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## Bio-efficacy of biocontrol agents against Fall armyworm *Spodoptera frugiperda* (J. E. Smith) under laboratory conditions

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### Abstract

Laboratory bioefficacy of six test bioagents viz., entomopathogenic fungi (*Metarhizium anisopliae*, *Nomjraea rileyi*, *Beauveria bassiana*), entomopathogenic bacteria *Bacillus thuringiensis*, entomopathogenic nematode *Steirnerernema carpocapsae* and botanicals *Azadirachtin* were evaluated against 1<sup>st</sup>, 3<sup>rd</sup>, 5<sup>th</sup> instar larva of fall armyworm at 1<sup>st</sup>, 3<sup>rd</sup> 5<sup>th</sup> and 7<sup>th</sup> days after treatment. Among all evaluated bioagents treatment with *Bt* showed the highest mortality i.e 85.92%, 64.44% and 50.00% against 1<sup>st</sup>, 3<sup>rd</sup>, and 5<sup>th</sup> instar larvae, respectively. The next best treatment was *Nomurea rileyi* showed 71.48%, 57.04% and 36.68% mortality against 1<sup>st</sup>, 3<sup>rd</sup> and 5<sup>th</sup> instar larvae respectively. In present study Bioefficacy depicted as *Bt* > *N. rileyi* > *M. anisopliae* and *Azadirachtin* > *S. carpocapsae* and *B. bassiana* against 1<sup>st</sup>, 3<sup>rd</sup> and 5<sup>th</sup> instar larvae of fall armyworm.

**Keywords:** entomopathogenic fungi, entomopathogenic bacteria, entomopathogenic nematode, botanicals, fall armyworm *Spodoptera frugiperda*

### Introduction

The Fall armyworm *Spodoptera frugiperda* is an economically important pest native to tropical and subtropical America has recently invaded India, causing more damage to maize and sorghum. It is a notorious pest with high dispersal ability, wide host range and high fecundity makes the fall armyworm one of the severe economic pest. Pest management in agriculture is a challenging task in the context of increasing agricultural productivity without upsetting the ecological balance and deteriorating the environment. Chemical insecticides in agriculture are useful for protecting crop against pests and play the significant role to boost the production. To obviate the effects of chemical insecticides, there has been increased demand for the alternative and selective pest control agents particularly bioagents that in turn are silent workers from nature. Biological control is regarded as more beneficial than pesticide-based control due to their target specificity, eco-safety, reduced number of applications, yield and quality improvement, higher acceptability and export value of produce and suitability for rural areas. Several biopesticides with novel mode of action are now available in the market and therefore, it is necessary to use safe, effective, ecologically sound biocontrol agents. Due to the seriousness of the pest problem and relative paucity of information regarding pest management the present investigations were conducted with objective to study bioefficacy of different biocontrol agents against *S. frugiperda* under laboratory conditions.

### Materials and Methods

The efficacy of test bioagents were evaluated on larvae by adopting the leaf dip method (Ahmad *et al.*, 1995). Tender succulent maize leaves were brought and after thorough cleaning with water. The leaves were dipped in requisite concentration of bioagents for 10 seconds. The leaves were air dried under ceiling fan for 4 hr and then the leaves were placed in each plastic container. Ten larvae were randomly selected from nucleus culture and then were placed in each plastic container. Larval mortality was recorded after every 1, 3, 5 and 7 DAT. The moribund larvae were considered as dead. Mean larval mortality was computed for larval instar. The data were subjected to the arc sin transformation and statistical analysis thereafter.

