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# Endophytes, promising option for pest management in agriculture: Review

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#### Abstract

Endophytes are the group of minute, microscopic organisms invisible to naked eye normally establish their close relationships with the plant. The relationship may be harmful or beneficial. Some of them complete entire part of their life cycle or some passes at least one part of their life cycle within the plant. By the perusal literature it has been proved that they can offer the greatest opportunity for taking the management strategies against the insect pest and disease alternative to agrochemicals. Their application in plant protection field is ecofriendly and indicates the long term sustainability of ecosystems. Keeping these views in mind, the present article has been written to highlight their contribution in pest and disease management in modern agriculture field.

Keywords: Endophyte, bacterial endophyte, fungal endophyte, insect pest, disease

#### Introduction

Now a day the modern agriculture system is continuously facing a tremendous challenge against the increasing world population and with the continuous supply of food commodities. To sustain in the competition and make the agriculture more profitable it is manadatory to reduce the cost of cultivation and to maintain the pollution free long term sustainable environment. To shift on this philosophy, there is a need for searching the alternative option in place of the use of agrochemicals. The plant growth and its defence mechanism was enhanced by the use of beneficial microorhanisms viz. Fungi and bacteria and this result was popularize for the opening a wide scope to expose and use of the plant associated microorganisms <sup>[45]</sup>. These plant associated tiny organisms were termed as Endophyte and they broadly categorized into two main groups' viz. Bacteria and Fungus<sup>[5]</sup>. Studies on the life cycle of endophytes suggested that they can able entire life cycle or a single part of their life cycle within the plant system <sup>[3]</sup>. Extensive research work on the bioecology, function and survivability of endophytes reveal that approximately 3,00,000 plant species show their intimate association with the endophytes <sup>[60]</sup> in different plant parts viz. leaves, stems, branches <sup>[23]</sup>. Besides it is also true that their interaction with their respective host plant was solely depen upon on their host plant habitat <sup>[16]</sup>. Endophytes are classified as harmful or beneficial depending on their interaction with their host plant. Many biologists throughout the globe have documented the different beneficial functions of endophytes. They help the plant by enhancing the capacity of essential nutrient elements including both macro and micro from soil environment [80, 40, 46, 6], provide the protection of the plant from major insect pest and disease [58, 20, 76], provide the major contribution in promoting of the plant growth and development [74, 24]. It has been reported that endophytes take part in production of several important secondary metabolic compounds with different mode of actions and sturucture which includes viz. alkaloids, benzopyranones, flavonoids, phenolic acids, quinones, steroids, terpenoids, tetralones and xanthones which showed excellent performance against different destructive plant pathogens <sup>[2]</sup>. Their association with the field crop make the crop more tolerant against the notorious weeds and suppress the weed floral population in field condition. Due to supression of weed problem the plant can able to utilize the natural resources in a better way <sup>[81]</sup>. It has been proved that endophytic association with the plant boost up the tolerance and resistant capacity of plant against abiotic and biotic stresses <sup>[49, 24]</sup>. Extensive studies on endophytes showed that they are very much promising and helpful regarding the enhancement of yield attributing characters of crop <sup>[21, 68]</sup>. The role of endophytes in insect and disease management in agriculture field by their antibiosis and antixenosis activity was proved by several scientists and this is the best weapon in plant protection arena in agriculture which leads to reduction of

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the use of plant protection chemicals and make the environment and ecosystem more stable and sustainable for long period <sup>[4]</sup>. There has no uncertainity in future biocontrol will be the major option in place of chemical control. Considering the all information, here the present review article elaborately discussed about the contribution of bacterial and fungal endophytes in managing the biotic stress of crop special emphasize to insect and disease pest.

