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Jumade Pratibha

Department of Parasitology, College of Veterinary Science & Animal Husbandry, Anjora, Durg, Chhattisgarh, India

Pal S

Department of Parasitology, College of Veterinary Science & Animal Husbandry, Anjora, Durg, Chhattisgarh, India

Corresponding Author: Jumade Pratibha Department of Parasitology, College of Veterinary Science & Animal Husbandry, Anjora, Durg, Chhattisgarh, India

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Influence of temperature and humidity on conidial germination and colony growth of entomopathogenic fungi

Jumade Pratibha and Pal S

Abstract

The effect of temperature and humidity on conidial germination and growth performance of four isolates of entomopathogenic fungi isolated from organic environment of Durg District of Chhattisgarh namely; *Metarhizium majus* as well as from naturally infected ticks namely; *Fusarium beomiforme, Aspergillus amstelodami* and *Beauveria bassiana* were studied in Potato Dextrose Agar media at 4 °C, 16 °C, 29 °C, 40 °C temperature and 75% relative humidity. Significant colony diameter was observed as 9.0 cm and 9.0 cm of *Fusarium beomiforme,* 4.0cm and 4.5cm of *Aspergillus amstelodami,* 9.0 cm and 9.0 cm of *Beauveria bassiana,*7.0 cm and 9.0 cm of *Metarhizium majus* at 16 °C and 29 °C temperature respectively. The lowest growth of fungal isolated was observed at 4 °C followed by 40 °C. The best suitable temperature for conidial germination and growth of entomopathogenic fungi was observed as 29 °C and 75% relative humidity.

Keywords: entomopathogenic, fungi, growth, temperature, humidity, conidia

Introduction

Control of ticks using chemical acaricides was considered as one of the best methods, but ticks have developed resistance against a range of acaricides (Martins *et al.*, 1995)^[9]. Acaricides as an artificial organic compounds can remain in the environment for many years and may be transported over a long distance (Kunz and Kemp, 1994)^[6]. More emphasis is given nowadays on use of biological control agents to overcome the environmental pollution and food safety. Entomopathogenic fungi are considered as natural mortality agents for insect pests. As per the recent and existing research, the entomopathogenic fungi show minimal adverse effects on the animals and other non-target organisms. They can be used in integrated pest management replacing the conventional chemical insecticides (Pell *et al.*, 2001)^[12]. The present research work was conducted to study the effects of various range of temperature on growth performance of isolates the entomopathogenic fungi explored from organic environment as well as from naturally infected ticks from Durg district of Chhattisgarh.

Materials and Methods

The exploration of entomopathogenic fungi was carried from agricultural soil under cultivation of paddy and maize from Durg district of Chhattisgarh. The isolation of fungi from soil samples were enumerated by using serial soil dilution and soil plate method. (Waksman,1922)^[15]. The serially diluted soil samples at the concentration of 10² dilutions were cultivated on Potato Dextrose Agar medium by incubating at 29 °C temperature and 75% relative humidity in BOD incubator until the full mycelial growth was achieved. The surface sterilized ticks were exposed to treatment of conidial suspension of fungal isolates explored from soil samples. The fungi causing mortality in ticks were separated and assumed as entomopathogenic fungi isolated from soil samples. The ticks collected from body of animals were observed for natural fungal infection and growth of fungal hyphae on their body surfaces. The fungal colonies infecting ticks were collected, cultivated, pure culture was maintained and assumed as entomopathogenic fungi isolated from naturally infected ticks. The species confirmation of entomopathogenic fungi was carried out by PCR using ITS gene as a molecular marker.

In replicates of five, the entomopathogenic fungus was grown on Potato Dextrose Agar medium at 16 $^{\circ}$ C, 29 $^{\circ}$ C and 40 $^{\circ}$ C temperature and 75% relative humidity in BOD incubator.

The plates were maintained in refrigerator by adjusting the temperature of 4 °C. An agar disc of 4.0 mm plug was scooped from the periphery of 4 week old mother cultures and placed at the center of the media plates (90 mm) and incubated at 4 °C, 16 °C, 29 °C and 40 °C. Two measurements of colony diameter in 90 mm petri plates of mycelial spread were recorded daily till the plates of one of the temperatures were full-grown and daily mean diameter was worked out. The data was subjected to statistical analysis by employing WASP (ICAR).

