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Oxygen uptake of *Olyra longicaudata* (McClelland, 1842) in relation to body weight

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Abstract

Oxygen uptake is a valid measure of metabolism on which production of aquaculture depends. The weight specific oxygen uptake in *Olyra longicaudata* was still unknown. So, the fishes were collected from Chisang river basin, Letang from February to July, 2019. Oxygen uptake rate of the fishes was carried out in laboratory of Post Graduate Campus, Biratnagar conducted by the cylindrical glass respirometer with the aim of estimating oxygen consumption. Data were analyzed from logarithmic transformations of body weight and oxygen uptake expressed by the allometric equation. The total oxygen uptake increased with increasing body weight by a power of 0.580. There was positive correlation coefficient equal to 0.9586 (p<0.01). The weight specific oxygen uptake declined with increasing body weight by power of 0.419 with correlation coefficient equal to 0.9252 (p<0.01). The study was concluded that the specific metabolic rate is higher in smaller than in larger fishes.

Keywords: Chisang, respirometer, metabolic rate, correlation coefficient

Introduction

Oxygen is essential for the function, growth and welfare of animals. Metabolism of the fish is necessary in order to estimate the requirement for promoting and sustaining growth. Regulation of metabolism is directly reflected in respiration. The uptake of respiration by the fish is taken as a measure of its metabolism. Increase in metabolic activity increase the rate of oxygen uptake. Oxygen uptake in fish is influenced by various extrinsic and intrinsic factors viz. temperature, photoperiod, pollutants, season and time, pH, salinity, oxygen, carbon dioxide tensions, body weight, state of activity, nutrition, stage in life cycle, sex, gonadal cycle and hormones etc. ^[1]. The metabolic rate is known to increase with temperature and feeding level ^[2], swimming speed and stress ^[3-6], as well as to decrease with increasing size ^[7]. Killen et al.^[8] studied the intraspecific scaling of metabolic rate in fishes. Kumari et al.^[9] determined the relationship between oxygen uptake and body weight in *Notopterus notopterus* at 24.0 ± 1.0 ⁰C under continuous water flow system. Subba and Ghosh ^[10] studied the oxygen uptake in relation to body weight in a hill stream fish Glyptothorax telchitta from the Saptakoshi river. They found that with a unit increase in body weight, the oxygen uptake per unit time (mlO_2h^{-1}) increases by a power of 0.930 showing significant positive correlation whereas the weight specific oxygen uptake decreases by a power of 0.070 showing significant but negative correlation. Akbulut et al. [11] resolved the oxygen consumption rate of Black Sea Trout (Salmo trutta labrax). Segovia et al. ^[12] studied the oxygen consumption rate in four groups of Oplegnathus insignis under three different water temperature 13 °C, 18 °C and 23 °C. Mohapatra et al. ^[13] determined oxygen consumption of advanced fry and fingerlings of Indian major carps (*Catla catla*, *Labeo rohita* and *Cirrhinus mrigala*). Hallikhed ^[14] gave a detail account on oxygen uptake in relation to temperature variation and showed a gradual increase in the oxygen consumption during heat adaptation and a gradual decrease during cold adaptation. Kumar and Ahsan^[15] studied the bimodal oxygen uptake in an air breathing fish Channa marulius and concluded that with unit increased in body weight, aquatic, aerial and total oxygen uptake increased by an fractional power of 0.8309, 0.7729 and 0.7737 respectively whereas aquatic, aerial and total oxygen uptake per unit body weight decreased by the fractional power of 0.1901, 0.2796 and 0.2302 respectively. Prakoso et al. [16] gave the detailed account on oxygen consumption of *Mugil celphalus* at different temperature (15 °C, $20 \,^{\circ}\text{C}$ and $25 \,^{\circ}\text{C}$) and found that the rate was high at $25 \,^{\circ}\text{C}$.