#### Role of Bacterial endophytes in insect pest management

Bacteria are the prokaryotic unicellular microscopic organisms show the great variation in their body shape and size <sup>[19]</sup>. Bacterial endophytes are the special group of living organisms which survive by the close association with their respective host plant. Besides their survivability they exert the growth promoting nature and defence mechanism to the plant against many insect and deadly pathogens <sup>[59]</sup>. Extensive studies during the last few several decades discovered many beneficial strains of commercially available bacterial endophytes which showed as very promising in suppression of plant disease, promoting the plant growth and development <sup>[34, 41, 1, 31]</sup>. Induced systemic resistance (ISR) is one of the most important phenomena in the plant body which is triggered by when a plant come to contact with an endophyte by production of several bioactive metabolic compounds. The development of ISR ultimately produce Systemic acquired resistance (SAR) when the plant gets infested by any insect or pathogen <sup>[68]</sup>. Among the several groups of bacterial endophytes, Bacillus amyloliquefaciens is considered as one of the most promising entomopathogenic endophyte [60, 11, 31] <sup>[12]</sup> by producing the main metabolic compounds lipopeptide in nature solely responsible for development induced plant resistance against the fall army worm (Spodoptera furgiperda) by reducing the body weight <sup>[9, 36, 15]</sup>. Enterobacter cloacae another endophytic rod shaped and gram negative bacteria were reported as good biocontrol agent against the White Backed Plant Hopper (WBPH) (Sogatella furcifera), important pest of paddy by production of their Pinellia ternate agglutinin (PTA) protein [83]. Eendophytic bacteria Clavibacter xyli subsp. cynodontis was made as transgenic by the incorporation of cry1A (Crystal endotoxin) gene isolated from Bacillus thurigiensis [64] and this transgenic bacteria showed insecticidal activity against many lepidopteran insect pest specially emphasizing to the European corn borer (Ostrinia nubilalis). Diamond back moth (DBM) (Plutella xylostella) is the major yield reducing lepidopteran insect pest in cabbage, cauliflower and broccoli throughout the world <sup>[18]</sup>. Several studies had been carried out for this management aspect of this insect pest by utilizing the bacterial endophytes. The isolates of Enterobacter cloacae (ENF14), Alcaligenes piechaudii (EN5) or K. ascorbata (EN4) were observed to have control action against diamondback moth [55, 61, 62, 63].

**Role of Bacterial endophytes in plant disease management** Plant diseases are characterized by any abnormality in plant leads to hindrance to the plant normal growth and reduce the economical yield. Majority of plant disease are caused by fungus belongs to the Ascomycetes and Zygomycetes group followed by bacteria and virus. For suppressing the plant disease indiscriminate use of chemicals has augmented leads to development of hazardous environment. The endophytc bacteria contribute huge impact on plant growth promotion and induce the resistance power into the plant body by production of their secondary metabolites which was considered as natural biocontrol against the disease [34]. Similarly Bacillus sp. produce secondary metabolic compounds which is peptidic in nature showing antibiosis property against many plant diseases [60]. Lipopeptide compounds with volatile and low molecular weight nature were produced by many endophytic bacteria showed specific antifungal and antibacterial activities [82]. The isolated lipopeptidic compounds were further categorized based on their chemical nature and mode of action on the plant pathogens viz. surfactin, fengycin, polymyxin, bacitracin and iturin <sup>[36, 46]</sup>. Among those compounds Fengycin and iturin are reported as antifungal agents <sup>[51]</sup>. From the research work it was established that Iturin, suppressed the pathogenecity of Pectobacterium carotovorum in potato, carrot and onion and *Xanthomonas campestris* in paddy<sup>[81]</sup>. Fengycin was found to be effective on Botrycis cinerea, causing the grey mold disease in many plants especially in apple <sup>[65]</sup>. Different types of phenazine compounds having antifungal properties like Phenazine-1-carboxamide, phenanazine-1-carboxylic acid and phenanzine-1-carboxamide are released by the Bacillus sp. successfully control Rhizoctonia solani, Xanthomonas oryzae, Pythium myriotylum, Pythium splendens in different agricultural and horticultural plant by exerting their antibiosis property [41, 40, 53]. Plant growth promoting Rhizobacteria (PGPR) normally colonize in the plant rhizospheric zone help to develop induced systemic resistance (ISR) in the plant and also take part in plant growth promotion. One of the major PGPR, Pseudomonas fluorescens strains WCS417R, WCS365 and 89B-61 protect the cucumber plant from their anthracnose disease [68, 25, 69, 76, 26]. Other promising bacterial endophytes are Bacillus amyloliquefaciens, Bacillus pumilus, Bacillus subtilis, Pseudomonas fluorescens, Pseudomonas syringae and Serratia marcescens which were reported as major factor for the development of systemic resistance in the plant against the different diseases <sup>[26]</sup> and also against different plant parasitic nematodes (PPN)<sup>[22]</sup>. From the enzymatic study it has been revealed that endophytic bacteria produced different kind of enzymes like peroxidases, lipoxygenases, chitinases and glucanases having diversified activity [10, 50]. These enzymes are found to be effective against the palnt pathogen by inhibition of their growth and development. Peroxidises enzyme effectively reduced the incidence of damping-off pathogen Pythium aphanidermatum in rice, wheat and cucumber <sup>[80]</sup>. Production of phytoalexins ultimately leads to formation of lipoxynase enzyme showed to be inhibitory action against the incidence of many plant diseases <sup>[30]</sup>. Scientists proved that there was a positive correlationship in between the more production of enzymne and better control of plant disease <sup>[17, 35, 49]</sup>.