Results and Discussions

In vitro growth profiles of entomopathogenic fungi, belonging to *Fusarium beomiforme, Aspergillus amstelodami, Beauveria bassiana* and *Metarhizium majus* in PDA plates incubated at 4 °C, 16 o C, 29 °C and 40 °C and 75% relative humidity were recorded and are expressed as diameter in cm (Table 1-2; Figure 1-4; Plate 1-4).

Fusarium beomiforme

Maximum colony diameter of *Fusarium beomiforme* was observed as 0.2cm, 9.0 cm, 9.00 cm and 1.0 cm at 4 °C, 160 C, 29 °C and 40 °C temperature respectively, on 10^{th} day of incubation. The maximum diameter of 9.0 cm was observed at 16 °C and 29 °C whereas lowest diameter of 0.2 cm was observed at 4 °C. The statistically significant higher conidial growth of *Fusarium beomiforme* was observed at 16 °C and 29 °C on 10th day of incubation. Maina *et al.* (2017) ^[8], recorded the colony diameter of 71 to 85 mm in 90 mm petri plates of *Fusarium oxysporum f. sp. phaseoli* isolates at 7th day of growth was maximum when incubated at 25-26 °C on PDA whereas Desai *et al.* (2003) ^[2], reported maximum growth of *F. oxysporum f. sp. Ricini* that *F. oxysporum f. sp. Ricini* showed maximum growth and sporulation at 27 ± 2 °C on PDA. According to Daami- Remadi *et al.* (2006) ^[1], *F. solani* was the most aggressive at temperatures superior to 30 °C. Gupta *et al.* (2010) ^[5] reported that the temperature range of 25–35 °C is favorable for the growth and sporulation of *Fusarium* spp.

Aspergillus amstelodami

The isolates of *Aspergillus amstelodami* showed maximum colony diameter of 0.00 cm, 4.0 cm, 4.5 cm and 1.5 cm at 4 $^{\circ}$ C, 160 C, 29 $^{\circ}$ C and 40 $^{\circ}$ C temperature respectively, on 10th day of incubation. The higher growth of *Aspergillus amstelodami* was observed at 29 $^{\circ}$ C on 10th day of incubation. According to Mannaa and Kim (2018) ^[7], while observing the effect of different temperatures (10, 20, 30, and 40 $^{\circ}$ C) and relative humidities (RHs; 12, 44, 76, and 98%) on grain fungi (*Aspergillus candidus, Aspergillus flavus, Aspergillus fumigatus*) were significantly enhanced by both increased temperature and RH.

Table 1: Colony diameter in cm. (Mean ± SEM) of Fusarium beomiforme and Aspergillus amstelodami in Potato Dextrose Agar (PDA) platesincubated at 4 °C, 16 °C, 29 °C and 40 °C temperature and 75% relative humidity

Time (Day)	<i>Fusarium beomiforme</i> Temperature °C				
	1	0.00±0.00	0.00 ± 0.00	0.00 ± 0.00	0.00 ± 0.00
2	0.00±0.00	0.90 ± 0.08^{b}	0.50±0.00	0.00 ± 0.00	
3	0.00±0.00	1.5±0.19 ^b	1.0±0.14 ^{2b}	0.00±0.00	
4	0.00±0.00	2.5±0.08 ^b	2.0±0.14 ^b	0.00±0.00	
5	0.00±0.00	3.8±0.14 ^b	4.0±0.16 ^b	0.00 ± 0.00	
6	0.00±0.00	5.0±0.16 ^b	4.8±0.08 ^b	0.00 ± 0.00	
7	0.00±0.00	5.8±0.16 ^b	6.0±0.20 ^b	0.00±0.00	
8	0.00±0.00	7.8±0.10 ^b	6.0±0.18 ^b	0.5 ± 0.06^{b}	
9	0.2±0.06 ^a	8.8 ± 0.08^{b}	7.8±0.28 ^b	1.0 ± 0.08^{b}	
10	0.2±0.04 ^a	9.0±0.16 ^b	9.0±0.30 ^b	1.0±0.09 ^b	
T	reatments found Sig	nificant at 5% level of	of Significance CD(0.0	05)= 0.044	
T*	Aspergillus amstelodami				
Time	Temperature °C				
(Day)	4 °C	16 °C	29 °C	40 °C	
1	0.00±0.00	0.00 ± 0.00	0.00 ± 0.00	0.00 ± 0.00	
2	0.00±0.00	0.90±0.14 ^b	0.500±0.00	0.50±0.14 ^{ab}	
3	0.00±0.00	1.5±0.12 ^b	0.60±0.14 ^b	0.50±0.24 ab	
4	0.00±0.00	1.5±0.18 ^b	1.0±0.20 ^b	0.50±0.11 ab	
5	0.00±0.00	1.8±0.24 ^b	1.5±0.18 ^b	0.50±0.20 ^{ab}	
6	0.00±0.00	1.8±0.42 ^b	2.0±0.20 ^b	0.50±0.46a ^b	
7	0.00±0.00	2.0±0.32 ^b	2.5±0.18 ^b	0.80±0.18 ^{ab}	
8	0.00±0.00	2.8 ± 0.18^{b}	2.8±0.24 ^b	1.0±0.32 ab	
9	0.00±0.00	3.8 ± 0.32^{b}	3.6±0.20 ^b	1.0±0.18a ^{ab}	
10	0.00±0.00	4.0 ± 0.46^{b}	4.5±0.26 ^b	1.5±0.46 ^{ab}	
T	reatments found Sig	nificant at 5% level of	of Significance CD(0.0)5)= 2.599	

