hrs after induction were; 61.80 \pm 1.54, 59.43 \pm 1.76, 50.10 \pm 2.28, 47.23 \pm 1.83 and 53.27 \pm 2.14 (table no2) respectively.

Biochemical observations

Alanine transaminase (IU/L)

The alanine transaminase (Mean \pm SE) in all the animals before anaesthesia, immediately after induction (0 minute), and then at 30 minutes, 60 minutes and 24hrs after induction were; 24.50 \pm 2.71, 23.67 \pm 1.45, 23.67 \pm 2.19, 26.17 \pm 2.68 and 27.83 \pm 3.74 (table no3) respectively.

Aspartate transaminase (IU/L)

The Aspartate transaminase (Mean \pm SE) in all the animals before anaesthesia, immediately after induction (0 minute), and then at 30 minutes, 60 minutes and 24hrs after induction were; 69.00 \pm 11.60, 73.83 \pm 10.11, 74.17 \pm 11.06, 91.00 \pm 11.97 and 94.20 \pm 14.98 (table no3) respectively.

Creatinine (mg/dl)

The Creatinine (mg/dl) (Mean \pm SE) in all the animals before anaesthesia, immediately after induction (0 minute), and then at 30 minutes, 60 minutes and 24hrs after induction were; 1.73 \pm 0.16, 1.74 \pm 0.16, 1.89 \pm 0.14, 1.81 \pm 0.7 and 1.82 \pm 0.25 (table no3) respectively.

Serum Urea Nitrogen (mg/dl)

Serum Urea Nitrogen (mg/dl) (Mean \pm SE) in all the animals before anaesthesia, immediately after induction (0 minute), and then at 30 minutes, 60 minutes and 24hrs after induction were; 27.63 \pm 3.64, 27.27 \pm 4.15, 27.95 \pm 3.74, 31.15 \pm 4.47 and 26.60 \pm 2.26 (table no3) respectively.

4. Discussion

Haematological observations

Haemoglobin decreased significantly from 60 minutes to 24hrs after induction, and there was gradual increase in the post-anaesthetic period, however, it remained come to physiological limits after 24hrs hours after anaesthesia.

Packed cell volume, total erythrocyte count, total leucocyte count significant decreased from 30 minutes to 60 minutes and after 24 hrs its come to normal physiological limits of post induction. The decrease in haemoglobin during sedation may be caused by the shifting of fluid from the extravascular compartment to the intravascular compartment in order to maintain normal cardiac output (Wagner *et al.*, 1991) [8]. The decreased haemoglobin has been reported after administration of dexmedetomidine in dogs (Gupta, 2010) [9] and sheep (Monsang, 2011) [10]. De Moor and Desmet (1979) [11] reported a decrease in haemoglobin after giving xylazine in cattle. Decrease in total erythrocyte count is due to stress response towards anaesthetic drugs. Amreshkumar *et al.* (1979) [12] stated that these changes could be due to result of animal response to stress caused by anaesthetic drugs. A significant increase in neutrophils with a subsequent significant decrease in lymphocytes was observed under ketamine-isoflurane anaesthesia in all the animals. These changes were nearer to normal levels and were probably related to response of animal to anaesthesia and surgery.

Peshin *et al.* (1980) [13] reported a decrease in lymphocytes with subsequent increase in neutrophils in dogs administered with xylazine.

Biochemical observations

Alanine transaminase and aspartate transaminase fluctuated within normal limits in all animals. Alanine transaminase and aspartate transaminase increased significantly at 24 hours after induction and they remained significantly higher even 48 hours after anesthesia in all the animals similar findings has been reported by Nuh (2008) [14] after detomidine-midazolam-ketamine anesthesia in calves, however, they reported that the values returned to the pre-anesthetic level by 24 hours after anesthesia. Abu-Ahmed (2013) [15] observed no significant change in the alanine transaminase and aspartate transaminase during midazolam ketamine anesthesia in goats. All the general anesthetics lower the circulation to liver (Malik and Singh, 2007) [16] and the changes in alanine transaminase and aspartate transaminase during present study might be due to this fact.

