

E-ISSN: 2320-7078 P-ISSN: 2349-6800 www.entomoljournal.com JEZS 2020; SP-8(3): 60-66

© 2020 JEZS Received: 01-03-2020 Accepted: 03-04-2020

Sudhanshu Pratap Singh

Department of Veterinary Gynaecology and Obstetrics, Bihar Veterinary College, Bihar Animal Sciences University, Patna, Bihar, India

Ankesh Kumar

Department of Veterinary Clinical Complex, Bihar Veterinary College, Bihar Animal Sciences University, Patna, Bihar, India

Prakrutik Prafulchandra Bhavsar College of Veterinary Science and Animal Husbandry, Anand Agricultural University, Anand,

Gujarat, India Mukesh Sahu

Department of Veterinary Gynaecology and Obstetrics, College of Veterinary and Animal Sciences G. B. Pant University of Agriculture and Technology, Pant Nagar, Uttarakhand, India

Praveen Kumar

Department of Veterinary Gynaecology and Obstetrics, Bihar Veterinary College, Bihar Animal Sciences University, Patna, Bihar, India

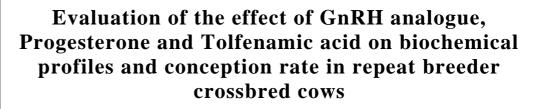
Sushil Kumar

Department of Veterinary Clinical Complex, Bihar Veterinary College, Bihar Animal Sciences University, Patna, Bihar, India

Corresponding Author: Sudhanshu Pratap Singh Department of Veterinary Gynaecology and Obstetrics, Bihar Veterinary College, Bihar Animal Sciences University, Patna, Bihar, India

Journal of Entomology and Zoology Studies

Available online at www.entomoljournal.com



Journal of Entomology and

Zoology Studies

Z

Sudhanshu Pratap Singh, Ankesh Kumar, Prakrutik Prafulchandra Bhavsar, Mukesh Sahu, Praveen Kumar and Sushil Kumar

Abstract

The present study was focused to study the effects of parenteral administrations of GnRH analogue-Buserelin acetate, Progesterone and Tolfenamic acid on biochemical parameters viz., serum glucose, serum cholesterol and serum total protein in repeat breeding animals. A total of 32 repeat-breeder crossbred cows were selected and divided in 4 equal groups (n=8). Group 1 was kept as positive control *i.e.* without any treatments, Group 2 animals were treated with Buserelin acetate, Group 3 animals were treated with Buserelin acetate + Progesterone and Group 4 animals were treated with inj. Buserelin acetate + Progesterone + Tolfenamic acid . Biochemical analysis was done by commercially available diagnostic kits. The mean blood glucose and serum protein concentration differed non- significantly without any changes in the normal range. The mean value of total serum cholesterol concentration on day 0 varies significantly (p<0.05) in group 2 & 4 compared to other groups. Moreover, on other days, cholesterol mean value was significantly (p<0.05) higher in group 4 as compared to other groups. Highest conception rate was observed in group 4 in which combinations of drugs were used.

Keywords: GnRH analogue, progesterone, Tolfenamic acid, conception rate, cholesterol

Introduction

Repeat breeding (RB) is a considerable problem in cattle breeding which leads to large economic losses due to increasing in the number of inseminations, increasing in the length of calving interval and moreover increasing in culling rates. A repeat breeder is generally defined as any cow that has not conceived after three or more services associated with true estrus ^[1].

The bovine conceptus produces interferon-tau (IFN τ) which prevents luteolysis over and above functional corpus luteum (CL) produces progesterone which is very much essential for supporting pregnancy ^[2]. There are two crucial periods of bovine pregnancy. The first period is first week after breeding and second period is from day 8 to 28, at which maternal recognition of pregnancy (MRP) takes place. Approximately 32% of total embryonic loss reported in this second period ^[3]. This leads the problem of repeat breeding.

Several factors like nutritional stress ^[4], heat stress ^[5], transportation stress ^[6] or other stress promotes secretion of prostaglandin F2 α from the uterine endometrium. This can cause lysis of the functional corpus luteum and leads to early embryonic death ^[7]. The two cyclooxygenase (COX) enzymes convert arachidonic acid into prostaglandin-H2 which is further converted into PGF2 α through the enzyme prostaglandin-F-synthase. The NSAIDs exhibits antiinflammatory activity mainly by the inhibition of the cyclo-oxygenase (COX) enzyme, results in inhibition of prostaglandin synthesis ^[8]. Besides tolfenamic acid, several studies also evaluated the effects of NSAIDs of Flunixine meglumine ^[9, 10] and meloxicam ^[11, 12]. Whereas administration of tolfenamic (NSAIDs) acid has significantly improved embryo transfer rate and pup delivery in mice ^[13].