Olyra longicaudata is rare fresh water hill stream cat fish inhabiting under vegetations and stony beds in shallow water.

They prefer shallow and clear cold running water with high level of dissolved oxygen enriched with larval and aquatic insects, crustaceans, annelids, mollusks etc. ^[17]. The conservation status of the fish is least concern, IUCN 2010. In Nepal, it was reported from Kadya tributary of Trijuga river, Udayapur ^[18] and also found in Chisang river, Letang ^[19].

The knowledge about oxygen uptake in relation to body weight permits the prediction of the oxygen consumption by an animal of given size which may be useful to know the time period for transportation of live fries, fingerlings and adults and also in their storage. So, an attempt was made to know the correlation coefficient between oxygen consumption and body weight of the fish. The study provides information about the fish and might help on its culture and conservation.

Materials and Methods

Study area

The study was carried out in laboratory of Post Graduate Campus, Biratnagar (Alt. 72 MSL, $26^{\circ} 28' 60''$ N & $87^{\circ} 16' 60''$ E,) located in Morang district of Province 1, Nepal. It is 2^{nd} largest industrial and busiest city with population of 2, 40,000 & area of 103.88 km².

Specimen collection

Live twenty five specimens of the *O. longicaudata* of different body weight (0.8g to 5.2g) were collected from Chisang river basin nearby Lamitar, Letang (Alt. 291 MSL, $26^{0}45'46.3"$ N & $87^{0}30'0.15"$ E) with the help of local fisherman using nylon mosquito nets from February to July , 2019.

The collected specimens were acclimatized in a large glass aquarium inside Zoology laboratory of Post Graduate Campus, Biratnagar for a month. The temperature acclimation at each sampling date was similar to that of the natural environment. The experimented fish was undernourished for 12 hours before experimentation. The fishes were weighed before introduced into the experiment. Atmospheric temperature, ambient temperature and operculum frequency were also recorded.

Measurement of oxygen consumption

Among 25 specimens, only 15 specimens were introduced in the experiment. A flow-through cylindrical glass respirometer (24 cm long and 7 cm in diameter) of 722 ml volume, was used in order to measure the inflow and the out flow water. The Winkler's method ^[20] was used to measure the oxygen content of the inspired and expired water. The constant flow of water was maintained in experiment at the rate of 600 ml per hour. Metal clops are used to control the constant flow of water. The oxygen uptake per unit time and body weight was determined by the difference of dissolved oxygen of inspired and expired water and body weight of the fishes. The respirometer was connected to a large water tank with constant water level. The tank was connected to chlorine free tap water. At first rate of flow of water was high and then slowed down. The fish was acclimatized in the respirometer for one hour before experimentation. Visual disturbance was refrained by a black cloth cover over the respirometer. Both ends of the respirometer were connected to the conical flasks from where inspired and expired water were collected for measurement of concentration of the dissolved oxygen content using Winkler's volumetric method ^[20]. Oxygen consumption per unit time (mlO₂/hr) and per unit body weight (mlO₂/g/hr) was calculated by the differences of dissolved oxygen levels between inspired and expired water and body weight of the fish. At every half an hour interval, three readings were taken and then an average value was calculated out of the three readings.

Regression analysis after logarithmic transformation was done to show regression coefficient between oxygen uptake and body weight and also between specific oxygen consumption and body weight. The correlation coefficient and estimated value were calculated by power equation. The expression of the relationship between oxygen uptake and body weight was made by the following allometric equation:

$$VO_2 = {}_aW^b$$

or

 $Log VO_2 = lag a + b log W.$

Where, $VO_2 = oxygen$ uptake per unit time a = intercept b = regression coefficient W= weight of the fish

Statistical analysis

Coefficient of correlation between body weight and oxygen uptake per unit time with number of fishes collected were performed by using Karl Pearson's method. The coefficient of correlation was calculated by using excel software.