Creatinine values remained within normal limits and no

significant change in the values were observed throughout anesthesia in all animals. Similar findings were recorded after midazolam-ketamine anesthesia in goats (Abu-Ahmed, 2013) [17] and isoflurane anesthesia in sheep (Hikasa *et al.*, 2000) [18].

A significant increase in the serum urea nitrogen was observed in all the animals. Increased hepatic urea production from amino acid degradation might account for the observed increase in serum urea nitrogen. Increase in serum urea nitrogen was reported after detomidine-midazolam-ketamine anesthesia in calves (Nuh, 2008) [19], however Abu-Ahmed (2013) [20] observed no significant change in the serum urea nitrogen during midazolam-ketamine anesthesia in goats.

5. Conclusion

In conclusion Haematological and biochemical observations revealed that haemoglobin, packed cell volume and total erythrocyte count decreased significantly at maximum depth of anesthesia in all the animals followed by all the biochemical parameters are within limits, the above said combination for surgery in cattle may be good for general anaesthesia.

Table 1: Mean \pm SE values of Haematological parameters at different intervals in all the animals

S. No	Parameters	Time	Midazolam+ propofol+ isoflurane Anaesthetic combinations
1	Haemoglobin (g/dl)	Before	10.15 \pm 0.75
		0 Min	10.48 \pm 0.62
		30 Min	09.35 \pm 0.86*
		60 Min	09.23 \pm 0.90*
		24 Hr	09.20 \pm 0.62**
2	Packed Cell Volume (%)	Before	35.48 \pm 2.83
		0 Min	33.15 \pm 2.93
		30 Min	30.40 \pm 2.96*
		60 Min	28.40 \pm 3.25**
		24 Hr	30.42 \pm 2.17*
3	Total Erythrocyte Count ($\times 10^6$ / μ l)	Before	7.62 \pm 0.53
		0 Min	7.26 \pm 0.54
		30 Min	6.79 \pm 0.52**
		60 Min	6.49 \pm 0.56**
		24 Hr	7.11 \pm 0.40*
4	Total Leucocyte Count ($\times 10^3$ / μ l)	Before	11.72 \pm 1.49
		0 Min	09.63 \pm 0.94
		30 Min	08.20 \pm 0.57* ^b
		60 Min	08.77 \pm 0.56*
		24 Hr	10.12 \pm 0.84

Means bearing superscript differ significantly at $P < 0.05$ from interval 'before' within the group

Means bearing superscript differ significantly at $P < 0.01$ from interval 'before' within the group a, b means bearing superscript a, b differ significantly at $P < 0.05$ level between groups at corresponding intervals

Table 2: Mean \pm SE of Differential leucocyte count (%) at different intervals in all the animals

S. No	Parameters	Time	Midazolam+ propofol+ isoflurane Anaesthetic combinations
1	Neutrophils (%)	Before	32.40 \pm 1.33
		0 Min	34.10 \pm 1.47
		30 Min	41.30 \pm 1.37**
		60 Min	42.43 \pm 1.47***
		24 Hr	40.10 \pm 2.12***
		48 Hr	32.33 \pm 2.13
2	Lymphocytes (%)	Before	61.83 \pm 1.54
		0 Min	59.43 \pm 1.76*
		30 Min	50.10 \pm 2.28***
		60 Min	47.23 \pm 1.83***
		24 Hr	53.27 \pm 2.14***
		48 Hr	61.00 \pm 2.19

Means bearing superscript differ significantly at $P \leq 0.05$ from interval 'before' within the group

Means bearing superscript differ significantly at $P \leq 0.01$ from interval 'before' within the group