Administration of GnRH or GnRH analogue before artificial insemination induces preovulatory LH (luteinizing hormone) surge which controls ovulation, while post inseminations with supplementation of exogenous P4 support early embryonic development ^[11]. At days 3 to 5 post-ovulation, the embryos usually enter the uterus, undergoing genomic activation and increases in P4 concentration; therefore, this may be a physiologically important time in the cattle, so administration of a low dose of P4 on days 4, 5 and 6 of the estrous cycle

increase the conception rate among repeat breeder cattle ^[14]. This extend the life span of the bovine corpus luteum (CL) so this is one of the strategies aimed at reducing embryo loss by inhibiting the PGF2 α in the endometrium during the critical period ^[15, 16], Inhibition of PGF2 α enhances the CL lifespan and avoiding detrimental and toxic effects of PGF2 α on the embryo ^[15].

Glucose is an important biochemical factor which regulate and modulate the metabolic functioning of the cell at different levels, it also acts at hypothalamus-pituitary gonadal axis and affect the reproductive function in animals, moreover, it is essential to regulate the quality of oocytes ^[17, 18]. It was noted that glucose within the oocyte regulates meiotic maturation and the glucose transporter (GLUT) content increased in rat ovary following gonadotropin use ^[19, 20]. Thus, Glucose called as central nutrient in metabolism and must for reproductive tract function. Cholesterol is a precursor of reproductive hormone and also a biochemical factor which represents the condition of lipid metabolism and have a positive relationship with the health of the animals ^[21]. Demand for cholesterol for the biosynthesis of progesterone, estrogen by the avascular granulosa cells increases during the LH surge ^[22]. Low levels of cholesterol with reduced levels of steroidogenesis [23]. Although, the high incidences of repeat breeding and anestrus are associated with the deficiencies of cholesterol. Serum concentration of protein reflects the nature of the uterine environment which is one of the most important factors for embryo development ^[24]. The dynamic of uterine lumen exhibits a marked difference at the stages of the estrous cycle as a consequence of ovarian steroidal regulation of endometrial secretion. Intakes of high protein diet alter the pH and the concentrations of other ions in uterine secretions, but only during the luteal phase of estrous cycle [25]. A study report showed that if the higher amount of rumen degradable protein fed to heifers reduces fertility by altering the uterine pH^[24].

Materials and Methods

Preparation of animals before commencement of treatment

The study was conducted on 32 apparently healthy, 3-8 years old repeat breeding crossbred cattle. All animals were dewormed with Fenbendazole @ 7.5 mg per kg body weight 60 days prior to the commencement of experiment and they were supplemented with 40 gm mineral mixture daily prior to experiment.

Grouping of animals and treatment

Selected animals were randomly divided into four groups (each group containing 8 animals). In group 1, animals were inseminated on spontaneous estrous without any treatment, in group 2, animals were treated with injection Buserelin acetate @ 20 μ g IM at the time of artificial insemination (AI), in group 3, animal were treated with injection Buserelin acetate @ 20 μ g IM at the time of AI followed by Injection P4 @100mg IM on days 4,5,6 after AI and in group 4, animals were treated with injection Buserelin acetate @ 20 μ g IM at the time of AI followed by Injection P4 @100mg IM on days 4,5,6 after AI and in group 4, animals were treated with injection Buserelin acetate @ 20 μ g IM at the time of AI, Injection P4 @100mg IM on days 4,5,6 and Inj. Tolfenamic acid @4 mg/kg body weight IM On days 16,17,18 after AI. All the animals were inseminated on spontaneous heat.