**Table 1:** Bioefficacy of test bioagents against first instar larvae

TN	Treatments	Per cent Mortality at Days After Treatment			
		1 DAT	3 DAT	5 DAT	7 DAT
T <sub>1</sub>	<i>Metarhizium anisopliae</i> @ (1 x 10 <sup>8</sup> cfu/ml) 5g / l	3.33 (8.06)*	13.33 (21.14)	42.59 (40.69)	67.77 (55.42)
T <sub>2</sub>	<i>Beauveria bassiana</i> @ (1 x 10 <sup>8</sup> cfu/ml) 5g / l	0.00 (2.87)	6.67 (13.25)	31.85 (34.21)	38.89 (38.31)
T <sub>3</sub>	<i>Nomuraea rileyi</i> @ (1 x 10 <sup>8</sup> cfu/ml) 5g / l	3.33 (8.06)	17.41 (24.05)	53.33 (46.92)	71.48 (57.80)
T <sub>4</sub>	<i>Bacillus thuringiensis</i> @ (3.5% ES) 2ml / l	16.67 (23.86)	26.37 (31.00)	71.48 (39.26)	85.92 (68.16)
T <sub>5</sub>	<i>Steinernema carpocapsae</i> @ (10,000 IJs) 4ml / l	0.00 (2.87)	10.00 (18.43)	39.26 (38.77)	46.29 (42.87)
T <sub>6</sub>	<i>Azadirachtin</i> @ (10,000 ppm) 2ml / l	10.00 (18.43)	13.33 (21.14)	46.30 (42.87)	58.52 (49.91)
T <sub>7</sub>	Untreated check	3.33 (8.85)	3.33 (8.06)	6.67 (13.25)	6.67 (13.25)
CD at 5%		-	4.83	9.03	9.32
SE (m) ±		3.45	3.39	2.97	3.07

\*Figures in parentheses are the arc sin transformed values.

### Bioefficacy of test bioagents against 1<sup>st</sup> instar larva

At 1 DAT, all the six test bioagents were found to be statistically non-significant with the untreated check exhibiting meager effect.

At 3 DAT, *Bt* (26.37%) was found to be significantly superior over rest of the treatments. *N. rileyi* (17.41%), Azadirachtin (13.33%) and *M. anisopliae* (13.33%) were observed to be the next best treatments which were on par. In the descending order of preference was *S. carpocapsae* (10%) followed by *B. bassiana* (6.67%).

At 5 DAT, *Bt* (71.48%) remained to be promising recording higher mortality over rest of treatments. *N. rileyi* (53.33%), Azadirachtin (46.30%), *M. anisopliae* (42.59%) and *S. carpocapsae* (39.26%) stood next best treatments which were on par with each other. *B. bassiana* (31.85%) was found to be least effective.

At 7 DAT, *Bt* (85.92%) was found the most promising treatment and in the order of merit next treatments were *N. rileyi* (71.48%), *M. anisopliae* (67.77%) and Azadirachtin (58.52%) which were on par. *S. carpocapsae* (46.29%) and *B. bassiana* (38.89%) recorded lower mortality and were at par with each other.

### Bioefficacy of test bioagents against 2<sup>nd</sup> instar larva

At 1 DAT, all the test bioagents were found to be non-significant with the untreated check exhibiting no harmful effect.

At 3 DAT, *B. thuringiensis* (20%) and *N. rileyi* (13.70%) were found to be the most effective treatments which were on par, followed by Azadirachtin (10%) and *M. anisopliae* (6.67%) which were on par followed by *S. carpocapsae* (3.33%) and *B. bassiana* (0.00%) and were at par with each other.

At 5 DAT, *Bt* (57.04%) remain to be significantly superior over rest of the treatment followed by *N. rileyi* (35.56%), Azadirachtin (27.41%), *M. anisopliae* (31.85%) and *S. carpocapsae* (24.81%) which were on par. Comparatively *B. bassiana* (17.41%) was observed to be the least effective treatment.

At 7 DAT, *Bt* (64.44%), *N. rileyi* (57.04%) and *M. anisopliae* (53.33%) were found superior over rest of the treatments which were at par with each other followed by Azadirachtin (46.30%) followed by *S. carpocapsae* (46.30%) and *B. bassiana* (24.81%) which were on par.

**Table 2:** Bioefficacy of test bioagents against third instar larvae

TN	Treatments	Per cent Mortality at Days After Treatment			
		1 DAT	3 DAT	5 DAT	7 DAT
T <sub>1</sub>	<i>Metarhizium anisopliae</i> @ (1 x 10 <sup>8</sup> cfu/ml) 5g / l	0.00 (2.87)*	6.67 (13.25)	31.85 (34.21)	53.33 (46.92)
T <sub>2</sub>	<i>Beauveria bassiana</i> @ (1 x 10 <sup>8</sup> cfu/ml) 5g / l	0.00 (2.87)	0.00 (2.87)	17.41 (24.05)	24.81 (29.82)
T <sub>3</sub>	<i>Nomuraea rileyi</i> @ (1 x 10 <sup>8</sup> cfu/ml) 5g / l	0.00 (2.87)	13.70 (21.49)	35.56 (36.59)	57.04 (49.05)
T <sub>4</sub>	<i>Bacillus thuringiensis</i> @ (3.5% ES) 2ml / l	10.00 (18.43)	20.00 (26.07)	57.04 (49.05)	64.44 (53.71)
T <sub>5</sub>	<i>Steinernema carpocapsae</i> @ (10,000 IJs) 4ml / l	0.00 (2.87)	3.33 (8.06)	24.81 (29.32)	31.85 (34.21)
T <sub>6</sub>	<i>Azadirachtin</i> @ (10,000 ppm) 2ml / l	6.67 (13.25)	10.00 (18.43)	27.41 (31.51)	46.30 (42.87)
T <sub>7</sub>	Untreated check	3.33 (8.85)	3.33 (8.06)	6.67 (13.25)	6.67 (13.25)
CD at 5%		-	5.47	9.29	8.19
SE(m)±		2.77	3.45	3.06	2.70

