



E-ISSN: 2320-7078

P-ISSN: 2349-6800

www.entomoljournal.com

JEZS 2020; 8(2): 1092-1097

© 2020 JEZS

Received: 04-01-2020

Accepted: 06-02-2020

Pushpa M Makwana

Assistant Professor, Department of Veterinary Microbiology, College of Veterinary Science & A.H., Navsari Agricultural University, NAU, Navsari, Gujarat, India

IH Kalyani

Professor & Head, Department of Veterinary Microbiology, College of Veterinary Science & A.H., Navsari Agricultural University, NAU, Navsari, Gujarat, India

Dhruv Desai

Ph.D. Research Fellow, Department of Veterinary Microbiology, College of Veterinary Science & A.H., Navsari Agricultural University, NAU, Navsari, Gujarat, India

JM Patel

Associate Professor, Department of Veterinary Pathology, College of Veterinary Science & A.H., Navsari Agricultural University, NAU, Navsari, Gujarat, India

JB Solanki

Professor & Head, Department of Veterinary Parasitology, College of Veterinary Science & A.H., Navsari Agricultural University, NAU, Navsari, Gujarat, India

PD Vihol

Associate Professor & Head, Department of Veterinary Pathology, College of Veterinary Science & A.H., Navsari Agricultural University, NAU, Navsari, Gujarat, India

DR Patel

Assistant Professor, Department of Veterinary Microbiology, College of Veterinary Science & A.H., Navsari Agricultural University, NAU, Navsari, Gujarat, India

DM Muglikar

Associate Professor & Head, AKNP College of Veterinary Science Shirwal, Dist. Satara, Maharashtra, India

PS Sakhare

Assistant Professor, RCDP on Cattle, M.P.K.V. Rahuri, Dist. Ahmednagar, Maharashtra, India

Corresponding Author:**Pushpa M Makwana**

Assistant Professor, Department of Veterinary Microbiology, College of Veterinary Science & A.H., Navsari Agricultural University, NAU, Navsari, Gujarat, India

Detection of bovine rotavirus (BRV) infection in neonatal calves of in and around Navsari district of South Gujarat, India

Pushpa M Makwana, IH Kalyani, Dhruv Desai, JM Patel, JB Solanki, PD Vihol, DR Patel, DM Muglikar and PS Sakhare

Abstract

The present study reports the incidence of bovine rotavirus infection in bovine calves of in and around areas of Navsari district. 157 faecal samples comprising (122 diarrheic and 35 non diarrheic) from 104 cattle and 53 buffalo calves of < 45 days' age were collected. All the samples were screened by LAT, ELISA and VP6 gene based RT-PCR and 17 (10.82%), 11 (7.0%) and 16 (10.19%) samples were found positive respectively. Diarrheic sample wise 13.11% incidence was recorded while all the non diarrheic samples were found negative. Species wise higher incidence was observed in cattle calves (10.58%) than buffalo calves (9.43%). Sex wise higher incidence was observed in female calves (10.61%) than male calves (6.81%). Age wise higher incidence was recorded in 0-15 days age group (11.58%) followed by 15-30 days age group (10.26%) and 30-45 days age group (4.35%).

Keywords: LAT, ELISA, BRV, TLP, RT PCR

1. Introduction

Diarrhea is one of the important causes of calf mortality, morbidity and economic losses in the dairy industry, especially in developing countries [1]. Calf diarrhea is attributed to both infectious and non-infectious factors. Multiple gastro-enteric pathogens (e.g., viruses, bacteria, and protozoa) are involved in the development of this disease. Amongst gastro enteric infectious agents, Rotavirus, Coronavirus, *Cryptosporidium* and *Escherichia coli* are collectively responsible for 75-95% of infection in neonatal calves worldwide and Rotavirus alone accounts for about 27- 36% [2, 3]. Rota-viral diarrhea are common in calves and affected young calves may die as a result of severe dehydration or secondary bacterial infections which in turn causes major economic losses [4].