Blood collection and biochemical assay

Blood samples were collected from all the cows at day 0, 7, 16, 17, 18 of the oestrous cycle. Estrous day assumed as day '0'. After restraining of the cow 10 ml of blood was collected aseptically from jugular vein. 2ml blood sample transferred into fluoride vial for serum glucose estimation. 8 ml blood sample transferred into clot activator vial for separation of serum for estimation of total cholesterol, total protein and progesterone. Blood sampling done in clot activator vial was allowed to clot in slanting position for about one hour at room temperature. Then it transferred into a centrifuge tube with the help of micro pipette and centrifuge for 15 minutes at 3000 rpm. Blood sample of fluoride vial was also centrifuge for plasma separation. The serum and plasma samples transfer in eppendorf tube with the help of micropipette and kept at (-20) degree centigrade for further use.

Estimation of glucose, cholesterol and protein was carried out according to manufacturer's protocol. Glucose was estimated by Glucose Test Kit by GOD-POD method. Cholesterol was estimated by Cholesterol Test Kit by CHOD-PAP method and the plasma total protein in blood serum was determined by using diagnostic kit manufactured by Coral Clinical Systems. Statistical analysis

Repeated Measures ANOVA was used to analysis the obtained result. The multiple comparisons between group, day and interaction for different parameters were done by using Tukey test at 5% level of significance. The analysis was done using JMP 9.0 software.

Results and Discussion

Total Blood Glucose

The mean blood glucose concentration was observed with non-significant difference among all groups (group 1, group 2, group 3 and group 4) on day '0', 7, 16, 17 and 18 (Table-1).

The mean blood concentration of glucose reported in the present study was nearly similar to that of ^[26–28]. Administration of GnRH and leads to significant differences in mean serum concentration of glucose, but the observations were found similar in the present study ^[29]. Flunixin meglumine at the dose rate of 1.1 mg/kg body weight in dogs for 5 days intravenously causes non-significant changes in the blood glucose concentration ^[30], though, there is an increased in blood glucose level after using dexamethasone and prednisolone in normal cows ^[31]. Supplementation of flaxseed to cow was observed to have higher value of glucose compared to present study and similar higher value was also observed in a study by ^[32, 33]. Some studies also showed the lower value with respect to present study ^[34, 35].

Circulating glucose concentration were lower in severe negative energy balance (SNEB) compared to mild negative energy balance (MNEB) in mares ^[36].

Glucose is also essential for gonads to maintain the quality of oocytes, development of embryo ^[17, 18]. There were no significant changes seen in blood serum glucose concentration and this might be due to no effects of GnRH on metabolic profile of animals. A study also showed the decreased in blood glucose concentration in cows with advancement of pregnancy in winters ^[37].

 Table 1: Mean (± SE) blood glucose concentration (mg/dl) in different groups on 0, 7th, 16th, 17th and 18th day of estrous cycle in repeat breeder cows.

Group	Day 0	Day 7	Day 16	Day 17	Day 18
1	56.32 ± 0.84	56.13 ± 0.84	55.71 ± 0.94	56.02 ± 1.32	56.08 ± 1.34
2	54.01 ± 1.54	55.29 ±1.61	54.59 ± 1.65	55.11 ± 1.99	56.12 ± 2.01
3	52.53 ± 1.77	53.65 ± 2.28	53.37 ± 2.04	55.11 ± 2.20	54.94 ± 2.58
4	55.29 ± 1.21	56.05 ± 1.09	57.45 ± 1.24	57.74 ± 0.98	57.59 ± 1.31

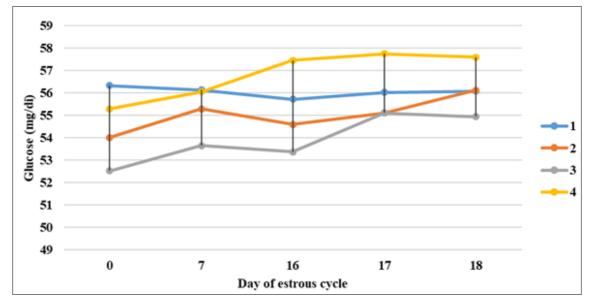


Fig 1: Mean (± SE) blood glucose concentration (mg/dl) in different groups on 0, 7th, 16th, 17th and 18th day of estrous cycle in repeat breeder cows.

Total Serum Cholesterol

The significant difference was observed in mean total serum cholesterol concentration between groups 4 and 3 as compared to group 1 and 2 on day '0'. There was observed significantly (p<0.05) higher level of serum cholesterol on days 0,7,16,17& 18 in group 4 compared to all three groups i.e. group 1, 2& 3 (Table-2).

