In vitro assessment of ACE inhibitory activity of A1 and A2 cow milk casein hydrolysate

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Abstract

The aim of the present study was in vitro assessment of ACE inhibition property of A1 and A2 cow milk casein digested with trypsin. Casein peptide was reconstituted (1%, 3% and 5% concentration) and trypsin was added at 1:100 (E:S), hydrolysis was carried out for 6 h at 37 °C and samples were drawn at 2 h intervals from both A1 and A2 casein hydrolysates. Bioactive peptides are encrypted within parent protein and are inactive, upon hydrolysis it become biologically active and shows positive health impact. Both A1 and A2 casein hydrolysates manifested ACE inhibition, whereas improved activity was recorded with progress in time and degree of hydrolysis. The results stipulate that hypertension was partially control by ACE inhibitory properties of casein-derived peptides.

Keywords: A1/A2 cow milk, enzymatic hydrolysis by casein, A1/A2 casein hydrolysate, ACE inhibition

Introduction

Hypertension can lead to cardiovascular disease complexes such as myocardial infarction, stroke, and heart failure; almost one-third of the western population is suffering from hypertension. Numerous lifestyle changes (low salt intake, weight reduction, increased physical activity, and cessation of alcohol consumption) and pharmacological treatments are applied to treat hypertension [1]. In clinical practice, vasodilators, diuretics, calcium channel blockers, angiotensin II receptor blockers, and angiotensin I-converting enzyme (ACE) inhibitors were utilized [2]. ACE inhibitors interfere with the renin-angiotensin and kinin-kallikrein pathway in the body. ACE inhibitory peptides are present in many food protein sources, and among them, milk protein is an immeasurable source of bioactive peptides [3]. Both casein and whey derived peptides have shown ACE inhibitory activity [1]. A product named “casein DP peptio drink” made up of casein hydrolyzed with trypsin was prepared to control the blood pressure [4]. Intake of 20 g of casein hydrolysate per day for 4 weeks would decrease the blood pressure in hypertensive patients [5]. Another product named “C12 Peption” in the Netherlands is composed of tryptic hydrolyzed casein, prescribed in hypertensive animals and humans to control the blood pressure [6, 7]. Calpis, fermented milk, cultured using Saccharomyces cerevisiae and Lactobacillus helveticus, contained two well-known ACE inhibitory peptides, Val-Pro-Pro and ile-Pro-Pro from β-casein and κ-casein, respectively [8]. Cow milk is deemed as a major source of functional peptides with the positive repercussions on the promotion of health and prevention of disease [9]. Upon hydrolysis, the bioactive peptides get released from its native protein [10] and perform multi-functional role like antioxidant [11], antihypertensive [12, 13], antidiabetic [14], anti thrombotic [15], and immunomodulation [16]. Milk-derived bioactive peptides have the potential to be used as an ingredient in functional foods and nutraceuticals. Casein is considered as the major source of indispensable amino acids and has high biological value. Casein consists of four fractions, namely αs1, αs2, β and κ-casein. β-casein consists of 209 amino acids and has two variants A1 and A2 on the basis of the presence of amino acid at 67th position. A1 type consists of histidine at the 67th position whereas A2 consists of proline at the same position.

Therefore, this research designed to investigate the in vitro health-promoting benefits namely ACE inhibition of A1 and A2 cow milk casein digested with trypsin at different casein concentrations, and at a various time of hydrolysis and also the comparison between the casein variant derived from the A1 and A2 cow milk.
Materials and Methods

Milk

Milk was collected from cattle belonging to A1A1 and A2A2 genotyped from Cattle and Buffalo Farm, ICAR-IVRI, Izatnagar. The cattle belonging to Vrindavani breeds (closbred) and Tharparkar breeds (indigenous) were taken as a source of A1 and A2 milk respectively. The cows herd designated as A1A1 and A2A2 was genotyped by AGB Division, ICAR-IVRI, Izatnagar.