Results

The oxygen uptake rate was determined in fifteen weight groups of *O. longicaudata*. The body weight was ranged from 0.8 g to 5.2 g, atmospheric temperatures from 26 $^{\circ}$ C to 30 $^{\circ}$ C, ambient temperature from 25 $^{\circ}$ C to 27 $^{\circ}$ C,operculum frequency per minute from 101 to 123, oxygen consumption (mlO₂/hr) from 0.7044 to 2.032 and weight specific oxygen uptake (mlO₂/g/hr) from 0.8805 to 0.3907 (Table 1). The dissolved oxygen of the ambient water ranged from 4.032mg/l to 5.360mg/l and pH of water was 7.

Table 1: (Oxygen	uptake in	relation	to body	weight
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Body	Atmospheric	Ambient	Operculum		Estimated		
weight (g)	temperature (⁰ C)	temperature (⁰ C)	frequency (per minute)	mlo2/hr	mlo2/g/hr	mlo ₂ /kg/hr	value
0.8	26.5	26.0	123	0.7044	0.8805	880.50	0.7395
1.4	26.0	25.0	122	0.8543	0.6102	610.20	0.9108
1.6	27.5	26.0	122	1.0081	0.6300	630.06	0.9842
1.8	28.0	27.0	122	1.0792	0.5995	599.50	1.0539
2.0	26.0	25.0	122	1.0346	0.5173	517.30	1.1203
2.4	28.5	27.0	120	1.1533	0.4805	480.50	1.2455
2.6	26.5	26.0	119	1.5491	0.5958	595.80	1.3047
2.8	28.0	26.0	119	1.1109	0.3967	396.70	1.3620
3.0	27.0	26.0	116	1.4113	0.4704	470.40	1.4177

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3.2	27.0	26.0	115	1.5110	0.4721	472.10	1.4716
3.4	30.0	26.0	115	1.6802	0.4941	494.10	1.5244
4.0	27.0	26.0	107	1.6400	0.4100	410.00	1.6750
4.2	27.0	26.0	102	1.7541	0.4176	417.60	1.7234
4.6	28.0	27.0	102	1.7877	0.3886	388.60	1.8167
5.2	27.0	27.0	101	2.0320	0.3907	390.70	1.9511
Avg. =2.87	Avg.=27.33	Avg=26.13	Avg.=107	Avg.=1.3540	Avg.= 0.5169	Avg.= 516.93	Avg. = 1.3533

Body weight of fishes(X), oxygen uptake(Y), logX and logY are presented as the parabolic chart for statistical analysis (Table 2).Value of logX, logY and their square and products are increased as increasing body weight.

	Table 2: Parabolic chart							
S.N.	X	logX	logX ²	Y	Logy	logY ²	logX.logY	
1	0.8	-0.09691	0.009392	0.7044	-0.15218	0.023159	0.014748	
2	1.4	0.146128	0.021353	0.8543	-0.06839	0.004677	-0.00999	
3	1.6	0.204120	0.041665	1.0081	0.003504	0.000012	0.000715	
4	1.8	0.255273	0.065164	1.0792	0.033102	0.001096	0.008450	
5	2.0	0.301030	0.090619	1.0346	0.014772	0.000218	0.004447	
6	2.4	0.380211	0.144561	1.1533	0.061942	0.003837	0.023551	
7	2.6	0.414973	0.172203	1.5491	0.190079	0.036130	0.078878	
8	2.8	0.447158	0.199950	1.1109	0.045675	0.002086	0.020424	
9	3.0	0.477121	0.227645	1.4113	0.149619	0.022386	0.071387	
10	3.2	0.505150	0.255177	1.5110	0.179264	0.032136	0.090555	
11	3.4	0.531479	0.282470	1.6802	0.225361	0.050788	0.119775	
12	4.0	0.602060	0.362476	1.6400	0.214844	0.046158	0.129349	
13	4.2	0.623249	0.388440	1.7541	0.244054	0.059563	0.152107	
14	4.6	0.662758	0.439248	1.7877	0.252295	0.063653	0.167210	
15	5.2	0.716003	0.512661	2.0320	0.307924	0.094817	0.220474	
	Σ=43.8	Σ=6.1698	Σ=3.2130	Σ=20.312	Σ=1.7018	Σ=0.4407	Σ=1.09207	