The mean serum cholesterol concentration of treated animal found in the present experiment were in agreement with reports of ^[26, 32, 34, 37] but the mean serum cholesterol concentration of non-treated animal group was lower. However, lower concentration has been reported by ^[38] in buffaloes during different stages of estrous cycle.

The mean serum cholesterol concentration in the present experiment showed the higher values as compared to results reported by $^{[39-41]}$, in fertile (112.10±3.57) and repeat breeder cows (126.10±9.74) $^{[33]}$.

Effect of GnRH, hCG and progesterone impregnated device on blood biochemical profile in repeat breeder crossbred cows had the lower value of cholesterol compared to the present study^[42].

All values observed in the present study were within normal range of 65-220 mg/dl ^[43]. Demand for cholesterol increases during the biosynthesis of progesterone, estrogen by the avascular granulosa cells under the influence of LH surge ^[44]. It is suggested that the low levels of cholesterol is related with reduced levels of steroidogenesis ^[23].

Increase in cholesterol level in serum causes greater availability of cholesterol in ovarian follicular fluid, and luteal tissue which has been associated with increase in E2 synthesis by the follicles result into better LH surge which finally helps in improving the embryo quality and similarly increase in P4 synthesis within CL ^[45].

 Table 2: Mean (± SE) serum cholesterol concentration (mg/dl) in different groups on 0, 7th, 16th, 17th and 18th day of estrous cycle in repeat breeder cows

Group	Day 0	Day 7	Day 16	Day 17	Day18
1	148.22±1.99 ^{JK}	154.06±1.83 ^{HI}	159.11±2.00 ^{EFG}	160.71±2.85 ^{DEF}	160.69±2.85 ^{DEF}
2	150.69±1.68 ^{IJ}	155.07±2.01 ^{GHI}	158.95±2.20 ^{EFG}	161.53±2.22 ^{DE}	162.86±2.60 ^{CDE}
3	145.27±4.11 ^K	152.73±2.91HI ^J	156.68±3.30 ^{FGH}	162.48±2.68 ^{CDE}	164.85±3.87 ^{BCD}
4	153.95±2.78 ^{HI}	162.21±3.11 ^{CDE}	166.55±3.22 ^{ABC}	168.55±3.28 ^{AB}	170.35±3.32 ^A

The values bearing the different superscripts across the row and column differ significantly from each other, P<0.05

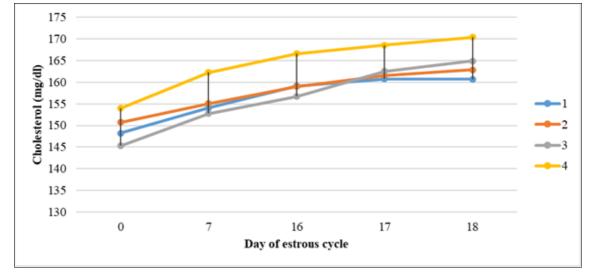


Fig 2: Mean serum cholesterol concentration (mg/dl) in different groups on 0, 7th, 16th, 17th and 18th day of estrous cycle in repeat breeder cows

3.3 Total Serum Protein

There have been non-significant differences observed among the entire treatment group on day 0, 7, 16, 17, &18 in the present study (Table-3).

The mean serum concentration of total blood protein was observed in the present study is 6-7 g/dl, which is comparable with the reports of other scientists ^[46]. The mean total serum protein concentration observed in the present study is at par with the reports of different studies ^[42, 47, 48]. On the contrary the values of mean total serum protein were lower than ^[35]. Similar lower values of mean total protein concentration during different stages of estrous cycle in buffaloes were also reported by ^[38]. The present finding is in agreement with report of in which they showed that the value of serum total protein concentration does not differ significantly following administration of Flunixin Meglumine by repeatedly at 12-hour interval for 12 days in crossbred goats ^[49]. Flunixin Meglumine at the dose rate of 1.1 mg/kg body weight in foal

and observed decreased in protein level with non-significant difference ^[50]. Administration of Flunixin Meglumine at the dose rate of 1.1 mg/kg body weight in dogs for 5 days by intravenously and the observed serum protein concentrations were not consistent with the present findings ^[30]. The total protein level in pregnant and non-pregnant with non-significant difference in all the treatment groups on different days of estrus cycle i.e. 0, 5, 10, 15 and day 20 ^[29].