#### Role of fungal endophytes in insect pest management

Fungi are the group of eukaryotic, unicellular to multicellular organisms showed wide diversity in their survivability and sustainability. The pioneering work on fungal endophyes was done on *Phomopsis oblonga*, the endophytic fungus gave protection to the Elm trees against the beetle *Physocnemum brevilineum* <sup>[75]</sup>. Later many research workers were focused on the study of insect pest management by using the fungal endophytes. Thereafter, many naturally occurring endophytic fungui like *Beauveria bassiana*, *Clonostachys rosea*, *Isaria farinosa* were isolated from their respective host plant <sup>[7, 13, 42, 70, 37]</sup>. *Beauveria bassiana* was found to be effective against insect pest in many cases. Damage incidence of Poppy stem

gall caused by cynipid gall wasp, Iraella luteipes was reduced by application of *B. Bassiana*<sup>[45]</sup>. It was found to be effective against European corn borer (Ostrinia nubilalis) and Pink borer Sesamia calamistis in maize [8, 28, 14]. Acremonium sp., occurring another important naturally endophytic entomopathogenic fungus enhanced the plant defence property by producing secondary metabolites were reported as effective against many insect species. Lolines, aminopyrrolizidine alkaloids produced by Neotyphodium sp. showed broad spectrum insecticidal activity <sup>[52]</sup>. Peramine, pyrrolopyrazine alkaloid produced by fungal endophytes showed effectiveness against the argentine stem weevil [48]. Another report revealed that Lolines and Permaines were very useful in managing the aphids including Rhopalosiphum padi and *Schizaphis graminum* in cereal crops <sup>[54]</sup>. Besides fighting with biotic stress, fungal endophytes were equally effective against the abiotic stress and help the plant to overcome the stress <sup>[77, 54]</sup>. By application of fungal endophytes in insect pest management in agriculture should be the best alternative in future to protect our environment for long lasting future <sup>[71]</sup>.

## Role of fungal endophytes in plant disease management

Like insect pest management, the fungal endophytes were also exposed against the suppression of many plant diseases by production of their many bioactive metabolic compounds. Cryptocin, tetrameric acid analog from endophytic fungus Cryptosporiopsis quercina was isolated found to be good suppressing agent against the rice blast disease caused by *Pyricularia oryzae*<sup>[29]</sup>. *Colletotrichum* sp. another endophytic fungus was used for the isolation of 6-isoprenvlindole-3carboxylic acid, an Indole derivative compound. This compound was effective against the many deadly plant pathogens like Phytophthora capsici, Rhizoctonia cerealis and Gaeumannomyces graminis var. tritici [32]. Colletotric acid, a phenolic antifungal compound was isolated from Colletotrichum gloeosporioides shown to be effective against Helminthosporium sativum, Bacillus subtilis, Staphylococcus aureus and Sarcina lutea [84]. Chokols, a sesquiterpene compound produced by Epichloe typhina, an found to be antifungal agent to the leaf spot disease caused by *Cladosporium phlei* <sup>[27]</sup>.

#### Conclusion

Today's agriculture is undergoing through a serious challenge due to indiscriminate use of plant protection chemicals for pest and disease management. In the present scenario the use of biocontrol agents got the considerable attention due to their contribution to shift the environment in a stable and sustainable way. Endophytes play the important role in spression of plant disease and insect management. There is a huge scope to explore the role of endophytes in plant protection field for achieving the best alternative way for promoting the healthy and stable ecosystem and environment for the next generation.