Relationship between body weight and oxygen uptake per unit time (mlO₂/hr) at 26.13±0.16°c

The log/log plot of the oxygen uptake in relation to body weight shows the straight line as shown in fig.1, when different scores were plotted by using least square regression method. The slope of regression line from logarithmic equation was 0.580. The oxygen uptake (mlO₂/hr) increased from 0.7044 to 2.0320 with increasing body weight from 0.8 g to 5.2 g by a power of 0.580. (Figure 1). There was positive significance between oxygen consumption and body weight with the correlation coefficient (r) equal to 0.9586 (p<0.01). The increased oxygen consumption exhibited an exponential relationship with increasing body weight. The value of regression coefficient and intercept was 0.580 and 0.749 respectively.

Here, $VO_2 = {}_aW^b$

Or, $\log VO_2 = \log a + b \log W = \log 0.749 + 0.580 \log W = 0.1255 + 0.580 \log W$

The estimated value for 1 g, 10 g, 100 g and 1000 g the fishes were 0.749, 2.847, 10.826 and 41.160 respectively using power equation $y = 0.749x^{0.580}$.

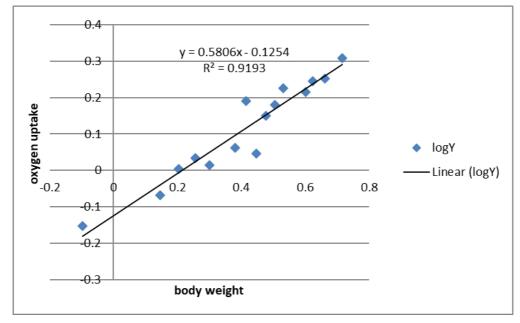


Fig 1: Log/log plots showing the relationship between oxygen uptake and body weight in O. lognicaudata.

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Relationship between body weight and oxygen uptake per unit body weight (mlO₂/g/hr) at 26.13±0.16 °c

Body weight of fishes (X), weight specific oxygen uptake(Y),

logX and logY are presented as the parabolic chart for statistical analysis (Table 3).

S.N.	X	logX	logX ²	Y	logy	logY ²	logX.logY	
1	0.8	-0.09691	0.009392	0.8805	-0.055270	0.003054	0.0053562	
2	1.4	0.146128	0.021353	0.6102	-0.214527	0.046021	-0.031348	
3	1.6	0.204120	0.041665	0.6300	-0.200659	0.040264	-0.040958	
4	1.8	0.255273	0.065164	0.5995	-0.222210	0.049377	-0.056724	
5	2.0	0.301030	0.090619	0.5173	-0.286257	0.081943	-0.861719	
6	2.4	0.380211	0.144561	0.4805	-0.318306	0.101318	-0.121023	
7	2.6	0.414973	0.172203	0.5958	-0.224899	0.050579	-0.093327	
8	2.8	0.447158	0.199950	0.3967	-0.401537	0.161231	-0.179550	
9	3.0	0.477121	0.227645	0.4704	-0.327532	0.107277	-0.156272	
10	3.2	0.505150	0.255177	0.4721	-0.325966	0.106253	-0.164661	
11	3.4	0.531479	0.282470	0.4941	-0.306185	0.093749	-0.162730	
12	4.0	0.602060.	0.362476	0.4100	-0.387216	0.149936	-0.233127	
13	4.2	0.623249	0.388440	0.4176	-0.379239	0.143822	-0.236360	
14	46	0.662758	0 439248	0 3886	-0 410497	0 168507	-0.272060	