Use of Flunixin Meglumine there was no any direct effect on protein level ^[49]. The values of protein level were decreased if it is given repeatedly due to Gastro-enteropathy which causes loss of protein.

Our findings are also comparable to the findings, who also observed the non-significant difference in serum protein levels between stages of the estrous cycles in cows^[34].

Whereas, the values of mean serum Protein concentration in fertile (7.09 ± 0.27) and repeat breeder cow (7.92 ± 0.31) were higher as compared to present study ^[33].

 Table 3: Mean (± SE) serum protein concentration (mg/dl) in different groups on 0, 7th, 16th, 17th and 18th day of estrous cycle in repeat breeder cows.

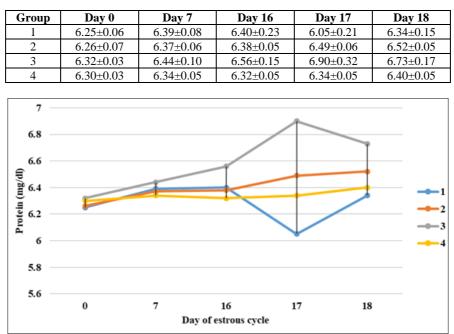


Fig 3: Mean serum protein concentration (mg/dl) in different groups on 0, 7th, 16th, 17th and 18th day of estrous cycle in repeat breeder cows.

Conception Rate

The conception rate was observed increased in the entire treated group with respect to non-treated group and the maximum conception was found in the group-4, where combinations of all treatment were given ^[51]. The combinations in group-4 were Buserelin acetate, Exogenous P_4 and Tolfenamic acid, these therapies help to maintain the P_4 level and significantly increases the conception rate by six times than control group, moreover, the level of cholesterol found high in pregnant animal with respect to non-pregnant shows that cholesterol level supports the pregnancy. The conception rate in groups 1, 2, 3 & 4, were 12.5%, 37.5%, 50%, 75% respectively.

Conclusion

In partial report of this study, conception rate had been reported increased in all the treated group with respect to nontreated group and the maximum conception was observed in the groups where combinations of all treatment were given as well as the level of cholesterol was also observed significantly higher in group 4 as compared to other groups. Moreover, the serum cholesterol level were also observed higher in pregnant animals with respect to non-pregnant animals. There are non-significant changes observed in glucose and protein level, but significantly increased in the level of serum cholesterol which acts as a precursor of steroid hormone following treatment might have supported the increased in conception rate.

Acknowledgements

We thank DRI-cum-Dean PGS, BASU, Patna for the financial support received through Bihar Animal Sciences University, for timely completion of my research work and Dr Gyanendra Singh, Principal Scientist, Division of Physiology & Climatology, IVRI, Izatnagar for his kind cooperation for analysis of Hormone is also gratefully acknowledged.

References

- 1. Purohit GN. Recent developments in the diagnosis and therapy of repeat breeding cows and buffaloes. CAB Reviews: Perspectives in Agriculture, Veterinary Science, Nutrition and Natural Resources, 2008, 3. https://doi.org/10.1079/PAVSNNR20083062.
- Spencer TE, Bazer FW. Biology of progesterone action during pregnancy recognition and maintenance of pregnancy. Frontiers in Bioscience: A Journal and Virtual Library. 2002; 7:1879-98. https://doi.org/10.2741/spencer.
- Wiltbank MC, Baez GM, Garcia-Guerra A, Toledo MZ, Monteiro PLJ, Melo LF *et al.* Pivotal periods for pregnancy loss during the first trimester of gestation in lactating dairy cows. Theriogenology. 2016; 86:239-53. https://doi.org/10.1016/j.theriogenology.2016.04.037.
- 4. Bender RW, Hackbart KS, Dresch AR, Carvalho PD, Vieira LM, Crump PM, *et al.* Effects of acute feed restriction combined with targeted use of increasing luteinizing hormone content of follicle-stimulating hormone preparations on ovarian superstimulation, fertilization, and embryo quality in lactating dairy cows. Journal of Dairy Science. 2014; 97:764-78. https://doi.org/10.3168/jds.2013-6926.
- 5. Sakatani M. Effects of heat stress on bovine preimplantation embryos produced *in vitro*. Journal of Reproduction and Development. 2017; 63:347-52.

https://doi.org/10.1262/jrd.2017-045.