Rotavirus belongs to the genus *Rotavirus* of the *Reoviridae* family possess distinct cartwheel structure while viewed in electron microscopy. The fully infectious RV particle is also termed as triple-layered particle (TLP) which is similar to wheels (latin word *Rota* means wheel) and this has led to the name of rotavirus for the genus [5]. The virus particles are about 80-100 nm in diameter, have icosahedral symmetry and has a segmented genome with 11 segments of double stranded (ds) RNA. The virus particle possesses 6 structural (VP1, VP2, VP3, VP4, VP6 and VP7) and 6 non-structural (NSP1, NSP2, NSP3, NP4, NSP5, NSP6) proteins. The virus consists of three concentric proteinaceous layers, the inner most viral core is formed of VP2 molecules, the VP1, VP3 and the dsRNA genome lie inner to the VP2 protein layer. The middle layer is composed of VP6 molecules and outer layer (outer capsid) is composed of VP4 and VP7 protein molecules.

Many enteric pathogens are responsible for neonatal calf diarrhea and therefore confirmatory diagnostic tests are necessary for specific agents. From Navsari district, few studies pertaining to bacterial neonatal calf diarrhea has been carried out, but study regarding virus association is significantly low, so present study was aimed to detect incidence of rotavirus infection in bovine neonatal calves this region.

2. Materials and Methods**2.1 Sample collection from calves**

A total of 157 faecal samples comprised of 122 diarrhoeic and 35 non diarrhoeic from 104 cattle and 53 buffalo calves were aseptically collected.

Calves ageing from birth to 45 days of age were selected and grouped as < 15 days, 15- 30 days and 30 -45 days from different regions in and around regions of Navsari district. Collected samples were immediately transported with ice pack to the department and then initially screened by LAT and then stored at -20 °C for processing by other tests.

2.2 Latex agglutination test (LAT)

LAT was performed to detect rotavirus antigen from faecal samples for initial screening of all the samples using HiRotavirus Latex Test Kit (HiMedia Laboratories, Mumbai, Cat. No LK08-50NO) as per the kit instructions.

2.3 Enzyme Linked Immunosorbent Assay (ELISA)

For antigenic diagnosis of bovine rotavirus, Bio X Bovine Rota virus ELISA kit (Bio X diagnostic, Belgium, Product ID - BIO K 343/2) was used and performed as per the kit instructions. The 96 well plate is coated with by specific antibodies for the rotavirus which allows specific capture of the rotavirus antigens present in the fecal samples. The net optical density of each sample was calculated at 450 nm by formula given by kit and value that obtained $\geq 22\%$ considered as positive sample.

$$\text{Value} = \frac{\text{Delta OD of Sample}}{\text{Delta OD of Positive}} \times 100$$

2.4 Reverse transcriptase Polymerase Chain Reaction (RT-PCR)

2.4.1 Extraction of Viral RNA

Pure quality viral RNA was extracted from faecal supernatants by QIAamp® Viral RNA Mini kit (Cat. No. 52904) as per the kit protocol. The extracted total RNA was quantified using nanodrop (NanoDrop 1000, Genaxy) at the ratio of 260/280 and stored at -20 °C until further use.

Table 3: Thermal cycler condition for Reverse Transcriptase PCR (RT-PCR)

cDNA Synthesis and pre-denaturation		Denaturation	Annealing temp according to primer	Extension	Final extension
1 cycle		35 Cycles			1 cycle
55 °C	94 °C	94 °C	47 °C	68 °C	68 °C
30 minutes	2 minutes	15 seconds	30 seconds	1 minute	5 minutes

Thermal cycler condition to amplify VP6 gene was set/ followed as per the kit instructions. The annealing temperature was set 10°C than the melting temperature of primer according to kit instructions. Hence, final thermal cyclic condition used for RT-PCR described in Table 3.

For confirmation of desired amplified gene, 8.0 µl of PCR product was mixed with 5X gel loading dye (2.0 µl) using 1 X TBE (Tris Borate EDTA) buffer along with 100 bp DNA molecular weight marker (6.0 µl) and analyzed on 1.5% agarose gel containing 0.5µg/ml ethidium bromide and Electrophoresis was run at 90 V for 1 hour. The amplified gene was examined under UV transilluminator and photographed in gel documentation unit (SynGene, Gene Genius Bio Imaging System, UK).