\*Figures in parentheses are the arc sin transformed values

**Bioefficacy of test bioagents against 5<sup>th</sup> instar larva**

At 1 DAT, all the test bioagents were found to be non-significant with the untreated check exhibiting no harmful effect.

At 3 DAT, *Bt* (10.37%) was found to be the most superior treatment followed by Azadirachtin (6.67%), *N. rileyi* (3.33%) and *M. anisopliae* (3.33%) which were on par followed by *S. carpocapsae* and *B. bassiana* which were at par with each other.

At 5 DAT, *Bt* (24.07%) was found most significantly superior

treatment. Next treatments in the order of merit were *N. rileyi* (20.74%), *M. anisopliae* (17.04%) and Azadirachtin (13.33%) which were found at par, followed by *S. carpocapsae* (6.67%) and *B. bassiana* (6.67%) which were on par with each other.

At 7 DAT, *Bt* (50.00%) and *N. rileyi* (36.68%) were observed to be the most promising treatments which were on par followed by *M. anisopliae* (30.00%), Azadirachtin (24.07%), *S. carpocapsae* (20.00%) and *B. bassiana* (17.04%) and were at par with each other.

**Table 3:** Bioefficacy of test bioagents against 5<sup>th</sup> instar larva

TN	Treatments	Per cent Mortality at Days After Treatment			
		1 DAT	3 DAT	5 DAT	7 DAT
T <sub>1</sub>	<i>Metarhizium anisopliae</i> @ (1 x 10 <sup>8</sup> cfu/ml) 5g / l	0.00 (2.87)*	6.67 (13.25)	17.04 (24.20)	30.00 (33.21)
T <sub>2</sub>	<i>Beauveria bassiana</i> @ (1 x 10 <sup>8</sup> cfu/ml) 5g / l	0.00 (2.87)	0.00 (2.87)	6.67 (13.25)	17.04 (24.20)
T <sub>3</sub>	<i>Nomuraea rileyi</i> @ (1 x 10 <sup>8</sup> cfu/ml) 5g / l	0.00 (2.87)	6.67 (13.25)	20.74 (27.08)	36.68 (37.22)
T <sub>4</sub>	<i>Bacillus thuringiensis</i> @ (3.5% ES) 2ml / l	6.67 (13.25)	10.37 (18.78)	24.07 (29.30)	50.00 (45.00)
T <sub>5</sub>	<i>Steinernema carpocapsae</i> @ (10,000 IJs) 4ml / l	0.00 (2.87)	3.33 (8.06)	6.67 (13.25)	20.00 (26.57)
T <sub>6</sub>	Azadirachtin @ (10,000 ppm) 2ml / l	3.33 (8.06)	3.33 (8.06)	13.33 (21.49)	24.07 (29.30)
T <sub>7</sub>	Untreated check	3.33 (8.85)	3.33 (8.06)	3.33 (8.06)	3.33 (8.06)
CD at 5%		-	5.04	7.58	10.51
SE(m)±		2.77	4.72	4.31	2.50