- Merrill ML, Ansotegui RP, Burns PD, MacNeil MD, Geary TW. Effects of flunixin meglumine and transportation on establishment of pregnancy in beef cows. Journal of Animal Science. 2007; 85:1547-54. https://doi.org/10.2527/jas.2006-587.
- Hockett ME, Rohrbach NR, Schrick FN. Alterations in embryo development in progestogen-supplemented cows administered prostaglandin F2α. Prostaglandins and Other Lipid Mediators. 2004; 73:227-36. https://doi.org/10.1016/j.prostaglandins.2004.02.002.
- Malm H, Borisch C. Analgesics, non-steroidal antiinflammatory drugs (NSAIDs), muscle relaxants, and antigout medications. Drugs During Pregnancy and Lactation: Treatment Options and Risk Assessment: Third Edition, 2015, 27–58. https://doi.org/10.1016/B978-0-12-408078-2.00002-0.
- Kasimanickam R, Kasimanickam V, Gold J, Moore D, Kastelic JP, Pyrdek D *et al.* Injectable or transdermal flunixin meglumine improves pregnancy rates in embryo transfer recipient beef cows without altering returns to estrus. Theriogenology. 2019; 140:8-17. https://doi.org/10.1016/j.theriogenology.2019.08.011.
- 10. Kasimanickam RK, Hall JB, Estill CT, Kastelic JP, Joseph C, Abdel Aziz RL *et al.* Flunixin meglumine improves pregnancy rate in embryo recipient beef cows with an excitable temperament. Theriogenology. 2018; 107:70-7.

https://doi.org/10.1016/j.theriogenology.2017.10.043.

- Amiridis GS, Tsiligianni T, Dovolou E, Rekkas C, Vouzaras D, Menegatos I. Combined administration of gonadotropin-releasing hormone, progesterone, and meloxicam is an effective treatment for the repeatbreeder cow. Theriogenology. 2009; 72:542-8. https://doi.org/10.1016/j.theriogenology.2009.04.010.
- McDougall S, Abbeloos E, Piepers S, Rao AS, Astiz S, van Werven T *et al.* Addition of meloxicam to the treatment of clinical mastitis improves subsequent reproductive performance. Journal of Dairy Science. 2016; 99:2026-42. https://doi.org/10.3168/jds.2015-9615.
- Schlapp G, Goyeneche L, Fernández G, Menchaca A, Crispo M. Administration of the nonsteroidal antiinflammatory drug tolfenamic acid at embryo transfer improves maintenance of pregnancy and embryo survival in recipient mice. Journal of Assisted Reproduction and Genetics. 2015; 32:271-5.
- https://doi.org/10.1007/s10815-014-0378-x. 14. Ferguson CE, Kesler DJ, Godke RA. Improving
- pregnancy rates in problem breeder cattle by administration of 15 mg of progesterone on days 3-5 post-mating. Journal of Applied Animal Research. 2012; 40:173-8.

https://doi.org/10.1080/09712119.2011.645038.

- Binelli M, Thatcher WW, Mattos R, Baruselli PS. Antiluteolytic strategies to improve fertility in cattle. Theriogenology. 2001; 56:1451-63. https://doi.org/10.1016/S0093-691X(01)00646-X.
- 16. Pugliesi G, Shrestha HK, Hannan MA, Carvalho GR, Beg MA, Ginther OJ. Effects of inhibition of prostaglandin F 2α biosynthesis during preluteolysis and luteolysis in heifers. Theriogenology. 2011; 76:640-51. https://doi.org/10.1016/j.theriogenology.2011.03.017.
- 17. Sutton-McDowall ML, Gilchrist RB, Thompson JG. The pivotal role of glucose metabolism in determining oocyte

developmental competence. Reproduction 2010; 139:685-95. https://doi.org/10.1530/REP-09-0345.