0.3907

Σ=7.754

Table 3: Parabolic chart

The weight specific oxygen uptake $(mlO_2/g/hr)$ declined from 0.8805 to 0.3907 with increasing body weight from 0.8 g to 5.2 g (Table 1). The slope of regression coefficient by log-log plot of the relationship between body weight and weight specific oxygen uptake was negative i.e. -0.419. Weight specific oxygen uptake $(mlO_2/g/hr)$ was decreased with increasing body weight by power of 0.419 (Figure 2). The

0.716003

 $\Sigma = 6.1698$

0.512661

Σ=3.2130

5.2

 $\Sigma = 43.8$

correlation coefficient obtained by power equation was negative. It is equal to 0.9252 (p<0.01) which showed significance.

0.166591

 $\Sigma = 1.4699$

-0.292240

 $\Sigma = -2.8967$

Here, $VO_2 = {}_aW^b = 0.749x^{-0.41}$

-0.408156

 $\Sigma = -4.4684$

The estimated value for 1g, 10g, 100g and 1000g fishes were 0.749, 0.2913, 0.1133 and 0.0441 respectively.

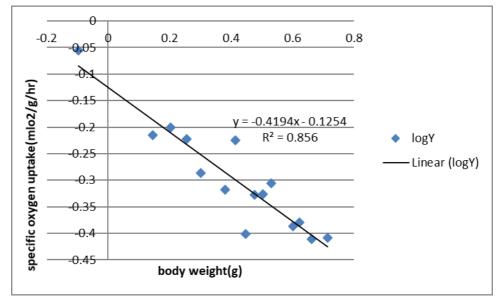


Fig 2: log/log plot showing the relationship between body weight and weight specific oxygen consumption at 26.13±0.16 °c

The oxygen uptake with intercept, slope and correlation coefficient is summarized in table 4.

 Table 4: Oxygen uptake showing intercept, slope and correlation coefficient in O. longicaudata.

Body weight Vs oxygen consumption	Intercept (a)	Slope (b)	Correlation coefficient (r)
VO ₂ (mlO ₂ /hr)	0.749	0.580	0.9586
VO ₂ (mlO ₂ /g/hr)	0.749	-0.419	0.9252

Discussion

The oxygen uptake rate was determined in fifteen weight groups of *O. longicaudata*. The ambient temperature of water

was ranged from 25 0 C to 27 0 C, operculum frequency per minute from 101 to 123 and the dissolved oxygen of water ranged from 4.032mg/l to 5.360 mg/l and pH of water was 7. These characters of water were suitable for the fishes.

The oxygen uptake (mlO_2/hr) increased from 0.7044 to 2.0320 with increasing body weight from 0.8 g to 5.2 g by a power of 0.580. There was positive significance between oxygen consumption and body weight with the correlation coefficient (r) equal to 0.9586 (p<0.01). The increased oxygen consumption exhibited an exponential relationship with increasing body weight. The value of intercept was 0.749.

The weight specific oxygen uptake $(mlO_2/g/hr)$ declined from 0.8805 to 0.3907 with increasing body weight from 0.8 g to

5.2 g. The slope of regression coefficient between body weight and weight specific oxygen uptake was negative i.e. - 0.419. The correlation coefficient obtained by power equation was negative. It is equal to 0.9252 (p<0.01) which showed significance. Weight specific oxygen uptake (mlO₂/g/hr) in the present study was decreased with increasing body weight by power of 0.419. The low exponent value i.e. 0.580 (rate of oxygen uptake) and 0.419 (Weight specific oxygen uptake) indicates that rate of dissolved oxygen consumption by the fish is very slow as it is a catfish.

