

Journal of Entomology and Zoology Studies

J Journal of Entomology and Zoology Studies

Available online at www.entomoljournal.com

ISSN 2320-7078 JEZS 2014; 2 (2): 52-71 © 2014 JEZS Received: 17-02-2014 Accepted: 07-04-2014

Ghoneim K.

Faculty of Science,
Al-Azhar University,
Cairo, Egypt
Email: Kar Ghoneim@yahoo.com

Predatory insects and arachnids as potential biological control agents against the invasive tomato leafminer, *Tuta absoluta* Meyrick (Lepidoptera: Gelechiidae), in perspective and prospective

Ghoneim K.

ABSTRACT

Although the tomato leafminer *T. absoluta* is a native pest to South America, it invaded many countries in center and north Europe and North Africa. Moreover, it has become an economically important insect in the major tomato-producing countries because its larvae reduce tomato yield and cause great losses of the fruit quality. The use of predators, as biocontrol agents, is an important alternative to the chemical insecticides. The present article reviewed the record and feasible use of different predator species belonging to several insect orders and families, as well as some arachnid species. This review provided, also, an insight on the predator as potential biocontrol agents against *T. absoluta* in different parts of the world, especially the tomato producing countries. In addition, the integration between predators and some other biocontrol agents, and conservation of the indigenous natural predators in relation to *T. absoluta* had been discussed.

Keywords: Arachnida, Coleoptera, Dermaptera, Heteroptera, Hymenoptera, Neuroptera, Thysanoptera.

1. Introduction

The tomato leafminer, *Tuta absoluta* (Meyrick) (Lepidoptera: Gelechiidae), is an invasive pest, native to South America, where it is considered one of the most devastating pests of tomato *Solanum lycopersicum* L. (or *Lycopersicon esculentum* (Mill.) (Solanales: Solanaceae) in several parts of the world [1-3]. Although *T. absoluta* is a pest native to South America, the first detection of it in Europe was in late 2006 (Iberian Peninsula) [4, 5]. It has expanded very quickly in three years (2007-2009) to many countries in center and north Europe [6]. Moreover, it has become an economically important pest in the major tomatoproducing countries in the Mediterranean Basin countries of Europe and North Africa [2, 7-9]. Generally, it spreads very quickly in many countries of the western Palearctic Region [10]. More recently, the pest was reported in several Middle East countries [11].

This pest causes a very high level of damage (quantity and quality) to tomato crops [6, 12, 13], particularly if no control measures are adopted [11]. Larvae reduce tomato yield and fruit quality losses of up to 80-100% by attacking leaves and flowers, burrowing stalks, apical buds, green and ripe fruits [14-16]. Tomato plants may be attacked at any developmental stage from seedlings to mature plants in greenhouses and in open fields. Beside tomato, *T. absoluta* is also able to attack and cause damage on different genera and species of the Solanaceae plants [17]. As control strategies for *T. absoluta* consist mainly of early detection through sexual pheromone traps [18, 19], cultural methods also may be important [20]. The primary *T. absoluta* management strategy in most native home, South America, or invaded European and northern African countries, is chemical control [21, 22]. However, pesticides are only partially successful because of the general endophytic behaviour of the larval instars and the rapid selection of resistant populations [21, 22-24]. However, the resistance development in this pest to chemicals had been reported by several authors [15, 21- 23, 25, 26]. For implementing environmentally safe strategies, several eco-sustainable control methods and integrated pest management (IPM) programs have been recently evaluated [27-31].

Correspondence:
Ghoneim K.
Faculty of Science,
Al-Azhar University,
Cairo, Egypt
Email: Kar_Ghoneim@yahoo.com

Biological control (BC), within an integrated pest management program, is a challenging but potentially very beneficial tactic to develop [32]. BC programs have been used against crop pest insects belonging to the orders Homoptera, Diptera, Hymenoptera, Coleoptera and Lepidoptera, among others [33]. BC agents (living antagonists-natural enemies: predators, parasitoids and pathogens) are considered as one possible solution of the *T. absoluta* crisis ^{[2,} ^{34]}. This strategy offers a more sustainable and less expensive alternative to chemicals [35, 36]. As reported by several authors [37-39], the impact of opportunistic native natural enemies (fortuitous BC) on T. absoluta should be considered as one of the key factors for suppressing the population density of this pest. In his review, Urbaneja et al. [40] updated the available information on indigenous natural enemies in Mediterranean countries. In addition, natural enemies are able to learn. This ability allows them to improve their response to their host when they are reared sequential generations on a host [41].