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#### References

1. Alfonzo A, Piccolo SL, Conigliaro G, Ventorino V, Burr uano S, Moschetti G *et al.* Antifungal peptides produced by *Bacillus amyloliquefaciens* AG1 active against grapevine fungal pathogens. Annual Review of Microbiology. 2012; 62:1593-1599.

- Aravind R, Kumar A, Eapen SJ, Ramana KV. Endophytic bacterial flora in root and stem tissues of black pepper (*Piper nigrum* L.) genotype: isolation, identification and evaluation against *Phytophthora capsici*. Applied Microbiology. 2009; 48(1):58-64.
- 3. Arnold AE, Mejia LC, Kyllo D, Rojas EI, Maynard Z, Robbins N *et al.* Fungal endophyte limit pathogen damage in a tropical tree. Proceedings of the National Academy of Science, USA, 2003.
- 4. Azevedo JL, Maccheroni Jr W, Pereira JO, de Araújo WL. Endophytic microorganisms: a review on insect control and recent advances on tropical plants. Electronic Journal of Biotechnology. 2000; (3)1:40-65.
- Azevedo JL. Microorganismos endofíticos. In: Ecologia Microbiana. Melo, I.S. and Azevedo, J.L. (edts.). Editora EMBRAPA, Jaguariuna, São Paulo, Brazil, 1998, 117-137.
- 6. Beltrán-García MJ, White JF, Prado FM, Prieto KR, Yamaguchi LF, Torres MS *et al.* Nitrogen acquisition in Agave tequilana from degradation of endophytic bacteria. Scientific Reports. 2014; 4:6938.
- Bills GF, Polishook JD. Microfungi from *Carpinus* caroliniana. Canadian Journal of Botany. 1991; 69:1477-1482.
- Bing LA, Lewis LC. Suppression of Ostrinia nubilalis (Hübner) (Lepidoptera: Pyralidae) by endophytic Beauveria bassiana (Balsamo) Vuillemin. Environmental Entomology. 1991; 20:1207-1211.
- 9. Boning RA, Bultman TL. Atest for constitutive and induced resistance by tall Fescue (*Festua arundinacea*) to an insect herbivore: impact of fungal endophyte *Acremonium coenophialum*. American Midland Naturalist Journal. 1996; 136:328-335.
- Chen C, Belanger RR, Benhamou N, Paulitz TC. Defense enzymes induced in cucumber roots by treatment with plant-growth promoting rhizobacteria (PGPR). Physiological and Molecular Plant Pathology. 2000; 56:13-23.
- 11. Chen XH, Scholz R, Borriss M, Junge H, Mögel G, Kunz S *et al.* Difficidin and bacilysin produced by plant-associated *Bacillus amyloliquefaciens* are efficient in controlling fire blight disease. Journal of Biotechnology. 2009; 140:38-44.
- 12. Chen YT, Yuan Q, Shan LT, Lin MA, Cheng DQ, Li CY *et al.* Antitumor activity of bacterial exopolysaccharides from the endophyte *Bacillus amyloliquefaciens* isolated from *Ophiopogon japonicus*. Oncology Letters. 2013; 5:1787-1792.
- Cherry A, Lomer C, Djegui D, Schulthess F. Pathogen incidence and their potential as microbial control agents in IPM of maize stem borers in West Africa. Biocontrol. 1999; 44:301-327.
- 14. Cherry AJ, Banito A, Djegui D, Lomer C. Suppression of the stem-borer *Sesamia calamistis* (Lepidoptera; Noctuidae) in maize following seed dressing, topical application and stem injection with African isolates of *Beauveria bassiana*. International Journal of Pest Management. 2004; 50:67-73.
- Choudhary DK, Johri BN. Interactions of *Bacillus* spp. And plants with special reference to induced systemic resistance (ISR). Microbiological Research. 2009; 164:493-513.