Several researchers have reported that rate of oxygen consumption increases as the body weight increases ^[21, 22]. The exponent value relating to the oxygen uptake per unit time was always less than 1 as found in most of the fishes by several researchers. The exponent value in the present study was 0.580 which is near to finding of Ruhland ^[23] and Munshi and Dube ^[24]. They reported the exponent value as 0.5. Similarly the value for juveniles of Catla catla and Labeo rohita was recorded as 0.561 and 0.538 respectively ^[25]. Bhattacharya and Subba [26] reported the slope value in Esomus dandricus as 0.5722. In Notopterus notopterus the slope value was 0.421 ^[9] which was lower than the present study. The present value is lower than most of purely water breathing fishes viz 0.64 in mosquito fish Gambusia affinis ^[27], 0.662 in *Mystus cavasius* ^[28], 0.747 in *Catla catla* ^[29] and 0.796 in Cirrhinus mrigala [30]. Hakim et al. [31] in Channa punctatus estimated a slope value of 0.629. In Heteropneustes fossilis the oxygen consumption through gills increased by a power function of 0.799 ^[32]. The relationship between oxygen consumption rate and body size was increased by a power function with an exponent of 0.82 ^[33]. Oxygen uptake in Channa marulius was increased with unit increased in body weight by a fractional power of 0.8309^[15].

As compared to other hill stream fishes, the exponent value in the present fish was lower than others. Subba and Ghosh ^[10] estimated exponent value 0.93 in hill stream fish Glyptothorax telchitta. Similarly the estimated slope value in two hill stream fishes Noemacheilus supicola and Garra lamta was 1.201 and 0.875 respectively ^[34]. Scholander et al. ^[21] suggested (0.83) as the general role for the fishes. Prosser ^[35] suggested the exponent value could range from 0.67 to 1.0. These all differences in slope value of different fish species seemed to be due to different growth pattern, stages of their life cycle, physiological condition of the fish and changes in feeding behavior etc. ^[36, 37, 22]. The weight specific oxygen uptake (mlO₂/g/hr) in present study declined with increasing body weight. The reason may due to the sum of enzymes, metabolites and perhaps inhibitors which govern various metabolic activities of organism, is lower in adult in comparison to growing tissue.

The estimated value of oxygen uptake for 1 g of *O. longicaudata* in the present study was 0.7493 which was higher than most of the air breathing fishes and purely aquatic breathing fishes, *Clarias batrachus* (0.134) ^[38], *Channa marulius* (0.1225) ^[15] and *Notopterus notopterus* (0.508) ^[9]. As compared with other hill stream fishes, the estimated value for 1 g in *Garra lamta* was 0.86 and in *Noemachelius supicola* was 0.532 ^[34] and in *Glyptothorax telchitta* was 0.3202 ^[10]. The value 0.86 is higher and 0.532 & 0.3202 are lower than 0.7493 found in present study (*O. longicaudata*).

Conclusion

The rate of oxygen uptake was increased with increasing body mass but the rate of metabolism declined with increasing

body mass of the fishes. The specific metabolic rate is higher in smaller than in larger fishes.

Breathing frequency, rate of oxygen uptake, mass and size of the fish, temperature, dissolve oxygen and volume of water can predict the time period during transportation of live fishes. Smaller fishes are more preferable in live transportation.