Means bearing superscript differ significantly at $P \leq 0.001$ from interval 'before' within the group

a, b means bearing superscript a, b differ significantly at $P \leq 0.05$ level between groups at corresponding intervals

Table 3: Mean \pm SE values of biochemical parameters at different intervals in all the animals

S. No	Parameters	Time	Group-II
1	Alanine Transaminase (IU/L)	Before	24.50 \pm 2.71
		0 Min	23.67 \pm 1.45
		30 Min	23.67 \pm 2.19
		60 Min	26.17 \pm 2.68
		24 Hr	27.83 \pm 3.74 ^b
2	Aspartate Transaminase (IU/L)	Before	69.00 \pm 11.60 ^b
		0 Min	73.83 \pm 10.11
		30 Min	74.17 \pm 11.06
		60 Min	91.00 \pm 11.97
		24 Hr	94.20 \pm 14.98*
3	Creatinine (mg/dl)	Before	1.73 \pm 0.16
		0 Min	1.74 \pm 0.16
		30 Min	1.89 \pm 0.14
		60 Min	1.81 \pm 0.07
		24 Hr	1.82 \pm 0.25
4	Serum Urea Nitrogen (mg/dl)	Before	27.63 \pm 3.64 ^b
		0 Min	27.27 \pm 4.15 ^b
		30 Min	27.95 \pm 3.74 ^b
		60 Min	31.15 \pm 4.47 ^b
		24 Hr	26.60 \pm 2.26

Means bearing superscript differ significantly at $P \leq 0.05$ from interval 'before' within the group

Means bearing superscript differ significantly at $P \leq 0.01$ from interval 'before' within the group a, b means bearing superscript a, b differ significantly at $P \leq 0.05$ level between groups at corresponding