Predators help to maintain a balance among organisms, by consuming prey, altering prey behavior and prey habitat selection [42]. Thus, predators may increase the biodiversity of communities by preventing a single species from becoming dominant [43]. Vertebrate predators, such as birds, lizards and small mammals, are not really introduced for the pest control but the insect species are known to have some effect on insect pests [41]. Unfortunately there is a problem in the use of predators since many of them are seen as generalists and therefore not seen as highly effective BC agents [44]. Although specialized natural enemies are considered as most promising in BC [45], the generalist predators may also be of major importance in pest suppression [46]. In addition, the generalist predators may be better suited than specialists in diverse environments indicating their potential in conservation biological control [47]. Although predators did not attract the attention of tomato growers in various South American countries infested with T. absoluta as the parasitoids, predators could be responsible for up to 79.4% larval mortality and egg predation amounts to 5% [37]. Therefore, the present review is primarily concerned with the reported works on predaceous insects and arachnids as natural enemies and potential biocontrol agents against T. absoluta allover the world. It is secondarily interested in some related aspects, such as side-effects of pesticides on these predators, integration between predators and some other BC agents, and conservation of indigenous natural predators.

2. Predacious insects against Tuta absoluta

The predacious insects, as natural enemies or BC agents, with special emphasis on *T. absoluta*, can be reviewed herein according to insect orders and families.

2.1. Heteroptera

Miridae is a large and diverse insect family in the order Heteroptera. They are globally important pests of crops such as alfalfa, apple, cocoa, cotton, sorghum, and tea. Some also are predators of crop pests and have been used successfully in BC ^[48, 49]. *Macrolophus* species is worldwide marketed since the 1990s as a BC agent. They are generalist zoophytophagous predatory bugs naturally present in Mediterranean area ^[50]. Examples include *Macrolophus caliginosus* (Wagner) and *Dicyphus hesperus* (Knight) that are generally available and widely used in protected crops in North America and Europe, respectively ^[51]. Other mirids have been integrated into conservation biological control programs

in horticultural crops in Europe and elsewhere, including Nesidiocoris (=Cyrtopeltis) tenuis (Reuter) [52, 53].

In tomato crops, the predators *Macrolophus pygmaeus* Rambur and *M. caliginosus* have been reported as a junior synonym of *Macrolophus melanotoma* (Costa) ^[54]. The mirid *M. caliginosus* (=*M. pygmaeus*) is a highly polyphagous predatory bug ^[55] which has proven to be effective in controlling many insect pests of greenhouse vegetables ^[56, 57]. This mirid predator was observed on or used to biologically control *T. absoluta* ^[16, 15, 55-69].

The mirid bugs of genus *Dicyphus* include zoophytophagous species, some of which are well-known for their role in the control of several pests of horticultural crops. In particular, *Dicyphus errans* (Wolff) is the species usually found in horticultural crops of northwestern Italy, but other *Dicyphus* species have been observed elsewhere in the crops of the North Mediterranean Basin ^[70]. This mirid predator was observed on or used as a biocontrol agent against *T. absoluta* ^[2, 71]. In addition, each of *Dicyphus tamaninii* Wagner ^[63, 72], *Dicyphus marrocannus* Wagner ^[2, 15, 59-61, 73, 74] and *D. hesperus* ^[72] were observed preying on eggs and young larvae of *T. absoluta* and can be used to control this pest.

The mirid bug *N. tenuis* is a polyphagous predator widely distributed in the Mediterranean region, where it has been used as an augmentative BC agent for several plant pests ^[75-77]. It can make significant contributions to the control of greenhouse pests such as whiteflies, leafminers, lepidopterans and spider mites ^[53, 78]. This predator effectively suppress populations of the tomato fruit borer *Helicoverpa armigera* (Lepidoptera: Noctuidae) ^[79]. The same predatory mirid was observed preying on eggs and young larvae of *T. absoluta* and can be used to control it ^[15, 16, 58-61, 68, 69, 80-87].