References

- 1. Dejours P. Principles of comparative respiratory physiology. American Elsevier, Amsterdam, New York, North Holland, 1975, 252.
- Sayed-Aly T. Oxygen consumption of Nile Tilapia (*Oreochromis niloticus*) juveniles under different water temperature and feeding frequencies. Egyptian Journal of Aquatic Research. 2007; 33(1):411-417.
- 3. Deitch EJ, Fletcher GL, Petersen LH, Costa I, Shears MA, Driedzic WR *et al.* Cardiorespiratory modification, and limitation, in post-smolt growth hormone transgenic Atlantic salmon *Salmo salar*. Journal of Experimental Biology 2006; 209:1310-1325.
- 4. Wilson C, Friesen E, Higgs D, Farrell A. The effect of dietary lipid and protein source on the swimming performance, recovery ability and oxygen consumption of Atlantic salmon (*Salmo salar*). Aquaculture. 2007; 273:687-699.
- 5. Barnes R, King H, Carter CG. Hypoxia tolerance and oxygen regulation in Atlantic salmon *Salmo salar* from a Tasmanian population. Aquaculture 2011: 318:397-401.
- 6. Folkedal O, Torgersen T, Olsen RE, Ferno A, Nilsson J, Oppedal F *et al.* Duration of effects of acute environmental changes on food anticipatory behavior, feed intake, oxygen consumption, and cortisol release in Atlantic salmon parr. Physiology and Behavior. 2012; 105:283-291.
- Forsberg OI. Modeling oxygen consumption rates of post-smolt Atlantic salmon in commercial –scale, landbased farms. Aquaculture International. 1994; 2(3):180-196.
- Killen SS, Atkinson D, Glazier DS. The scaling of metabolic rate with body mass in fishes depends on lifestyle and temperature. Ecology Letters. 2010; 13:184-193.
- 9. Kumari R, Roy P, Ghosh T. Aquatic Oxygen Uptake of the Freshwater Dual Breathing Featherback, *Notopterus notopterus* (Pallas) Relation to Body Weight. Our Nature 2010; 8(1):131-138.
- 10. Subba BR, Ghosh T. Oxygen uptake in relation to body weight in a Hill stream fish *Gliptothorax telchitta* (Ham.). Nepalese Journal of Biosciences. 2011; 1:11-18.
- 11. Akbulut B, Cakmak E, Kurtoglu IZ, Alkan A. Routine oxygen consumption rate of the Black Sea Trout (*Salmo trutta labrax* Pallas, 1811). Journal of Fisheries Sciences.com 2012; 6(2):88-95.
- 12. Segovia E, Munoz A, Flores H. Water flow requirements related to oxygen consumption in juveniles of *Oplegnathus insignis*. Latin American Journal of Aquatic Research. 2012; 40(3):766-773.
- 13. Mohapatra BC, Das L, Mahanta SK, Sahu H, Sahoo P, Lenka S *et al.* Oxygen consumption in fry and fingerling stages of Indian major carps analyzed using indigenously developed respirometer. Indian Journal of Fisheries. 2017; 64(1):91-94.
- 14. Hallikhed NS. The relationship of temperature on oxygen

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consumption in *Catla catla* (Hamilton). Journal of Bio Innovation. 2018; 7(2):279-285.