6. References

- Abboud TK, Zhu J, Richardson M, Peres DE, Donovan M. Intravenous propofol versus thiomylal – isoflurane for caesarean section, comparative maternal and neonatal effects. *Acta Anesthesia Scandinavia*. 1995; 39(2):205-209.
- Abu-Ahmed, H., Sedative and haemato biochemical effects of midazolam and midazolam-ketamine combination in Baladi goats. *Global Veterinaria*. 2013; 10(6):742-747.
- Adams HK, Briggs LP, Bahar M, Douglai EJ, Dundee JW. Pharmacokinetic evaluation of ICI 35868 in man, single induction doses with different rates of injection. *British Journal of Anesthesia*, 1993; 55:97-103.
- Ajadi RA, Olusa TA, Adeniye SB. Comparative effects of xylazine and acepromazine on some haematological parameters and serum electrolytes in dogs. *Indian J Vet. Surg.* 2008; 29(1):45-46.
- Akhare SB, Pawshe DB, Mehsare SP, Joshi MV, Mode SG. Biochemical effects of ketamine with premedication of diazepam, haloperidone and acepromazine in goats. *J. Vet. Surg.* 2003. 24(2):99-100.
- AL-Redah. A comparative study between using of midazolam-ketamine and diazepam-ketamine combinations as anesthetic program in sheep. *Al. Qadisiya J of Vet. Med. Sci.* 2011; 10(1):66-72.
- Amandeep K, Singh SS. Clinical effect of midazolam-ketamine and midazolam-thiopentone anesthesia in bovines. *Indian J Vet. Surg.* 2004; 25(2):80-82.
- Amarpal P, Kinjavadekar, Aithal HP, Rekha Patak, Virendra Singh, Pratap K. Propofol with and without xylazine and medetomidine for general anesthesia in goats. *Indian J Vet. Surg.* 2000; 21(2):115.
- Ambrisko TD, Hikasa Y. Neurohormonal and metabolic effects of medetomidine compared with xylazine in Beagle dogs. *Can. J Vet. Res.* 2002; 66:42-49.
- Arai S, Yoshioka K, Suzuki C, Takahashi H, Itoh T, Nakano S *et al.* Development of a neurosurgical operating table for adult cattle and changes in intracranial pressure and blood pressure in adult cattle undergoing long-time isoflurane anaesthesia. *J Vet. Med. Sci.* 2006; 68(4):337-43.
- Baniadam A, Afshar FA, Balani AR. Cardiopulmonary effects of acepromazine-ketamine administration in sheep. *Bull. Vet. Inst. Pulawy.* 2007; 51:93-96.
- Bayan HS, Sarma KK, Thomas S. Studies on midazolam propofol anesthesia in canines. *The J Anim. Sci.* 2007, 77(5).
- Benjamin MM. *Outline of Veterinary Clinical Pathology*, 3rd Edn. Iowa State University Press, Iowa, USA, 1985, 64-75.
- Bishnoi P, Saini NS. Cardio-respiratory effects of midazolam in calves. *Vet. Pract.* 2005; 6(1):1-5.
- Brearley JC, Kellagher REB, Hall LW. Propofol anesthesia in Cats. *J Small Anim. Pract.* 1988; 29:315-322.
- Brett CM, Teitel DF, Heymann MA, Rudolph AM. The cardiovascular effects of isoflurane in lambs. *Anesthesiology.* 1987; 67(1):60-5.
- Brezeski W, Depta A, Jalynski M, Chyczewski M. General anaesthesia in sheep with the use of Diprivan (Propofol). *Vet. Bulletin.* 1994; 64:6546.
- Bryant CE, England GC, Clark KW. Comparison of sedative effects of medetomidine and xylazine in horses. *Vet. Rec.* 1991; 129:421-423.
- Butola V, Singh B. Physiological and clinical effects of midazolam and ketamine in dogs. *Indian Vet. J.* 2003; 24(2):95-96.
- Butola V, Singh B. Midazolam as tranquilizer in dogs. *Indian Vet. J.* 2007; 84:1141-1145.
- Cantalapiedra AG, Villanueva B, Pereira JL. Anesthetic potency of isoflurane in cattle: determination of the minimum alveolar concentration. *Vet. Anaesth. Analg.* 2000; 27(1):22-26.
- Carroll GL, Hartsfield SM. General anaesthetic techniques in ruminants. *Vet. Clin. North Am. Food Anim. Pract.* 1996; 12:627-661.
- Chandrasekhar EL, Jaipal R, Hargopal V. Clinical and physiological changes of ketamine-xylazine anaesthesia in buffalo calves. *Indian J Vet. surg.* 2003; 24(2):97-98.
- Cockshot ID, Briggs LP, Douglas EJ, White M. Pharmacokinetics of propofol in female patients. *Bri. J Anaesthesia.* 1987; 59:1103-1110.
- Correia D, Nolan AM, Reid J. Pharmacokinetics of propofol infusion, either alone or with ketamine in sheep, premedicated with acepromazine and papavertum. *Res. Vet. Sci.* 1996; 60:213-217.
- Correia D, Reid J, Nolan AM. The Pharmacodynamics and pharmacokinetics of propofol and propofol-ketamine infusions in sheep. *Proceedings of the 6th International Congress of the European Association of Veterinary Pharmacology and Toxicology* Ed. Lees, P. Blackwell Scientific, London, 1994, 32-33.
- Dzikiti TB, Stegmanna GF, Hellebrekers LJ. Effect of midazolam on isoflurane minimum alveolar concentration in goats. *Small Ruminant Research.* 2011; 97(1-3):104-109.
- Dzikiti TB, Stegmanna GF, Hellebrekers LJ, Auerc RE, Dzikitid LN. Sedative and cardiopulmonary effects of acepromazine, midazolam, butorphanol, acepromazine-butorphanol and midazolam-butorphanol on propofol anaesthesia in goats. *S. Afr. Vet. Ver.* 2009; 80(1):10-16.
- Edmond, Eger. *Isoflurane-A Review.* *Anesthesiology.* 1981; 55:559-576.