The mirid predator *Tupiocoris cucurbitaceus* (Spinola) attacks several crop and vegetable pests in different parts of the world. As for example, it is associated with whiteflies in tomato crops as a natural enemy ^[15, 88]. López et al. ^[89] evaluated some biological characteristics of this species. This mirid predator was observed on or used to control *T. absoluta* ^[90].

The mirid predator Heterotoma meriopterum (Scopoli) was found as an important factor regulating the filbert aphid, Myzocallis coryli (Homoptera: Aphididae), populations in the filbert orchards of Oregon (USA) [91]. It was also found as an aphidophagous predator in Belgium [92] and recorded among the predatory insects belonging to Heteroptera in different hazel groves in Piedmont (NW Italy) and in Sardinia Island [93]. The mirid predator Heterotoma sp. had been recorded on T. absoluta and can be used against this pest $^{[2,65]}$. The Anthocoridae are a family of bugs classified in order Heteroptera and commonly called 'minute pirate bugs' or 'flower bugs'. Pirate bugs prey on other small insects, spider mites and insect eggs. This family contains between 400 and 600 species distributed worldwide [94]. The genus *Orius* consists of omnivorous bugs feeding mostly on spider mites, thrips, Bemisia tabaci (Homoptera: Aleyrodidae) and their eggs, but will also feed on pollen and vascular sap [95]. The omnivorous predator Orius insidiosus (Say) is considered beneficial, as it feeds on small arthropod pests and their eggs. This predatory bug was recorded on T. absoluta and can be as a biocobnrol agent against it [2, 15, 96-98]. Another anthocorid predator, Orius laevigatus (Fieber), is an economically important BC agent of several small arthropod pests [99]. It was recorded on or used to control *T. absoluta* [87]. The predator Orius albidipennis was recorded from several parts in Europe, Africa and Asia [96, 100-103]. This predatory bug is used for BC of T. absoluta [84,100]. The anthocorid predacious bug Xylocoris flavipes (Reuter) appears to be a promising agent for population suppression of both Coleoptera and Lepidoptera in stored products, since they prey on most stages of many of these species $^{[104]}$. The *Xylocoris* sp. was recorded on or used as an egg predator for *T. absoluta* $^{[15,37]}$.

The **Berytidae** (stilt bugs) represent a small family of true bugs in order Heteroptera. Most of them are phytophagous. Some species are occasionally omnivorous and facultative carnivorous or saprophagous ^[105]. The stilt bug *Metacanthus tenellus* Stål is a predator with vary species of pests as preys in the tomato crop such as eggs and larvae of *T. absoluta* ^[106].

Nabidae (order Heteroptera) prey on a variety of small invertebrates, chiefly arthropods. The predaceous habit, together with the widespread occurrence of some species in a variety of ecosystems, particularly agro-ecosystems, has attracted the attention of entomologists [107, 108]. Multi-stage dynamic model for a prey-predator interaction was applied to the damsel bug *Nabis pseudoferus* Remane [109]. It has been described as an effective predator against aphids and lepidopteran eggs and larvae, although it can feed also on other preys. Preliminary results identified it as a candidate for BC of *T. absoluta* by preying on larvae [2, 15, 28, 110]. In addition, Cabello et al. [111] demonstrated that *N. pseudoferus* will feed on *T. absoluta* eggs.

The **Lygaeidae** (order Heteroptera) are known as true bugs. The family includes the insects commonly known as milkweed bugs, and also some of those known as seed bugs. The family includes about 60 genera in six subfamilies (for some details, see ^[112]). Bell and Whitcomb ^[113] reported that the predatory big-eyed seed bug *Geocoris punctipes* Say was found among the most abundant and important predators of bollworm eggs, aphids, plant bugs, and young larvae in Arkansas cotton fields (USA). Orhanides *et al.* ^[114] reported that *G. punctipes* was an effective predator of the pink bollworm, *Pectinophora gossypiella* (Lepidoptera; Gelechiidae), in southern California cotton fields (USA). Some biological and ecological aspects of *G. punctipes* had been studied ^[115, 116]. *G. punctipes* was reported as biocontrol agent against *T. absoluta* ^[98]. **Pentatomidae** are a family of insects (order Heteroptera) including