- 16. Compant S, Saikkonen K, Mitter B, Campisano A, Mercado-Blanco J. Editorial special issue: soil, plant and endophytes. Plant and Soil. 2016; 405(1):1-11.
- 17. Daniel M, Purkayastha RP. Handbook of phytoalexin metabolism and action. New York: Marcel Dekker, 1995, 615.
- Dickson MH, Shelton AM, Elgenbrode SD, Vamosy ML, Mora M. Selection for resistance to diamondback moth (*Plutella xylostella*) in cabbage. Horticultural Science. 1990; 25(12):1643-1646.
- Fredrickson JK, Zachara JM, Balkwill DL, Kennedy D, Li SM, Kostandarithes HM *et al.* Geomicrobiology of high-level nuclear waste-contaminated vadose sediments at the Hanford site, Washington State. Applied and Environmental Microbiology. 2004; 70(7):4230-41.
- Gond SK, Bergen M, Torres MS, White JF. Effect of bacterial endophyte on expression of defense genes in Indian popcorn against *Fusarium moniliforme*. Symbiosis. 2015; 66:133-140.
- Govindarajan M, Balandreau J, Kwon S-W, Weon H-Y, Lakshminarasimhan C. Effects of the inoculation of *Burkholderia vietnamensis* and related endophytic diazotrophic bacteria on grain yield of rice. Microbial Ecology. 2008; 55:21-37.
- 22. Hallmann J, Kloepper J, Rodriguez-Kabana R, Sikora RA. Endophytic rhizobacteria as antagonists of *Meloidogyne incognita* on cucumber. Phytopathology. 1995; 85:1136.
- Hardoim PR, van Overbeek LS, Berg G, Pirtilla AM, Compant S, Campisano A *et al.* The hidden world within plants: ecological and evolutionary considerations for defining functioning of microbial endophytes. Microbiology and Molecular Biology Reviews. 2015; 79(3):293-320.
- 24. Irizarry I, White JF. *Bacillus amyloliquefaciens* alters gene expression, ROS production, and lignin synthesis in cotton seedling roots. Journal of Applied Microbiology. 2018; 124:1589-1603.
- 25. Kamilova F, Validov S, Azarova T, Mulders I, Lugtenberg B. Enrichment for enhanced competitive plant root tip colonizers selects for a new class of biocontrol bacteria. Environmental Microbiology. 2005; 7:1809-1817.
- Kloepper JW, Ryu CM. Bacterial endophytes as elicitors of induced systemic resistance. In B. Schulz, C. Boyle, & T. Sieber (Eds.), Microbial root endophytes. Berlin/ Heidelberg: Springer, 2006, 33-52.
- 27. Koshino H, Togia S, Terada SI, Yoshihara T, Sakamura S, Shimanuki *et al.* New fungitoxic sesquiterpenoids, chokols A-G, from stromata of Epichloe typhina and the absolute configuration of chokol. Agricultural and Biological Chemistry. 1989; 53:789-796.
- 28. Lewis LC, Bruck DJ, Gunnarson RD, Bidne KG. Assessment of plant pathogenicity of endophytic *Beauveria bassiana* in Bt transgenic and non-transgenic corn. Crop Science. 2001; 41:1395-1400.
- 29. Li JY, Strobel G, Harper J, Lobkovsky E, Clardy J. Cryptocin, a potent tetramic acid antimycotic from the endophytic fungus *Cryptosporiopsis* cf. *quercina*. Organic Letters. 2000; 2:767-770.
- 30. Li WX, Kodama O, Akatsuka T. Role of oxygenated fatty acids in rice phytoalexin production. Agricultural and Biological Chemistry. 1991; 55:1041-1147.
- 31. Li HY, Shen M, Zhou ZP, Li T, Wei YL, Lin LB et al.

Diversity and cold adaptation of endophytic fungi from five dominant plant species collected from the Baima Snow Mountain, Southwest China. Fungal Diversity. 2012; 54:79-86.