Preparation of freeze-dried casein from A1 and A2 genotyped cows milk

The cow milk casein powder was prepared according to the method as narrated by Salami et al. (2011) with slight modification [13]. The fresh cow milk was warmed up to 37 °C and skimming was done by using a manual cream separator at Animal By-Product lab, Division of LPT. The pH of skim milk was reduced to 4.6 with 2% citric acid solution. Then the solution was mixed properly at 37 °C for 30 min and casein was precipitated and separated from whey proteins by centrifugation (10,000 rpm, 60 min, and 4 °C) and then lyophilized (Scan Vac, Cool Safe Freeze Dryers) and stored at -20 °C until use.

Preparation of A1 and A2 casein hydrolysates by enzymatic hydrolysis

Cow milk whole casein solutions (1%, 3% and 5% w/v on total solid basis) were made by reconstituting the casein in distilled water. The casein solution was heated in boiling water bath for 5 min to destroy the microorganisms, if present, which may produce proteolytic enzymes during hydrolysis process and to denature the native enzymes of the milk, if present, as well as to heat denature the proteins, which increase its susceptibility to proteolytic enzymes. Enzyme/substrate ratio of 1:100 (w/w) was kept constant for trypsin. The temperature and pH of trypsin were adjusted to 37 °C and 8.0 respectively. The hydrolysis was achieved by incubating the samples at 37 °C for 15 min, 30 min, 60 min, and 110 µl of Angiotensin I and phenylalanine were the most effective at the C-terminal position, with proline unexpectedly binding well to ACE [20]. Peptic hydrolyzed bovine casein had higher ACE inhibitory activity than whole casein, attributed to the presence of lower molecular weight peptides [19]. Proline, tryptophan, tyrosine, and phenylalanine were the most effective at the C-terminal position, with proline unexpectedly binding well to ACE [20]. Peptide hydrolyzed bovine casein had higher ACE inhibitory activity than whole casein, attributed to the presence of lower molecular weight peptides [19].

Results and Discussion

Determination of In vitro antihypertensive activity of casein hydrolysate

Angiotensin I-converting enzyme (ACE) inhibition

ACE inhibitory activity of casein hydrolysates were determined as per Cushman and Cheung (1971) [18]. 20 µl of hydrolysate samples mixed with 20 µl of an ACE solution. Following preincubation at 37 °C for 15 min, 110 µl of Hippuryl-Histidyl-Leucine (HHL) substrate solution in 0.1 M sodium borate buffer comprising 0.3 M NaCl, and pH 8.3 was added. After incubating at 37 °C for 60 min, the reaction was stopped, by adding 60 µl of 1 N HCl. To extract hippuric acid, add 1ml of ethyl acetate to the mixture and was centrifuged at 3000 rpm for 10 min. Then, 750 µl of the upper organic phase was evaporated by heating at 95 °C for 30 min in a water bath. The dried material was dissolved in 1 ml of distilled water, and the absorbance was measured at 228 nm using a spectrophotometer. The control sample was prepared by replacing the test sample with distilled water. The

inhibition (%) was calculated as below:

\[
\% \text{ACE inhibition} = \frac{\text{Absorbance}_{\text{control}} - \text{Absorbance}_{\text{sample}}}{\text{Absorbance}_{\text{control}}} \times 100
\]

Statistical analysis

Data are presented as mean±standard error (SE) of nine independent experiments. Data were analyzed using the SPSS package (SPSS 20, Version 20, IBM, USA). The difference between the mean was compared using a t-test. Further, Significant differences were determined using Tukey tests (p<0.05). Data obtained from the experiments were pooled. Firstly A1 and A2 samples were compared within the group based on the time of hydrolysis and then based on casein concentration furthermore a comparison was made between A1 and A2 at the same time period of hydrolysis.

Moreover, among different casein concentrations at the same time interval showed a significant (p<0.05) increase in ACE activity of A1 casein hydrolysates at 0 h of hydrolysis in case of 1% and 3% casein, afterward increase was not significant (p>0.05). However, in the case of A2 casein hydrolysate a significant (p<0.05) increase in activity at 2 h and 6 h of hydrolysis. A1 and A2 casein hydrolysate at 5% casein level showed a significant (p<0.05) increase at 2 h and 4 h followed by a non-significant (p>0.05) increase thereafter. Results indicated that the ACE inhibitory activity of casein hydrolysates increased with an increase in hydrolysis time and protein concentration (Fig. 1.).