30. England GC, Clark KW. Alpha 2 adrenoceptor agonists in the horse. *Br. Vet. J.* 1996; 152:641-657.
31. Glen JB, Hunter SC. Pharmacology of an emulsion formation of ICI 35868. *Bri. J Anaesthesia.* 1984; 56:617-625.
32. Goodman NW, Black AMS, Carter JA. Some ventilator effects of propofol as a sole anaesthetic agent. *Bri. J Anaesthesia.* 1987; 59:1497-1503.
33. Hall, Clarke. *Veterinary Anaesthesia*, 8th Edn. E.L.B.S. Bailliere Tindall London, 1983, 25-30.
34. Hall LW, Chambers JP. A clinical trial of propofol infusion anaesthesia in dogs. *J Small Anim. Pract.* 1987; 28:623-637.
35. Hall LW, Clarke KW, Trim CM. *Veterinary Anaesthesia*. 10th Edn, Saunders, Harcourt Publishers Limited, USA, 2001, 127-129.
36. Haskins SC, Patz JD, Farver TB. Xylazine and xylazine ketamine in dogs. *Am. J Vet. Res.* 1986; 47:636-641.
37. Haskins SC, Farver TB, Patz JD. Ketamine in dogs. *Am. J Vet. Res.* 1985; 46(9):1855-1860.
38. Hikasa Y, Saitob K, Takaseb K, Ogasawara S. Clinical, cardiopulmonary, haematological and serum biochemical effects of sevoflurane and is of luurane anesthesia in oxygen under spontaneous breathing in sheep. *Small Ruminant Research.* 2000; 36:241-249.
39. Hsu WH. Xylazine induced depression and its antagonism by alpha adrenergic blocking agents. *J Pharmacol. Exp. Ther.* 1981; 218(1):188-192.
40. Hsu WH, Lu ZX, Hembrough FB. Effect of xylazine on heart rate and arterial blood pressure in conscious dogs, as influenced by atropine, 4-aminopyridine, doxapram, and yohimbine. *J Am. Vet. Med. Assoc.* 1985; 186:153-156.
41. Ilback NG, Stalhandske T. Cardiovascular effects of xylazine recorded with telemetry in the dogs. *J Vet. Med. Assoc.* 2003; 50:479-483.
42. Ilkiw E, Pascoe PJ, Haskins SC, Partz JD. Cardiovascular & respiratory effects of propofol administration in hypovolemic dogs. *Am. J Vet. Res.* 1992; 53(12):2323-2327.
43. Jain NC. *Schalm's Veterinary Haematology*, 5th Edn. Lea and Febiger, Philadelphia, USA, 2000.
44. James R, Glen JB. Synthesis, biological evaluation and preliminary structural activity of a series of alkyl phenols as an intravenous anaesthetic agents. *J Med. Chem.* 1980; 23:1350-1357.
45. Jones DJ, Stehling LC, Zauder IL. Cardiovascular responses to diazepam and midazolam maleate in the dog. *Anesthesiology.* 1979; 51:430-434.
46. Karen JW, Eugene PS, Neil HW, Michael JW. Recovery of horses from inhalation anesthesia. *Am. J Vet. Res.* 1993; 54(10):1693-1702.
47. Kastner SB. Alpha2-agonists in sheep. *Vet. Anaesth. Analg.* 2006; 33:79-96.
48. Kastner SB, Von Rechenberg B, Keller K, Bettschart Wolfensberger R. Comparison of medetomidine and dexmedetomidine as premedication in isoflurane anaesthesia for orthopaedic surgery in domestic sheep. *J. Vet. Med. Assoc. Physiol. Pathol. Clin. Med.* 2001; 48:231-241.
49. Kelawala NH, Parasania RR, Patil DB. Clinical evaluation of propofol-ketamine anaesthesia in diazepam pre-medicated goats. *Indian J Vet. Surg.* 1993; 14(2):83-85.