- Rato L, Alves MG, Socorro S, Duarte AI, Cavaco JE, Oliveira PF. Metabolic regulation is important for spermatogenesis. Nature Reviews Urology 2012; 9:330-8. https://doi.org/10.1038/nrurol.2012.77.
- 19. Downs SM. The influence of glucose, cumulus cells, and metabolic coupling on ATP levels and meiotic control in the isolated mouse oocyte. Developmental Biology 1995; 167:502-12. https://doi.org/10.1006/dbio.1995.1044.
- Zhang C, Niu W, Wang Z, Wang X, Xia G. The effect of Gonadotropin on glucose transport and apoptosis in rat ovary. PLoS ONE. 2012; 7:42406. https://doi.org/10.1371/journal.pone.0042406.
- 21. Velhankar D. Growth, puberty and sexual maturity in gir heifers consequential to different dietary energy levels with critical studies on blood glucose, copper, cholesterol. KrishikoshEgranth.ac.in
- 22. Hightshoe RB, Cochran RC, Corah LR, Kiracofe GH, Harmon DL, Perry RC. Effects of calcium soaps of fatty acids on postpartum reproductive function in beef cows. Journal of Animal Science 1991; 69:4097-103. https://doi.org/10.2527/1991.69104097x.
- 23. Joe AA, Kathiresan D, Devanathan TG, Rajasundaram RC, Rajasekaran J. Blood biochemical profile in normal cyclical and anoestrus cows. Indian Journal of Animal Sciences. 1998; 68:1154-6.
- 24. Butler WR. Symposium: Optimizing Protein Nutrition For Reproduction And Lactation Review: Effect of Protein Nutrition on Ovarian and Uterine Physiology in Dairy Cattle, 1998, 81 https://doi.org/10.3168/jds.S0022-302(98)70146-8.
- 25. Elrod CC, Butler WR. Reduction of fertility and alteration of uterine pH in heifers fed excess ruminally degradable protein. Journal of Animal Science. 1993; 71:694-701. https://doi.org/10.2527/1993.713694x.
- Kappel LC, Ingraham RH, Morgan EB, Zeringue L, Wilson D, Babcock DK. Relationship between fertility and blood glucose and cholesterol concentrations in Holstein cows. American Journal of Veterinary Research. 1984; 45:2607-12.
- 27. Lager K, Jordan E. The Metabolic Profile for the Modern Transition Dairy Cow. Mid-South Ruminant Nutrition Conference. 2012: 9-16.
- 28. Sreedhar S, Rao KS, Suresh J, Moorthy PRS, Reddy VP, Sreedhar S *et al.* India 44 Sri Venkateswara Veterinary University. Veterinarski Arhiv. 2013; 83:171-87.
- 29. Pandey NKJ, Gupta HP, Prasad S, Sheetal SK. Plasma progesterone profile and conception rate following exogenous supplementation of gonadotropin-releasing hormone, human chorionic gonadotropin, and progesterone releasing intra-vaginal device in repeatbreeder crossbred cows. Veterinary World. 2016; 9:559-62. https://doi.org/10.14202/vetworld.2016.559-562.
- Erdogan HM, Gunes V, Gokce HI, Uzun M, Citil M, Yuksek N. Effects of prolonged intravenous of flunixin meglumine in healthy dogs. Acta Veterinaria Brno. 2003; 72:71-8. https://doi.org/10.2754/avb200372010071.
- 31. Maplesden DC, McSherry BJ, Stone JB. Blood Sugar Levels in Normal Cows before and after Treatment with Prednisolone and Dexamethasone. The Canadian Veterinary Journal La Revue Veterinaire Canadienne. 1960; 1:309-30912.
- 32. Sahu M. Studies on dietary manipulation with PUFA on

embryo production and conception in cattle. G.B. Pant University of Agriculture and Technology, Pantnagar. U.S Nagar, Uttarakhand, 2018.

- Guzel S, Tanriverdi M. Comparison of serum leptin, glucose, total cholesterol and total protein levels in fertile and repeat breeder cows. Revista Brasileira de Zootecnia 2014; 43:643-7. https://doi.org/10.1590/S1516-35982014001200003.
- 34. Maithani M. Studies on improving conception following embryo transfer in cattle, 2017.
- 35. Ahmad I, Lodhi LA, Qureshi ZI, Younis M. Studies on blood glucose, total proteins, urea and cholesterol levels in cyclic, non-cyclic and endometritic crossbred cows. Pakistan Veterinary Journal. 2004; 24:92-4.
- 36. Salazar-Ortiz J, Monget P, Guillaume D. The influence of nutrition on the insulin-like growth factor system and the concentrations of growth hormone, glucose, insulin, gonadotropins and progesterone in ovarian follicular fluid and plasma from adult female horses (*Equus caballus*). Reproductive Biology and Endocrinology 2014; 12. https://doi.org/10.1186/1477-7827-12-72.
- 37. Alameen AO, Abdalla AM. Metabolic and endocrine responses of crossbred dairy cows in relation to pregnancy and season under tropical conditions. American-Eurasian Journal of Agricultural & Environmental Sciences 2012; 12:1065-74. https://doi.org/10.5829/idosi.aejaes.2012.12.08.6619.
- Abd Ellah MR, Hussein HA, Derar DR. Ovarian follicular fluid constituents in relation to stage of estrus cycle and size of the follicle in buffalo. Veterinary World 2010; 3:263-7. https://doi.org/10.5455/vetworld.2010.263-267.
- 39. Ceylan A, Serin İ, Akşit H, Seyrek K, Gökbulut C. Investigation of Vitamins A, E, beta-carotene, cholesterol and triglyceride concentrations in dairy cows with repeat breeder and anestrus. Kafkas Universitesi Veteriner Fakultesi Dergisi, 2007.