2.5 Statistical analysis

Species wise, sample wise, sex wise age wise, season wise statistical analysis were calculated by WASP - Web Agri Stat

2.4.2 Processing of RNA samples for one step RT PCR

The pure quality RNA (3.0 µl) was taken in the RNase free PCR tube and kept at 95°C for 5 minutes in thermal cycler and immediately snap chilled on ice. Then it was used for one step RT-PCR technique.

2.4.3 Method of one step RT- PCR

One step RT- PCR was performed by using one step RT PCR kit (SuperScript™ III one step RT-PCR kit with platinum™ Taq DNA polymerase) as per the kit manufacturer instructions. Reaction mixture was prepared for RT- PCR (Table 1) and immediately added into the PCR tube containing snap chilled RNA. Primers (Table 2) for the present study were send for custom synthesis from Eurofins Genomic Pvt. Ltd., Bangalore.

Table 1: Reaction mixture for RT PCR

Component	Volume
2X Reaction Mix	25 µl
Forward primer (VP6)	1 µl
Reverse primer (VP6)	1 µl
SuperScript™ III RT	2 µl
Autoclaved distilled water	18 µl
RNA sample	3.0 µl
Total reaction volume	50.0 µl

Table 2: Primers used for detection of VP6 gene in RT-PCR technique

Gene	Primers Sequence (5'–3')	Amplicon size
VP6- F	5'GAC GGV GCR ACT ACA TGG T3'	379bp
VP6- R	5'GTC CAA TTC ATN CCT GGT GG3'	

Package Developed by Ashok Kumar Jangam and Pranjali Ninad Wadekar at ICAR Research complex, Goa.

3. Results and Discussion

All the 157 samples were screened for the detection of bovine rotavirus. Out of these 17 (10.82%), 11(7.00%) and 16 (10.19%) samples were found positive by LAT, ELISA and RT- PCR respectively.

3.1 Detection of bovine rotavirus by Latex agglutination test (LAT)

Out of 157 samples tested total 17 (10.82%) samples were found positive by LAT that showed clear visible agglutination in faecal samples (Figure 1). Similar to this 10.86% positivity of bovine rotavirus was recorded [6] by other researchers. Compared to this higher positivity 12.80% [7], 23.93% [8], 63% [9], 65% [10], and 91% [11] as well as lower positivity 9.60% [12], 6.75% [13] observed by various workers.

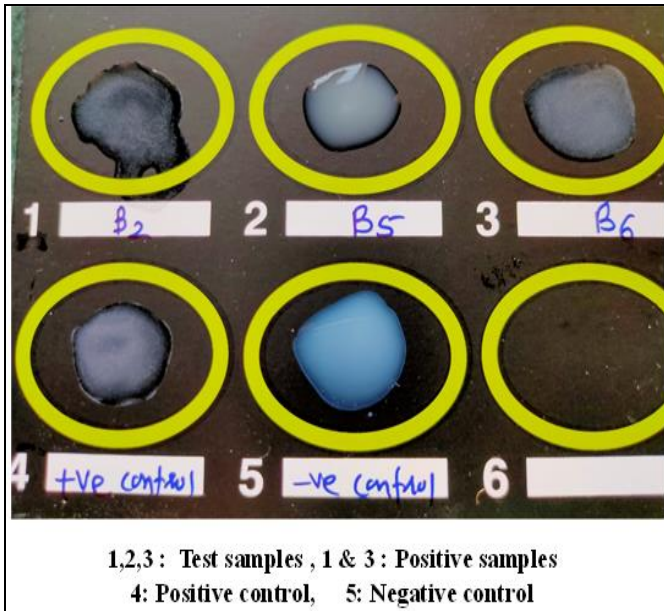


Fig 1: Detection of bovine rotavirus by Latex agglutination test

3.2 Detection of bovine rotavirus by ELISA

In the present study total 11 (7.00%) samples found positive by antigen capture ELISA (Figure 2). Value of each sample was calculated as per kit formula and value that obtained $\geq 22\%$ considered as positive sample. Similar to this findings various workers have used antigen detection based ELISA and reported lower positivity 7.0% and 4.61% by ELISA respectively [14, 15]. While others detected higher positivity 24.10%, 9.59%, 32.5%, 43.5%, 29.4%, 15.38% [16-20, 8] respectively, compared to present study.