 Table 2: Colony diameter in cm. (Mean ± SEM) of *Beauveria bassiana* and *Metarhizium majus* in Potato Dextrose Agar (PDA) plates incubated at 4 °C, 16 °C, 29 °C and 40 °C temperature and 75% relative humidity

Time (Day)	Beauveria bassiana Temperature °C				
	1	0.00±0.00	0.00±0.00	0.00±0.00	0.00±0.00
2	0.00±0.00	0.90±0.18 ^b	0.50±0.22 ^b	0.50±0.08 ^b	
3	0.00±0.00	1.5±0.05 ^b	1.0±0.10 ^b	1.0±0.02 ^b	
4	0.00±0.00	2.5±0.22 ^b	2.0±0.12 ^b	1.0±0.02 ^b	
5	0.00±0.00	3.8±0.26 ^b	3.0±0.22 ^b	1.0±0.12 ^b	
6	0.00±0.00	5.0±0.54 ^b	4.8±0.12 ^b	1.5±0.36 ^b	
7	0.00±0.00	5.8±0.20 ^b	5.0±0.66 ^b	1.5±0.29 ^b	
8	0.1±0.12 ^a	6.8±0.12 ^b	6.0±0.92 ^b	2.0±0.42 ^b	
9	0.1±0.10 ^a	7.0±0.32 ^b	7.2±0.41 ^b	2.2±0.34 ^b	
10	0.1±0.08 ^a	9.0±0.38 ^b	9.0±0.18 ^b	2.5±0.22 ^b	
Treatments fo	ound Significant at 1	% and 5% level of sig	nificance CD(0.01) = 0.0	031; CD(0.05) = 0.023	
Time (Day)	Metarhizium majus				
	Temperature °C				
	4 °C	16 °C	29 °C	40 °C	
1	0.00±0.00	0.90±0.18 ^b	0.50±0.08 ^b	0.00±0.00	
2	0.00±0.00	1.5±0.22 ^b	1.0±0.02 ^b	0.00±0.00	
3	0.00±0.00	2.5±0.18 ^b	2.0±0.22 ^b	0.00±0.00	
4	0.1±0.12 ^a	3.8±0.28 ^b	3.0±0.46 ^b	0.00±0.00	
5	0.1±0.16 ^a	4.0±0.22 ^b	4.8±0.52 ^b	0.00±0.00	
6	0.2±0.22 ª	4.0±0.18 ^b	5.0±0.18 ^b	0.00±0.00	
7	0.2±0.28 ^a	4.8±0.32 ^b	6.0±0.32 ^b	0.00±0.00	
8	0.2±0.38 ^a	5.8±0.36 ^b	7.8±0.43 ^b	1.0±0.52 ^b	
9	0.2±0.42 ^a	6.8±0.38 ^b	9.0±0.52 ^b	1.0±0.22 ^b	
10	0.2±0.32 ^a	7.0±0.22 ^b	9.0±0.56 ^b	1.0±0.18 ^b	
Treatments	found Significant at 1	% and 5% level of sign	ficance $CD(0.01) = 0.05$	6; CD(0.05) = 0.042	