50. Kelawala NH, Parasania RR, Patil DB. Haematological and biochemical studies on ketamine, propofol and propofol-ketamine as general anaesthesia in diazepam pre-medicated goats. *Indian J Vet. Surg.* 1991; 12(1):17-20.
51. Kerr DD, Jones EW, Huggins K. Sedative and other effects of xylazine given intravenously to horses. *Am. J Vet. Res.* 1972; 33:525-532.
52. Klide AM, Calderwood HW, Soma RL. Cardiopulmonary effect of xylazine in dogs. *Am. J Vet. Res.* 1975; 36:931-935.
53. Komar E, Silmanowicz P, Balicki I. Effects of propofol anaesthesia on gas exchange & haematological parameters in dogs. *Vet. Bulletin.* 1993; 63:4173.
54. Kumar SS, Dharmaceelan S, Selvarju P, Subramaniyamand M, Rajendran N. Isoflurane uptake in cattle - report of 18 cases. *Proceedings of XXXVI Annual Congress of ISVS and International Symposium*, 2012, 42.
55. Kumar SS, Rajendran N, Dharmaceelan S, Kathirvel S, Subramanian M, Selvaraj P *et al.*, Effect of butorphanol and buprenorphine on inhalant sparing and gas concentrations during low flow isoflurane anaesthesia in cattle. *Adv. Anim. Vet. Sci.* 2013; 1(2S):29-32.
56. Lele CM, Bhokre AP. Evaluation of xylazine as an anaesthetic agent in combination with certain pre-anaesthetic drugs in dogs. *Indian Vet. J.* 1985a; 62:675-682.
57. Lele CM, Bhokre AP. Evaluation of xylazine as an anaesthetic agent in combination with certain pre-anaesthetic drugs in dogs. *Indian Vet. J.* 1985b; 62:863-868.
58. Lele CM, Bhokre AP. Evaluation of xylazine as an anaesthetic agent in combination with certain pre-anaesthetic drugs in dogs. *Indian Vet. J.* 1985c; 62:1039.
59. Singh GD, Kinjavdekar P, Amarpal Aithal HP, Pawde AM, Zama MM. Clinicophysiological and haemodynamic effects of fentanyl with xylazine, medetomidine and dexmedetomidine in isoflurane-anesthetized water buffaloes (*Bubalus bubalis*). *JS. Afri. Vet. Assoc.* 2013; 84(1):67-77.
60. Topal A, Gul N, Ilcol Y, Gorgul OS. Hepatic effects of halothane, isoflurane or sevoflurane anesthesia in dogs. *J Vet. Med. Assoc.* 2003; 50(10):530-533.
61. Tranquilli WJ, Thurmon JC, Corbin JE, Benson JE, Davis LE. Halothane-sparing effect of xylazine in dogs and subsequent reversal with tolazoline. *J Vet. Pharmacol. Ther.* 1984; 7:23-28.
62. Vesal N, Sarchahi AA, Nikalival B, Karampour A. Clinical evaluation of the sedative properties of acepromazine-xylazine in dogs. *Vet. Arhiv.* 2011; 81(4):485-498.
63. Vescae G, Lucisano A. Crossing of placental barrier by propofol in small ruminants. *Vet. Bulletin*, 63 Abst, 1991, 6825.
64. Wagner AE, Muir WW, Hinchcliff KW. Cardiovascular effects of xylazine and detomidine in horses. *Am. J Vet. Res.* 1991; 52:651-657.
65. Waterman AE. Use of propofol in sheep. *Vet. Rec.* 1988; 12:26.
66. Watkins SB, Hall LW, Clarke KW. Propofol as intravenous anaesthetic agent in dogs. *Vet. Rec.* 1987; 120:326-329.
67. Weaver BMQ, Raptopoulos. Induction of anaesthesia in dogs & cats with propofol. *Vet. Rec.* 1990; 126:617-62.