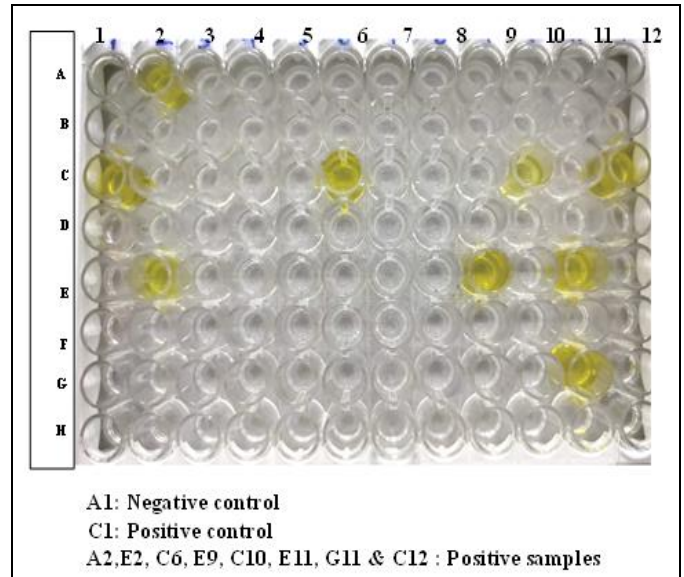


Fig 2: Detection of bovine rotavirus by ELISA

3.3 Detection of bovine rotavirus by Reverse Transcriptase Polymerase Chain Reaction (RT-PCR) by detecting VP6 gene

In the present investigation out of 157 samples 16 (10.19%) samples amplified for VP6 gene specific amplicon at 379 bp and revealed 10.19% positivity (Figure 3). Contrast to this lower infection rate 6.89% was recorded by others [21]. While others detected [22, 23, 4, 8] 24.10%, 36.22%, 22.28%, 22.22% higher infection rate by VP6 gene based RT-PCR.

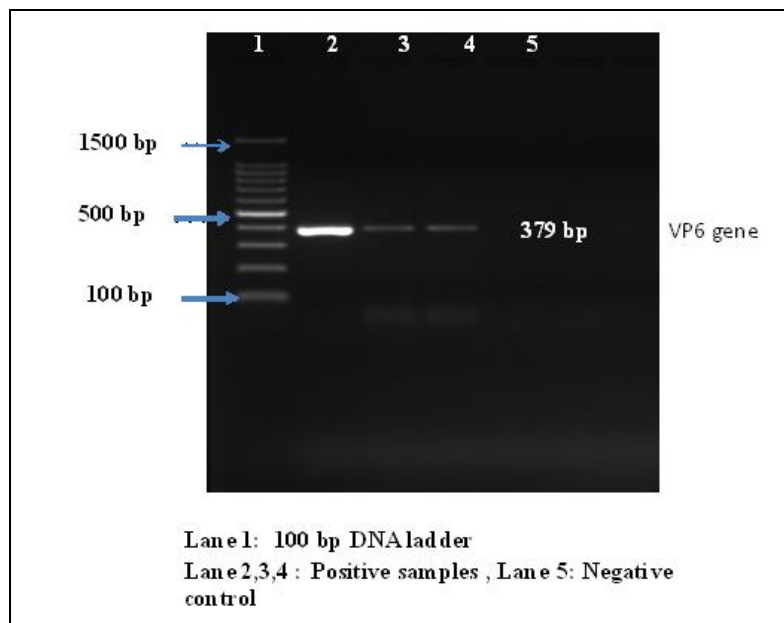


Fig 3: Detection of bovine rotavirus by RT - PCR

3.4 Overall detection of bovine rotavirus

As for confirmation of virus either virus isolation or nucleic acid based technique is more reliable, overall detection of rotavirus was considered on the basis of RT PCR. In the present study, overall positivity of rotavirus was detected 10.19%. Similar to this other workers [24] detected 10.0% positivity. Compared to this higher prevalence 63.50%

[25], 91% [11], 32.07% [26], 43.5% [19], 19.16% [27], 65% [10], 13.10% [28], 39.13% [8] as well as lower incidence 3%, 6.75%, 8.92%, 6%, 6.89%, 5.11% detected by various other workers [29, 13, 30, 31, 21, 1] respectively. For the present study, species wise, sample wise, sex wise, age wise and season wise positivity of rotavirus was considered on the basis of positivity of RT PCR (Table 4).