- 32. Lu H, Xou WX, Meng JC, Hu J, Tan RX. New bioactive metabolites produced by *Colletotrichum* sp., an endophytic fungus in *Artemisia annua*. Plant Science. 2000; 151:67-73.
- Mei C, Flinn BS. The use of beneficial microbial endophytes for plant biomass and stress tolerance improvement. Recent Patents on Biotechnology. 2010; 4:81-95.
- Muthukumar A, Udhayakumar R, Naveenkumar R. Endophytes: Crop Productivity and Protection, 2017, 133-161.
- 35. Nakkeeran S, Kavitha K, Chandrasekar G, Renukadevi P, Fernando WGD. Induction of plant de-fence compounds by *Pseudomonas chlororaphis* PA 23 and Bacillus subtilis BSCBE 4 in controlling damping-off of hot pepper caused by Pythium aphanidermatum. Bio-control Science and Technology. 2006; 16:403-416.
- 36. Ongena M, Jacques P. Bacillus lipopeptides: versatile weapons for plant disease biocontrol. Trends in microbiology. 2008; 16:115-125.
- Orole O, Adejumo T. Activity of fungal endophytes against four maize wilt pathogens. African Journal of Microbiology Research. 2009; 3:969-973.
- 38. Paungfoo-Lonhienne C, Rentsch D, Robatzrk S, Webb RI, Sagulenko E, Nasholm T *et al.* Turning the table: plants consume microbes as a source of nutrients. Plosone, 2010, 5.
- Pérez-García A, Romero D, Vicente A. Plant protection and growth stimulation by microorganisms: biotechnological applications of Bacilli in agriculture. Current Opinion in Biotechnology. 2011; 22:187-193.
- Perneel M, D'Hondt L, De Maeyer K, Adiobo A, Rabaey K, Hofte M *et al.* Phenazines andbiosurfactants interact in the biological control of soil-borne diseases caused by *Pythium* spp. Environmental Microbiology. 2008; 10:778-788.
- 41. Pierson LS, Thomashow LS. Cloning of heterologous expression of phenazinebiosynthesis locus from *P. aureofaciens* 30–84. Molecular Plant-Microbe Interactions. 1992; 53:330-339.
- 42. Pimentel IC, Glienke-Blanco C, Gabardo J, Stuart RM, Azevedo JL. Identification and colonization of endophytic fungi from soybean (*Glycine max* (L.) Merril) under different environmental conditions. Brazilian Archives of Biology and Technology. 2006; 49:705-711.
- 43. Pineda A, Loon JAV, Pieterse C. Helping plants to deal with insects: The role of beneficial soil-borne microbes. Trends in Plant Science. 2010; 15(9):507-514.
- 44. Prieto KR, Echaide-Aquino F, Huerta-Robles A, Valerio HP, Macedo-Raygoza G, Prado FM *et al.* Endophytic bacteria and rare earth elements; promising candidates for nutrient use efficiency in plants, in Plant Macronutrient Use Efficiency, ed. by Hossain M, Kamiya T, Burritt D, Tram L-SP and Fujiwara T. Academic Press, Cambridge, MA, 2017, 285-302.
- 45. Quesada-Moraga E, Munoz-Ledesma F, Santiago-Alvarez C. Systemic protection of *Papaver somniferum* L. against *Iraella luteipes* (Hymenoptera: Cynipidae) by an endophytic strain of *Beauveria bassiana* (Ascomycota: Hypocreales). Environmental Entomology.

2009; 38:723-730.