- 15. Kumar S, Ahsan W. Studies on metabolic rates in an air breathing fish, *Channa marulius* (Ham.). Proceedings of the Zoological Society of India. 2018; 17(1):61-64.
- 16. Prakoso VA, Kim KT, Ryu JH, Min BH, Chang YJ. Oxygen consumption of *Mugil cephalus* on several temperatures under brackish water conditions. IOP Conference Series. Earth and Environmental Science 2019, 278.
- Kachari A, Gogoi B, Dutta R, Aran K, Ghosh P, Maitra S et al. Habitat preference of an endangered Hill Stream Catfish Olyra longicaudata (McClelland) from Arunachal Pradesh, India. International Journal of Fisheries and Aquatic Studies 2014; 1:86-93.
- Subba BR. Repot on the occurrence of a hill stream fish, *Olyra longicaudata* (McClelland, 1842), Siluriformes, Olyridae from Kadya River of Nepal. Freshwater Biology. 1995; 7(2):155-157.
- Subba BR, Pokharel N, Pandey MR. Ichthyo-faunal diversity of Morang district, Nepal. Our Nature. 2017; 15(1-2): 55-67.
- 20. Welch PS. Limnological methods. Mc Graw Hill Book Co., New York 1948, 370.
- 21. Scholander PF, Flagg W, Walters V, Irving LB. Climatic adaptations in arctic and tropical poikilotherms. Physiological and Biochemical Zoology 1953; 26:67-82.
- 22. Kamler E. respiration in carps in relation to body size and temperature. Polskie Archiwum Hydrobiologii. 1972; 19:325-333.
- 23. Ruhland ML. Etude comparative de la consummation d' chez differentes especes de poisons teleosteans. Bulletin de la Societe Zoologique de France 1965; 90:347-353.
- 24. Munshi JSD, Dube SC. Oxygen uptake capacity of gills in relation to body size of air-breathing fish, *Anabus testudineus* (Bloch). Acta Physiologica Academiae Scientiarum Hungaricae 1973; 44:113-123.
- 25. Jha KK, Roy PK, Ghosh TK. Oxygen uptake in the early developmental stages of two Indian major carps *Catla catla* (Ham.) and *Labeo rohita* (Ham.) in relation to body weight. Journal of Inland Fisheries Society India. 2008; 40(2):7-12.
- 26. Bhattacharya H, Subba BR. Structure of Respiratory Organs of *Esomus dandricus* (Ham.). Our Nature. 2003; 6(1):49-52.
- Mitz SV, Newman MC. Allometric relationship between oxygen consumption and body weight of mosquito fish, *Gambusia affinis*. Environmental Biology of Fishes. 1989; 24(4):267-273.
- Ojha J, Singh R. Oxygen uptake in relation to body weight of the freshwater catfish, *Mystus cavasius* (Ham.). Indian Journal of Experimental Biology 1981; 19:126-129.
- 29. Kunwar GK, Pandey A, Munshi JSD. Oxygen uptake in relation to body weight of two freshwater major carps, *Catla catla* (Ham.) Indian Journal of Animal Science 1989; 59(5):621-624.
- Roy PK, Munshi JSD. Oxygen uptake in relation to body weight and respiratory surface area in *Cirrhinus mrigala* (Ham.) at two different seasonal temperatures. Proceedings of the National Academy of Sciences, Indian Section B. 1984; 50(4):387-394.
- 31. Hakim A, Mushi JSD, Hughes JM. Oxygen uptake from water through the respiratory organs in *Channa punctata*

(Bloch) in relation to body weight. Proceeding of the National Academy of Science, India, Section B. 1983; 49(2):73-78.

- 32. Munshi JSD, Pandey BN, Pandey KP, Ojha J. Oxygen uptake through gills and skin in relation to body weight of an air breathing siluroid fish, *Saccobrabchus* (*Heteropneustes*) *fossilis*. Journal of Zoology. 1978; 184:171-180.
- 33. Urbina MA, Glover CN. Relationship between Fish Size and Metabolic Rate in the Oxyconforming Inanga *Galaxias maculates* Reveals Size-Dependent Strategies to Withstand Hypoxia. Physiological and Biochemical Zoology. 2013; 86(6):740-749.
- Rooj NC. Structure and function of respiratory organs of certain hill stream fishes of Chhotanagpur Division, Ph.D. Thesis, Bhagalpur University, Bhagalpur, India 1984.
- 35. Prosser CL. Oxygen, respiration and metabolism. In Comparative Animal Physiology. 2nd edn (CL Prosser and FA Brown Eds.). WB Saunders and Company, Philadelphia, London. 153-197.
- 36. Smith HW. The metabolism of lung fish. I. General consideration of the fasting metabolism on active fish. Journal of Cellular and Comparative Physiology. 1935a; 6:43-67.
- 37. Smith HW. The metabolism of lung fish. II. Effect of feeding meat on the metabolism rate. Journal of Cellular and Comparative Physiology. 1935b; 6:335-349.
- Munshi JSD, Sinha AL, Ojha J. Oxygen uptake capacity of gills and skin in relation to body weight of the air breathing siluroid fish, *Clarias batrachus* (Linnaeus). Acta Physiologica Academiae Scientiarum Hungaricae, Tomna. 1976; 48(1):28-33.