Table 4: Detection of bovine rotavirus in bovine calves

Parameter		Total number of samples screened	Number of samples positive	%	χ^2 value
Species	Cattle	104	11	10.58	0.050 ($P>0.05$)
	Buffalo	53	05	9.43	
Type of sample	Diarrhoeic	122	16	13.11	5.111* ($P\leq 0.05$)
	Non diarrhoeic	35	00	00	
Sex	Male	44	04	9.09	0.081 ($P>0.05$)
	Female	113	12	10.61	
Age	<15 days	95	11	11.58	1.058 ($P>0.05$)
	15-30 days	39	04	10.26	
	30-45 days	23	01	4.35	
Season	Summer	12	01	8.33	0.061 ($P>0.05$)
	Rainy	34	03	8.82	
	Winter	111	12	10.81	

3.4.1 Species wise incidence of bovine rotavirus

In the present study, cattle calves showed 10.58% higher infection as compared to buffalo calves 9.43%. Similar to this, others have [32] detected more prevalence in cow calves (41.6% by RNA PAGE and 33.3% by ELISA) than buffalo calves (20% by RNA PAGE and 24% by ELISA). In contrast to this lower prevalence in cattle calves than buffalo calves (22.73% and 77.78%) and (13.33% and 22.01%) was reported [26, 33] respectively.

3.4.2 Sample wise incidence of bovine rotavirus

In the present study, 16 (13.11%) diarrhoeic samples found positive while from 35 non diarrhoeic samples all the samples were found negative. This result shows significant difference between diarrhoeic and non diarrhoeic samples for detection of rotavirus. Similar to this, others [23, 34] have also found positive samples from diarrhoeic calves while all the non-diarrhoeic samples were found negative.

3.4.2 Sex wise incidence of bovine rotavirus

Sex wise, female calves showed higher incidence 10.61% than male calves 9.09%. This finding showed agreement with other researchers [35, 36] who recorded higher infection rate in female than male. In contrast to this others [34, 8] found higher positivity in male than female calves.

3.4.3 Age wise incidence of bovine rotavirus

In the present study, highest infection rate was observed in age group pertaining to 0-15 days 11.58% followed by 15-30 days 10.26% and then in 30-45 days 4.35%. These findings are in accordance with various workers [8, 19, 35] who also observed higher infection rate in early days of age group i.e. 1-15 days. The age wise higher infection in present study in earlier age (0-15 days) might be due to immune system of very young calves is not fully mature to fight against infection. Other workers [37-39] also revealed that diarrhea in calves from 5 to 10 days of age is commonly due to rotavirus infection. Similarly other workers [34, 40-42] reported that the susceptibility of bovine calves to rotavirus infection decreases with age, probably due to loss of receptors on enterocytes. In the present study, lower infection rate in 30-45 days age group compared to 0-15 days age group recorded that may be due to with progression of age calves acquire increased natural resistance against enteropathogens [43, 44, 35, 19].

3.4.4 Season wise incidence of bovine rotavirus

In the present investigation, highest positivity (10.81%) found during winter month followed by rainy season (8.82%) and summer (8.33%). In accordance with this, others [1, 31] also

detected higher prevalence in winter season compared to summer season. Various other workers also stated that diarrhea in calves due to rotavirus mostly occurs during winter periods and it has been observed that the morbidity also increases during adverse weather conditions [45-47]. This may be due to overcrowding of population and cold temperature leads to stress to younger calves.

4. Conclusion

Present study reveals importance of Rotavirus as a one of etiological agent of diarrhea in bovine calves and has been associated with significant economic losses to dairy industry. For detection of bovine rotavirus antigen diarrhoeic samples should be screened by LAT and ELISA for initial screening of samples while for confirmation of rotavirus nucleic acid detection based technique should be used. As there is variation was seen in positivity by different tests used combination of two or more tests should be used.