- 46. Ramkumar G, Yu SM, Lee YH. Influence of light qualities on antifungal lipopeptide synthesis in *Bacillus amyloliquefaciens* JBC36. European Journal of Plant Pathology. 2013; 137:243-248.
- 47. Redman RS, Sheehan KB, Stout RG, Rodriguez RJ, Henson JM. Thermotolerance generated by plant/fungal symbiosis. Science. 2002; 298:1581.
- Rowan DD, Gaynor DL. Isolation of feeding deterrents against argentine stem weevil from ryegrass infected with the endophyte *Acremonium loliae*. Journal of Chemical Ecology. 1986; 12:647-658.
- 49. Saikia R, Kumar R, Arora DK, Gogoi DK, Azad P. *Pseudomonas aeruginosa* inducing rice resistance against *Rhizoctonia solani*: Production of salicylic acid and peroxidases. Folia Microbiologica. 2006; 51:375-380.
- 50. Saikia R, Kumar R, Singh T, Srivastava AK, Arora DK, Gogoi DK *et al.* Induction of defense related enzymes and pathogenesis related proteins in *Pseudomonas fluorescens*-treated chickpea in response to infection by *Fusarium oxysporum* f. sp. *ciceri.* Mycobiology. 2004; 32:47-52.
- 51. Savadogo A, Tapi A, Chollet M, Wathelet B, Traore AS, Jacques P *et al.* Identification of surfactin producing strains in Soumbala and Bikalga fermented condiments using polymerase chain reaction and matrix assistedlaser desorption/ionization-mass spectrometry methods. International Journal of Food Microbiology. 2011; 151:299-306.
- 52. Schulz B, Boyle C, Draeger S, Römmert AK, Krohn K. Endophytic fungi: a source of novel biologically active secondary metabolites. Mycological Research. 2002; 106:996-1004.
- 53. Shanmugaiah V, Mathivanan N, Varghese B. Purification, crystal structure and antimicrobial activity of phenazine-1-carboxamide produced by a growthpromoting biocontrol bacterium, *Pseudomonas aeruginosa* MML2212. Journal of Applied Microbiology. 2010; 108:703-711.
- 54. Siegel MR, Latch GC, Bush LP, Fannin NF, Rowan DD, Tapper BA *et al.* Fungal endophyte-infected grasses: alkaloid accumulation and aphid response. Journal of Chemical Ecology. 1990; 16:3301-3315.
- 55. Silva G, Araújo DV, Costa VSO, Mariano RLR, Barros R, Gomes AMA *et al.* Potencial de bactérias endofíticas para controle biológico da traça das crucíferas (*Plutella xylostella* L., 1758) (Lepidoptera: Plutellidae) em repolho. In: Mota, R.A., Faustino, M.A.G. & Almeida, M.G.A. (Eds.). Proceedings, IX Congresso de Iniciação Científica, Recife, PE, 1999, 70.
- 56. Soares MA, Li H, Bergen M, White JF. Functional role of an endophytic *Bacillus amyloliquefaciens* in enhancing growth and disease protection of invasive English ivy (*Hedera helix* L.). Plant and Soil. 2015; 405:107-123.
- 57. Stone JK, Bacon CW, White JF. An overview of endophytic microbes: endophytism defined. In: Bacon CW, White JF, editors. Microbial endophytes. New York: Marcel-Dekker, 2000, 3-30.
- 58. Strobel GA. Endophytes as sources of bioactive products. Microbes and Infection. 2003; 5:535-544.
- 59. Sun L, Lu Z, Bie X, Lu F, Yang S. Isolation and characterization of a co-producer of fengycins and surfactins, endophytic *Bacillus amyloliquefaciens* ES-2, from *Scutellaria baicalensis* Georgi. World Journal of

Microbiology and Biotechnology. 2006; 22:1259-1266.

- Tabbene O, Slimene IB, Bouabdallah F, Mangoni ML, Urdaci MC, Limam F *et al.* Production of antimethicillin-resistant staphylococcus activity from *Bacillus subtilis* sp. strain B38 newly isolated from soil. Applied Biochemistry and Biotechnology. 2009; 157:407-419.
- Thuler RT, Barros R, Mariano RLR, Vendramim JD. Efeito de bactérias promotoras do crescimento de plantas (BPCP) no desenvolvimento de *Plutella xylostella* (L.) (Lepidoptera: Plutellidae) em couve. Científica. 2006; 34(2):217-222.
- 62. Thuler RT, Bortoli SA, Hoffmann-Campo CB. Classificação de cultivares de brássicas com relação à resistência à traçadas-crucíferas e à presença de glucosinolatos. Pesquisa Agropecuária Brasileira. 2007; 42(4):467-474.
- 63. Thuler RT. Criação de *Plutella xylostella* (L.). In: Criação de Insetos: da base à biofábrica. ed. Jaboticabal: O autor, 2009, 58-68.
- 64. Tomasino SF, Leister RT, Dimock MB, Beach RM, KellyJL. Field Performance of *Clavibacter xyli* subsp. *Cynodontis* expressing the Insecticidal Protein Gene *cryIA*(c) of *Bacillus thuringiensis* against European Corn Borer in Field Corn. Biological Control. 1995; 5(3):442-448.
- 65. Toure Y, Ongena M, Jacques P, Guiro A, Thonart P. Role of lipopeptides produced by Bacillus subtilis GA1 in the reduction of grey mould disease caused by *Botrytis cinerea* on apple. Journal of Applied Microbiology. 2004; 96:1151-1160.
- 66. Uppala S, Beena S, Chapala M, Bowen KL. Amaranth endophytes and their role in plant growth promotion. In: Reddy MS, Desai S, Sayyed RZ, Rao VK, Sarma YR, Reddy BC, Reddy KRK, Podile AR, Kloepper JW (eds) Plant growth promotion by Rhizobacteria for sustainable agriculture. Scientific Publishers, Jodhpur, 2010, 531-537.
- 67. Van Loon LC, Bakker PA, Pieterse CMJ. Systemic resistance induced by rhizosphere bacteria. Annual Review of Phytopathology. 1998; 36:453-483.
- Van Loon LC, Bakker PAHM. In H. De Kroon & V. WJW (Eds.) Root ecology. Berlin: Springer, 2003, 297-330.
- 69. Van Wees SCM, Van der Ent S, Pieterse CMJ. Plant immune responses triggered by beneficial microbes. Current Opinion in Plant Biology. 2008; 11:443-448.
- 70. Vega FE, Posada F, Aime MC, Pava-Ripoll M, Infante F, Rehner SA *et al.* Entomopathogenic fungal endophytes. Biological Control. 2008; 46:72-82.
- 71. Vega FE. The use of fungal entomopathogens as endophytes in biological control: A review Mycologia. 2018; 110(1):4-30.
- 72. Verma SK, Kingsley K, Bergen M, English C, Elmore M, Kharwar RN *et al.*, Bacterial endophytes from rice cut grass (*Leersia oryzoides* L.) increase growth, promote root gravitropic response, stimulate root hair formation, and protect rice seedlings from disease. Plant and Soil. 2017; 422:223-238.
- 73. Verma SK, Kingsley K, Irizarry I, Bergen M, Kharwar RN, White JF *et al.* Seed vectored endophytic bacteria modulate development of rice seedlings. Journal of Applied Microbiology. 2017; 122:1680-1691.
- 74. Verma SK, Kingsley KL, Bergen MS, Kowalski KP,