In the case of 1%, 3%, and 5% casein, the distinctions between A1 and A2 casein hydrolysates did not vary significantly (p>0.05) at 0 h and 2 h thereafter, a significant difference (p<0.05) was discerned at 4 h and 6 h of hydrolysis with higher activity toward A2 casein hydrolysate (Table 1).

Peptide hydrolyzed bovine casein had higher ACE inhibitory activity than whole casein, attributed to the presence of lower molecular weight peptides [19]. Proline, tryptophan, tyrosine, and phenylalanine were the most effective at the C-terminal position, with proline unexpectedly binding well to ACE [20], hence, A2 casein hydrolysate showed higher activity than A1 casein hydrolysate. Proline-containing peptides are more resistant to hydrolysis by digestive enzymes, thus sustained for a longer period in the digestive tract and exert in vivo bioactivity [21]. Casein is superior to whey protein and showed >80% inhibition since casein is rich in proline that potentiates the activity of casein hydrolysate [22]. Tryptic milk protein hydrolysate showed higher ACE inhibitory activity due to the occurrence of the positively charged amino acid at

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the ultimate C-terminal position [23]. Lower activity of A1 variant than the A2 variant of β-casein was reported on trypsin digestion [23]. After oral feeding of tryptic digests of casein in spontaneous hypertensive rats, the decrease in blood pressure was reported [11]. Trypsin digested casein showed maximum inhibition in contrast to the papain and pancreatin digest further, a slower activity rate was observed after 1 h of hydrolysis [24]. IC50 value (ACE inhibition) of bovine κ-casein and ovine S2-casein hydrolysates was 9.97 and 41.8 µg/ml, respectively [25]. Several ACE inhibitory peptides were extracted from α-casein and β-lactoglobulin [12, 26]. Petrat-Melin et al. (2014) reported maximum ACE inhibition in the A1 and B variant as compared to the A2 and I variant of β-casein hydrolyzed with pepsin and pancreatic enzyme [27].

**Table 1:** ACE inhibitory potential of A1 and A2 casein hydrolysates with different casein concentrations at various time periods of hydrolysis (Mean±SE)

<table>
<thead>
<tr>
<th>Casein concentration</th>
<th>0 h</th>
<th>2 h</th>
<th>4 h</th>
<th>6 h</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>A1</td>
<td>A2</td>
<td>A1</td>
<td>A2</td>
</tr>
<tr>
<td>1% casein</td>
<td>47.54±0.41</td>
<td>48.15±0.39</td>
<td>62.14±0.42</td>
<td>62.61±0.43</td>
</tr>
<tr>
<td>3% casein</td>
<td>52.25±0.31</td>
<td>52.60±0.30</td>
<td>82.14±0.34</td>
<td>83.24±0.38</td>
</tr>
<tr>
<td>5% casein</td>
<td>53.56±0.42</td>
<td>53.96±0.31</td>
<td>84.59±0.44</td>
<td>84.92±0.26</td>
</tr>
</tbody>
</table>

(n=9); *significant (p<0.05) difference between A1 and A2 samples

Mean±SE with different superscripts row wise (small alphabets show difference among A1 samples; capital alphabets show difference among A2 samples) and column wise (number show difference among concentration) differ significantly (p<0.05)

**Fig 1:** ACE inhibition of A1 and A2 casein hydrolysates with different casein concentrations at various time periods of hydrolysis

**Conclusion**

The result of present study demonstrates that hydrolysis of A1 and A2 casein by the trypsin resulted in production of peptides with inhibitory properties against ACE and activity get improved with time of hydrolysis. These finding suggested that both A1 and A2 casein hydrolysates with such inhibitory traits may potentially improve the blood pressure regulation by means of their ability to inactivate the ACE enzyme. However, further study is required for a better understanding of the molecular mechanisms of action of the defined peptides. Besides, clinical studies are certain to validate the efficiency and bioavailability of casein-derived peptides in humans.

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**Conflicts of interest**

There are none potential conflicts between authors and others that bias our work.

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