Beauveria bassiana

The growth profile of Beauveria bassiana was observed as 0.1cm, 9.0 cm, 9.00 cm and 2.5 cm at 4 °C, 16 °C, 29 °C and 40 °C temperature respectively, on 10th day of incubation. Maximum colony diameter of Beauveria bassiana was observed as 9.0 cm at 16 °C and 29 °C temperature with conidia germination. Similarly in accordance with the present findings, Fergues et al. (1997) [4], observed that the temperature range of 16 °C to 29 °C was appropriate for maximum growth of B. bassiana isolates. Niranjana, (2004) ^[11] and Mwamburi *et al.* (2015) ^[10], reported that the optimal temperature for conidial germination of B. bassiana isolates was approximately 25 °C with an upper limit at 30 °C was able to germinate rapidly in a broad temperature range of 25-30 °C after 24 h. Ekesni et al. (1999) ^[3] and Soundarapandian and Chandra, (2007) ^[13], observed the optimum temperature for germination, radial growth and pathogenic activity of EPF ranged between 25-30 °C. The in vitro conidial germination of myco insecticides Beauveria bassiana, and Metarhizium anisopliae was slower at 10 and 15 °C than at 20 and 25 °C.

Metarhizium majus

The effect of temperature on Metarhizium majus in PDA was observed as 0.2 cm, 7.0 cm, 9.00 cm and 1.0 cm colony diameter at 4 °C, 16 °C, 29 °C and 40 °C temperature, respectively on 10th day of incubation. Maximum conidial germination and colony diameter of 9.0 cm of Metarhizium majus was achieved at 29 °C whereas lowest diameter of 0.2 cm was observed at 4 °C at 40 °C temperature. Yeo et al. (2003) ^[16], observed the reduction of conidial germination and colony growth of *M. anisopliae* isolates at 10 °C. The maximum radial growth of *M. anisopliae* was observed at 30 °C followed by 25 °C. At 35 °C retarded vegetative radial growth of all the fungal isolates was observed which were failed to grow or form conidia at 40 and 45 °C. The results of present investigation were in accordance with the findings of Teja and Rahman (2016)^[14]. From the results of growth of EPF in different temperature suggested that the temperature plays an important role in growth of entomopathogenic fungi. The very high and very low temperature may affect the conidial germination and fungal growth. The temperature range of 25-30 °C was found to be optimal for growth of fungi.

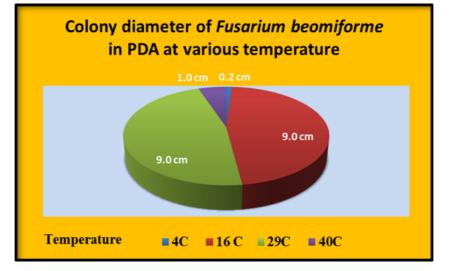


Fig 1: Colony growth of Fusarium beomiforme at various temperature

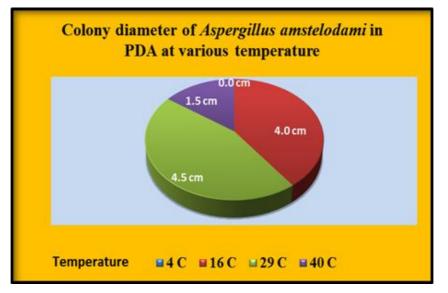


Fig 2: Colony growth of Aspergillus amstelodami at various temperature

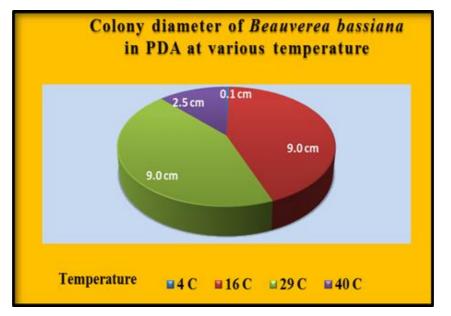


Fig 3: Colony growth of Beauveria bassiana at various temperature

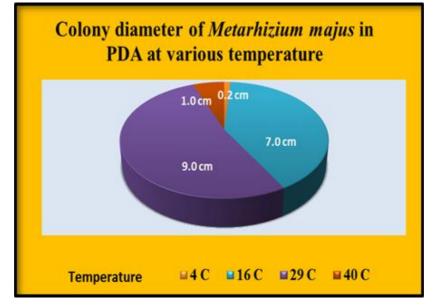


Fig 4: Colony growth of Metarhizium majus at various temperature



Plate 1: Colony diameter of Fusarium beomiforme in Potato Dextrose Agar (PDA), incubated at 4 °C, 16 °C, 29 °C and 40 °C temperature.