https://doi.org/10.9775/kvfd.2007.23-a.

- 40. Ramakrishima KV. Microbial and biochemical profile in repeat breeder cows. Indian Journal of Animal Reproduction. 1997; 17:30-2.
- 41. Singh M, Reprod HP-IJA U. Blood biochemical profile of normal and repeat breeder cows in Himachal Pradesh. Indian Journal of Animal Reproduction. 1998; 19:156-7.
- 42. Pandey NKJ, Gupta HP, Prasad S, Sheetal SK. Plasma progesterone profile and conception rate following exogenous supplementation of gonadotropin-releasing hormone, human chorionic gonadotropin, and progesterone releasing intra-vaginal device in repeatbreeder crossbred cows. Veterinary World. 2016; 9:559-62. https://doi.org/10.14202/vetworld.2016.559-562.
- 43. Radostits O, Gay C, Gay C, Hinchcliff K, Constable P. A Textbook of the Diseases of Cattle, Horses, Sheep, Pigs and Goats: veterinary medicine. Veterinary Medicine. 2007; 10:2045-50.
- 44. Hightshoe RB, Cochran RC, Corah LR, Kiracofe GH, Harmon DL, Perry RC. Effects of calcium soaps of fatty acids on postpartum reproductive function in beef cows. Journal of Animal Science. 1991; 69:4097-103. https://doi.org/10.2527/1991.69104097x.
- 45. Cordeiro MB, Peres MS, de Souza JM, Gaspar P, Barbiere F, Sá Filho MF *et al.* Supplementation with sunflower seed increases circulating cholesterol concentrations and potentially impacts on the pregnancy

rates in Bos indicus beef cattle. Theriogenology. 2015; 83:1461-8.

https://doi.org/10.1016/j.theriogenology.2015.01.022.

- 46. Kulkarni B, Sciences B U. Blood metabolic profiles in crossbred lactating cows. Indian Journal of Animal Sciences. 1993; 63:716-719.
- 47. Memon G. Study on the Blood Composition of Growing Calves and Pregnant and Lactating Cows. Indian Journal of Physiology and Allied Sciences. 1961; 95:1961.
- 48. Sheetal S. Follicular development, superovulation and embryo recovery following insulin and insulin like growth factor-I (IGF-I) treatment during mid Luteal phase of Estrous cycle. G.B. Pant University of Agriculture and Technology, Pantnagar. U.S Nagar, Uttarakhand, 2017.
- 49. Mozaffari AA, Derakhshanfar A. Evaluation of the brain, renal, and hepatic effects of flunixin meglumine, ketoprofen, and phenylbutazone administration in Iranian fat-tailed sheep. Tropical Animal Health and Production. 2011; 43:1389-93. https://doi.org/10.1007/s11250-011-9866-5.
- 50. Carrick JB, Papich MG, Middleton DM, Naylor JM, Townsend HGG. Clinical and pathological effects of flunixin meglumine administration to neonatal foals. Canadian Journal of Veterinary Research. 1989; 53:195-201.
- 51. Singh SP, Kumar A, Bhavsar PP, Sahu M, Kumar P, Kumar S. Evaluation of the Effect of GnRH Analogue, Progesterone and Tolfenamic Acid on Serum Progesterone Profile and Conception Rate in Repeat Breeding Crossbred Cattle. International Journal of Current Microbiology and Applied Sciences. 2020; 9:2630-7.

https://doi.org/https://doi.org/10.20546/ijcmas.2020905.3 01.