White JF. Fungal disease protection in rice (*Oryza sativa*) seedlings by growth promoting seed-associated endophytic bacteria from invasive *Phragmites australis*. MDPI: Microorganisms, 2018, 6.

- 75. Webber J. A natural control of dutch elm disease. Nature. 1981; 292:449.
- 76. Wei G, Kloepper JW, Tuzan S. Induction of systemic resistance of cucumber to *Colletotrichum orbiculare* by select strains of plant growth-promoting rhizobacteria. Phytopathology Journal. 1991; 81:1508-1512.
- 77. West CP. Physiology and drought tolerance of endophyte-infected grasses. In: Bacon C. W. & White J. F. (Eds), Biotechnology of endophytic fungi of grasses. Boca Raton, FL: CRC Press, 1994, 87-99.
- 78. White JF, Crawford H, Torres MS, Mattera R, Irizarry I, Bergen MA *et al.* Proposed mechanism for nitrogen acquisition by grass seedlings through oxidation of symbiotic bacteria. Symbiosis. 2012; 57:161-171.
- 79. White JF, Kingsley KL, Kowalski KP, Irizarry I, Micci A, Soares MA *et al.* Disease protection and allelopathic interactions of seed-transmitted endophytic pseudomonads of invasive seed grass (*Phragmites australis*). Plant and Soil. 2017; 422:195-208.
- Young SA, Guo A, Guikema JA, White F, Leach IE. Rice cationic peroxidase accumulation in xylem vessels during incompatible interaction with *Xanthomonas oryzae*. Plant Physiology. 1995; 107:1333-1341.
- 81. Zeriouh H, Romero D, García-Gutiérrez L, Cazorla FM, de Vicente A, Pérez-García A *et al.* The iturin-like lipopeptides are essential components in the biological control arsenal of *Bacillus subtilis* against bacterial diseases of cucurbits. Molecular Plant-Microbe Interactions. 2011; 24:1540-1552.
- 82. Zhang X, Li B, Wang Y, Guo Q, Lu X, Li S *et al.* Lipopeptides, a novel protein, and volatile compounds contribute to the antifungal activity of the biocontrol agent *Bacillus atrophaeus* CAB-1. Applied Microbiology and Biotechnology. 2013; 97:9525-9534.
- 83. Zhang XF. Insecticidal effect of recombinant bacterium containing *Pinellia ternate* agglutinin against *Sogatella furcifera*. Crop Protection. 2011; 30:1478-1484.
- Zou WX, Meng JC, Lu H, Chen GX, Shi GX, Zhang TY et al. Metabolites of Colletotrichum gloeosporioides, an endophytic fungus in *Artemisia mongolica*. Journal of Natural Products. 2000; 63:1529-1530.