Plate 2: Colony diameter of Aspergillus amstelodami in Potato Dextrose Agar (PDA), incubated at 4 °C, 16 °C, 29 °C, 40 °C temperature and 75% relative humidity



Plate 3: Colony diameter of Beauveria bassiana in Potato Dextrose Agar (PDA), incubated at 4 °C, 16 °C, 29 °C, 40 °C temperature and 75% relative humidity



Plate 4: Colony diameter of *Metarhizium majus* in Potato Dextrose Agar (PDA), incubated at 4 °C, 16 °C, 29 °C, 40 °C temperature and 75% relative humidity

References

- 1. Dammi-Remadi M, Jabnoun-Khiareddine H, Ayed F, EI Mahjoub M. Effect of temperature on aggerressivity of Tunisian *Fusarium sp.* causing potato (*Solanum tuberosum* L) tuber dry rot. J. Agron 2006;5(2):350-355.
- 2. Desai AG, Dange SRS, Patel DS, Patel DB. Variability of *Fusarium oxysporum f. sp. ricini* causing wilt of castor .J. Mycol. Plant Pathol 2003;33(1):37-41.
- 3. Ekesni S, Maniana K, Ampong Nyarko. Effect of Temperature on Germination, Radial Growth and Virulence of *Metarhizium anisopliae* and *Beauveria bassiana* on *Megalurothrips sjostedti*. Journal Biocontrol Science and Technology 1999.
- Fargues J, Goettel MS, Smits N, Ouedraogo A, Rougier M. Effect of Temperature on Vegetative Growth of *Beauveria bassiana* Isolates from Different Origins. Mycologia 1997;89(3):383-392
- Gupta, Vijai Kumar, Misra, Ashok Kumar, Gaur Rajarshi Kumar. Growth Characteristics of *Fusarium* Spp. Causing Wilt Disease in *Psidium guajava* L. in India. Journal of Plant Protection Research 2010;50(4):452-462.
- 6. Kunz SE, Kemp DH. Insecticides and acaricides: resistance and environmental impact 1994;13(4):1249-1286.
- Mannaa Mohamed, Kim Deok Ki. Effect of temperature and relative humidity on Growth of Aspergillus and *Penicillium* spp. and Biocontrol activity of *Pseudomonas* protegens AS15 against Aflatoxigenic Aspergillus flavus in Stored Rice Grains. Mycobiology. 2018;46(3):287-295.
- 8. Maina PK, Wachira PM, Okoth SA, Kimenju JW, Cultural. Morphological and Pathogenic Variability among *Fusarium oxysporum f. sp. phaseoli* Causing Wilt in French Bean (*Phaseolus vulgaris* L.). Journal of Advances in Microbiology 2017.
- Martins JR, Correa BL, Cereser VH, Arteche CCP. A situation report on resistance to acaricides by the cattle tick *Boophilus microplus* in the state of Rio Grande do Sul, southern Brazil. In C. Rodriguez, D. Sergio & H. Fragoso, eds. 3rd International seminary on Animal Parasitology. Acapulco, Guerrero, Mexico 1995.
- 10. Mwamburi Lizzy A, Laing Mark D, Miller Ray M. Effect of surfactants and temperature on germination and vegetative growth of Beauveria bassiana Braz. J. Microbiol. 2015;46(1).
- 11. Niranjana SR. Exploitation of entomopathogenic fungus *Beauveria bassiana* for efficient control of coffee berry borer in India. J Mycol Plant Pathol 2004;34(3):714-723.
- 12. Pell JK, Eilenberg J, Hajek AE, Steinkraus DC, Butt TM, Jackson C *et al*. Fungi as biocontrol agents: Progress, Problems and Potential. CAB International, Wallingford,

2001, 71-153.

- Soundarapandian P, Chandra R. Mass Production of Endomopathogenic Fungus *Metarhizium anisopliae* (Deuteromycota; Hyphomycetes) in the Laboratory. Research Journal of Microbiology 2007;2(9):690-695
- 14. Teja KNP, Chandra, Rahman SJ. Characterisation and evaluation of *Metarhizium anisopliae* (Metsch.) Sorokin strains for their temperature tolerance. An International Journal on Fungal Biology. 2016;7(4):171-179.
- 15. Waksman SA. A method for counting the number of fungi in the soil. J. Bact 1922;7(3):339-341.
- 16. Yeo H, Pell JK, Alderson PG, Clark SJ, Pye BJ. Laboratory evaluation of temperature effects on the germination and growth of entomopathogenic fungi and on their pathogenicity to two aphid species. Pest Manag Sci 2003;59(2):156-65.