\*Figures in parentheses are the arc sin transformed values

**Conclusions**

Laboratory bioefficacy of six test bioagents were evaluated against 1<sup>st</sup>, 3<sup>rd</sup> and 5<sup>th</sup> instar larva. In first instar larva, the trend of bioefficacy of bioagents at 3 DAT, exhibited as *Bt*>*N. rileyi*, Azadirachtin and *M. anisopliae*>*S. carpocapsae*>*B. bassiana*. At 5 DAT, shown as *Bt*>*N. rileyi*, Azadirachtin, *M. anisopliae* and *S. carpocapsae*>*B. bassiana*. At 7 DAT, presented as *Bt*>*N. rileyi*, *M. anisopliae* and Azadirachtin>*S. carpocapsae* and *B. bassiana*. In third instar larva, the trend of bioefficacy of bioagents at 3 DAT, presented as *Bt* and *N. rileyi*>Azadirachtin and *M. anisopliae*>*S. carpocapsae* and *B. bassiana*. At 5 DAT, shown as *Bt*>*N. rileyi*, Azadirachtin, *M. anisopliae* and *S. carpocapsae*>*B. bassiana*. At 7 DAT, exhibited as *Bt*, *N. rileyi*, *M. anisopliae*>Azadirachtin >*S. carpocapsae* and *B. bassiana*. In fifth instar larva, the trend of bioefficacy of bioagents at 3 DAT, unveiled as *Bt*>Azadirachtin, *N. rileyi* and *M. anisopliae*>*S. carpocapsae* and *B. bassiana*. At 5 DAT, shown as *Bt*>*N. rileyi*, Azadirachtin, *M. anisopliae*>*S. carpocapsae* and *B. bassiana*. At 7 DAT, presented as *Bt* and *N. rileyi*>*M. anisopliae*, Azadirachtin, *S. carpocapsae* and *B. bassiana*.

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**References**

1. Asi MS, Bashir MH, Afzal M, Zia K. Potential of Entomopathogenic Fungi for Biocontrol of *Spodoptera litura* (Fabricius) (Lepidoptera: Noctuidae). J Animal & Pl. Sci 2013;23(3):913-918.
2. Caccia MG, Valle ED, Dauset ME. Susceptibility of *S. frugiperda*, *H. gelatopoeon* to EPN *Steinernema diaprepesi* under laboratory conditions. J Agril Res 2014;74(1):123-126.
3. Cruz-Avalos AM, Hernández MA, Ibarra JE, Rincón Castro. High virulence of Mexican entomopathogenic fungi against fall armyworm, (Lepidoptera: Noctuidae). J Econ Entomol 2019;112:99-107.
4. Domecino Pavone, Mayri Diaz, Lesbia Trujillo, Blas Dorta. A Granular formulation of *Nomuraea rileyi* Farlow (Samson) for the control of *Spodoptera frugiperda* (Lepidoptera: Noctuidae). Interciencia. 2014;34:0378-1844.
5. Lalitha C, Muralikrishna T, Sravani S, Devaki kalyan. Laboratory evaluation of native *Bacillus thuringiensis* isolates against *Spodoptera litura* (Fabricius). J Biopestic 2012;5(1):4-9.
6. Mallapur CP, Anjan KNS, Hagari T, Praveen. Potentiality of *Nomuraea rileyi* (Farlow) against the fall armyworm, *Spodoptera frugiperda* (J. E. Smith) infesting maize. J Ent & Zo Stud. 2018;6(6):1062-1067.
7. Malarvannan SPD, Murali, Shanthakumar SP, Prabavathy RV. Laboratory evaluation of entomopathogenic fungi, *Beauveria bassiana* against Tobacco caterpillar, *Spodoptera litura* (Fabricius) (Noctuidae : Lepidoptera). J Biopestic 2010;3(1):167-170.
8. Polanczyk RA, Rogerio F, Pires, silva, Lidia MF. Effectiveness of *Bacillus thuringiensis* strains against *Spodoptera frugiperda* (Lepidoptera: Noctuidae). Brazilian J Microbiology 2000;31:165-167.
9. Ramanujam B, Poornesha B, Shylesha AN. Effect of entomopathogenic fungi against invasive pest *Spodoptera*

- frugiperda* (J. E. Smith) (Lepidoptera : Noctuidae ) in maize. Egyptian J Biol Pest Control 2020;30:100-102.
10. Silva MS, Sania M, Forti B, Roseane C, Predes T, Emerson SF, Ismael BG. Toxicity and application of neem in fall armyworm. *Communicata Scientiae* 2015;6(3):359-364.
  11. Sisodiya DB, Nainesh Patel, Patel PH, Raghunandan BL, Gohel VR, Chavada KM. Bioefficacy of entomopathogenic fungi and bacteria against invasive pest *Spodoptera frugiperda* (J. E. Smith) under laboratory condition. *J Entomol & Zoo stud* 2020;8(6):716-720.
  12. Umamaheshwari R, Sivakumar M, Subramanaian S. Biocontrol efficacy of entomopathogenic nematodes on *Spodoptera litura* in black gram. *Indian, J Nematol* 2006;36(1):19-22.