5. Acknowledgement

The authors are thankful to the Department of Veterinary Microbiology, Vanbandhu College of Veterinary science and Animal Husbandry, Navsari Agricultural University, Navsari for giving opportunity to do research study.

6. References

- Barua SR, Rakib TM, Rahman MM, Selleck S, Masuduzzaman M, Siddiki AZ *et al.* Disease burden and associated factors of rotavirus infection in calves in south-eastern part of Bangladesh. *Asian Journal of Medical and Biological Research.* 2019; 5(2):107-116.
- Uhde FL, Kaufmann T, Sager H, Albin S, Zanoni R, Schelling E *et al.* Prevalence of four enteropathogens in the faeces of young diarrhoeic dairy calves in Switzerland. *Veterinary Record.* 2008; 163:362-366.
- Dhama K, Chauhan R, Mahendran M, Malik S. Rotavirus diarrhea in bovines and other domestic animals. *Veterinary Research Communications.* 2009; 33(1):1-23.
- Das S, Medhi M, Khaound M, Doley P, Islam M, Borah DP. Detection of group a rotavirus infection in diarrhoeic calves by electropherotyping and reverse transcriptase polymerase chain reaction. *Journal of Entomology and Zoology Studies.* 2018; 6(3):1071-1075.
- Flewett TH, Bryden A, Davies H, Woode GN, Bridger JC, Derrick JM. Relation between viruses from acute gastroenteritis of children and newborn calves. *Lancet.* 1974; 2:61-63.
- Kadam AS, Tembhumne PA, Bonde SW, Chaudhary SP, Kurkure NV, Ingle VC. Molecular detection of group A

- rotaviruses from the Bhandara and Chandrapur district of Maharashtra state, India. *International Journal of Current Microbiology and Applied Sciences*. 2019; 8(9):2555-2564.
7. Hassan Mir Nadeem G. P typing and sequence analysis of group A rotavirus in calves and lambs in Kashmir. PhD thesis submitted to Sher-e-Kashmir University of Agricultural Sciences and Technology of Kashmir Division of Veterinary Microbiology & Immunology Shuhama Campus, Srinagar, 2014.
 8. Patel J, Mathakiya R, Golaviya A. Detection of bovine rotavirus from diarrheic bovine calves in Gujarat region, India. *International Journal of Current Microbiology and Applied Science*. 2019; 8(9):1282-1293.
 9. Yousif AY, Anderson J, Chard-Bergstrom C, Bustamante A, Muenzenberger M, Austin K *et al*. Evaluation of a Latex Agglutination Kit (Virogen Rotatest) for detection of bovine Rotavirus in fecal samples. *Clinical and Diagnostic Laboratory Immunology*. 2001; 8:496-498.
 10. Muhammid HA, AL- Shemmari IGM, Alkabi MAS. Estimating the sensitivity and specificity of Reverse transcriptase polymerase chain reaction (RT – PCR) with rotavirus latex agglutination test (Virogen Rotalex) in calves suffering from diarrhoea in Kabala province. *Scientific Journal of Medical Research*. 2017; 1(4):110-114.
 11. Reidy N, Lennon G, Fanning S, Power E, O'Shea H. Molecular characterisation and analysis of bovine rotavirus strains circulating in Ireland 2002–2004. *Veterinary microbiology*. 2006; 117(2):242-247.
 12. Okada N, Matsumoto Y. Bovine rotavirus G and P types and sequence analysis of the VP7 gene of two G8 bovine rotaviruses from Japan. *Veterinary Microbiology*. 2002; 84:297-305.
 13. Dulgheroff ACB, Pereira WAB, Sarmiento RR, Silva GAV, Naveca FG, Domingues ALS. Analysis of bovine rotavirus strains circulating in diarrheic dairy calves in Uberaba, Minas Gerais, Brazil, during 2008–2009. *The journal Arquivo Brasileiro de Medicina Veterinária e Zootecnia*. 2016; 68(4):1090-1094.
 14. Selim SA, Aziz KMS, Sarker AJ, Rahman H. Rota virus infection in calves in Bangladesh. *Veterinary Research Communications*. 1991; 15(4):327-333.
 15. Manuja BK, Prasad M, Manuja A, Gulati BR, Prasad G. A novel genomic constellation (G10P[3]) of Group A Rotavirus detected from buffalo calves in northern India. *Virus Research*. 2008; 138(1, 2):36-42.
 16. Tamilmani S, Ram BR, Kuldeep D, Pradeep MS, Deepak K, Prakash B. Determination of G and P type diversity of group A rotaviruses and detection of a new genotype from diarrhoeic calves in northern and southern states of India. *Veterinary Practitioner*. 2012; 13(1):1-15.
 17. Gill GS, Kaur S, Dwivedi P, Gill JS. Comparative prevalence and molecular characterization of group A rotavirus in cow calves of Punjab, India. *Journal of Animal Research*. 2017; 7:927-933.
 18. Ali YH, Khalafalla AI, Intisar KS, Halima MO, Salwa AE, Taha KM *et al*. Rota virus infection in human and domestic animals in Sudan. *Journal of Science and Technology*. 2011; 12(4):1605.
 19. Khamees AKS. Detection of Rota and Corona viral antigens in diarrheic newly born calves in Menofiya governorate. *Benha Veterinary Medical Journal*. 2015; 29:9-16.
 20. Soltan MA, Tsai YL, Lee PA, Tsai CF, Chang HG, Wang HT *et al*. Comparison of electron microscopy, ELISA, real time RT-PCR and insulated isothermal RT-PCR for the detection of Rotavirus group A (RVA) in feces of different animal species. *Journal of Virological Methods*. 2016; 235:99-104.
 21. Tumlam UM, Ingle VC, Tembhurne PA, Kurkure NV, Chaudhari SP, Chitambar SD *et al*. Detection of VP6 gene of rotavirus in feces of diarrhoeic calves, kids, lambs, piglets, pups and human infants by Reverse Transcriptase–Polymerase Chain Reaction. *The Indian Journal of Veterinary Sciences & Biotechnology*. 2018; 13(4):12-16.
 22. Suresh T, Rai RB, Dhama K, Sawant PM, Kumar D, Bhatt P. Detection of group A bovine Rotavirus in diarrhoeic calves by reverse transcriptase polymerase chain reaction (RT-PCR) and electrophoresis. *Veterinary practitioner*. 2011; 12(2):133-137.
 23. Ahmed SP, Hazarika RA, Bora DP, Tamuly S. Detection and genotypic characterization of Rotavirus from bovine calves of Assam, A North Eastern state of India. *The Journal of Animal & Plant Sciences*. 2017; 27(2):439-445.
 24. Kassem IK, Magouz AF, Desouky AY, Hagag MF. Isolation and Identification of Rotavirus Infection in diarrheic calves at El Gharbia Governorate. *Global Veterinaria*. 2017; 18 (3):178-182.
 25. Al-Yousif Y, Anderson J, Chard- Bergstrom C, Bustamante A, Muenzenberger M, Austin K *et al*. Evaluation of a latex agglutination kit (Virogen Rotatest) for detection of bovine rotavirus in faecal samples. *Clinical and Diagnostic Laboratory Immunology*. 2001; 8(3):496-498.
 26. Singh TC, Jhala MK. Comparing relative sensitivity and specificity of LA and RNA-PAGE in detecting bovine rotavirus. *Buffalo Bulletin*. 2011; 30(1):36-40.
 27. Sravani GVD, Kaur G, Chandra M, Dwivedi PN. Prevalence of group A bovine rotavirus in neonatal calves in Punjab, India. *Journal of Microbiology*. 2015; 2:9-14.
 28. Monney JD, Adjogoua VE, Karamoko Y, Akran V. Prevalence of rotavirus infection in diarrheic newborn calves in Abidjan region, Ivory Coast. *GSC Biological and Pharmaceutical Sciences*. 2018; 5(2):82-87.
 29. Gandhi SS. Genomic profile electrom microscopy and isolation of bovine rotavirus from diarrhoeic calves. M.V. Sc Thesis, Submitted to Gujarat Agricultural University, S.K. Nagar, 1992.
 30. Yilmaz V. Investigation of rotavirus infection in calves with diarrhea in Northeast Turkey. *American Journal of Animal and Veterinary Sciences*. 2016; 4:1-4.
 31. Mukhtar N, Yaqub T, Munir M, Nazir J, Aslam A, Masood A *et al*. Prevalence of group A bovine rotavirus in neonatal calves in Punjab, Pakistan. *The Journal of Animal & Plant Sciences*. 2017; 27(2):379-383.
 32. Jindal SR, Maiti NK, Oberoi MS. Genomic diversity and prevalence of rotavirus in cow and buffalo calves in northern India. *Revue scientifique et technique*. 2000; 19(3):871-876.
 33. Malik YS, Kumar N, Sharma K, Sharma R, Kumar HB, Anupamlal K *et al*. Epidemiology and genetic diversity of rotavirus 144 strains associated with acute gastroenteritis in bovine, porcine, poultry and human population of Madhya Pradesh, Central India, 2004–

2008. *Advances in Animal and Veterinary Sciences*. 2013; 1(4):111-115.
34. Dash SK, Tewari A, Kumar K, Goel A, Bhatia AK. Detection of rotavirus from diarrhoeic cow calves in Mathura, India. *Veterinary World*. 2011; 4:554-556.
35. Ammar SSM, Mokhtaria K, Tahar BB, Amar AA, Redha BA, Yuva B *et al*. Prevalence of rotavirus (GARV) and coronavirus (BCoV) associated with neonatal diarrhea in calves in western Algeria. *Asian Pacific journal of tropical biomedicine*. 2014; 4:S318-S322.
36. Hassan HA, Kshash QS, Mansur KA. Detection of bovine rotavirus in diarrheic calves by using rapid test in some Mid-Euphrates provinces. *AL-Qadisiya Journal of Veterinary Medicine Sciences*. 2014; 2:20-26.
37. Tzipori S. The relative importance of enteric pathogens affecting neonates of domestic animals. *Advances in Veterinary Science and Comparative Medicine*. 1985; 29:103-206.
38. Radostits OM. A veterinary clinician's perspective of diarrhea in neonatal food producing animals. *Proc. Int. Semi. On Diarr. Disease in S.E. Asia, Australia and W. Pacific regions*. *Journal of Clinical Microbiology*. 1985, 9-19.
39. Tewari A, Dash S, Jain B, Bhatia A. Electrophoretic detection of bovine Rotavirus by RNA-PAGE in Mathura. *Indian Journal of Animal Research*. 2012; 2:67-71.
40. Minakshi P, Prasad G, Malik Y, Pandey R. G and P genotyping of bovine group A rotaviruses in faecal samples of diarrhoeic calves by DIG-labelled probes. *Indian Journal of Biotechnology*. 2005; 4:93-99.
41. Udaykar A, Sharda R, Malik YS, Sharma V, Shrivastava N. Occurrence of group A rotavirus in diarrhoeic buffalo and cow calves, Madhya Pradesh. *Advances in Animal and Veterinary Sciences*. 2013; 4:51-53.
42. Kumari S, Anjay M, Kaushik P, Kumari S, Kumar P. Detection of rotavirus in diarrheic bovine calves by RNA-PAGE. *Journal of Agri Search*. 2019; 6:131-134.
43. Gumusova SO, Yazici Z, Albayrak H, Meral Y. Rotavirus and Coronavirus prevalence in healthy calves and calves with diarrhea. *Medycyna Weterinaria*. 2007; 63:62-64.
44. Suresh T, Rai RB, Wani MY, Damodaran T, Dhama K. Detection of bovine rotavirus in neonatal calf diarrhoea by ELISA, FAT and transmission electron microscopy. *International Journal of Current Research*. 2013; 5(7):1935-1939.
45. Woode GH, Bridger JC. Viral enteritis of calves. *The Veterinary Record*. 1975; 96(4):85-88.
46. Woode GH, Crouch CF. Naturally occurring and experimentally induced rotavirus infection of domestic and laboratory animals. *Journal of the American Veterinary Medical Association*. 1978; 173:522-526.
47. Chauhan RS, Dhama K, Mahendran M. Pathobiology of Rotaviral diarrhea in calves and its diagnosis and control: A Review. *Journal of Immunology and Immunopathology*. 2008; 10(1):1-13.