some of the stink bugs and shield bugs. The majority are herbivorous and some are predaceous on other insects. Many species, whether primarily herbivorous or predaceous, are generalist feeders [117]. Biology and ecology of the pentatomid stink bug *Podisus nigrispinus* (Dallas) and its use to control defoliating lepidopterous larvae in *Eucalyptus* plantations in Brazil were discussed [118]. Other Biological and behavioural studies had been conducted by some authors [119, 120]. *P. nigrispinus* was recorded as a BC agent for *T. absoluta* [15, 29, 35, 37, 80, 81, 121-123].

Reduviidae is a large, cosmopolitan family of predatory insects. It includes assassin bugs, ambush bugs, wheel bugs and threadlegged bugs. There are about 7000 species altogether, making it one of the largest families in the Heteroptera. Assassin bugs are essential components of ecosystems, but also important in agriculture and medicine (for some details, see [124-126]). *Debilia* sp. is believed to be a potential predator of adult leaf-footed bug *Leptoglossus* sp. (Heteroptera: Coreidae) [127]. This reduviid predator was observed as a natural predator of *T. absoluta* [2, 15].

2.2. Hymenoptera:

Vespidae constitutes a large family (nearly 5,000 species) in the order Hymenoptera. It is considered a diverse and cosmopolitan family of wasps, including nearly all the known eusocial wasps and many solitary wasps. Many species are pollen vectors contributing to the pollination of several plants, being potential or even effective

pollinators, while others are notable predators of insect pest species ^[128]. Predatory Vespidae are the most voracious predators which can harvest very large numbers of invertebrate preys ^[129]. Also, the social wasps are generalist predators and are important agents for natural BC of phytophagous insects on natural environments and agroecosystems ^[37, 130-134]. Predatory wasps are well adapted to control lepidopteran larvae in high population densities ^[135]. Predation behaviour of *T. absoluta* by vespid wasps had been described by Picanço *et al.* ^[136]. There is a relatively large body of literature on certain species of predaceous vespid wasps, in the genera *Polistes, Polybia, Brachygastra, Protonectarina* and *Synoeca*, as well as their promising role in the BC against several insect pests including *T. absoluta*.

The paper wasps Polistes are classified in a cosmopolitan genus. It is also the single largest genus within the family Vespidae, with over 300 recognized species and subspecies. Paper wasps feed on nectar, and insects, including lepidopterous caterpillars, flies, and beetle larvae [137]. Specialization on caterpillar preys makes *Polistes* suitable for augmentative BC of lepidopterous pests of field crops [138, 139]. With regard to the paper wasp *Polistes versicolor* (Olivier), it was reported as a natural enemy of the cassava hornworm (Erinnyis ello) (Lepidoptera: Sphingidae) [140] and the coffee-leafminer Leucoptera coffeella (Lepidoptera: Lyonetiidae) [141]. In a study, Prezoto et al. [132] found that 95% of prey caught by P. versicolor was lepidopteran larvae. Moreover, it is one of social wasps preying on larval forms of coleopterous and dipterous pests of agricultural and forest crops [142-144]. This paper wasp was recorded as predator on T. absoluta [2, 15]. In addition, the vespid wasps Polistes melanosoma (de Saussure) and Polistes carnifex (Fabricius) were recorded as biocontrol agents for *T. absoluta* ^[2, 15]. Another paper wasp, Polybia scutellaris (White), is a eusocial wasp in South America [145]. It hunts and stores a large number of prey insects from different orders in their nests [145]. It is one of predators of the coffee-leaf-miner L. coffeella found in Brazil [141]. Its role as a pollinator is interesting and its role as biocontrol agent for agricultural pests is possible [146]. P. scutellaris was recorded as a predator on *T. absoluta* [15, 136]. In addition, *Polybia* fastidiosuscula Saussure, Polybia (=Protopolybia) exigua (De Saussure) and Polybia ignobilis (Haliday) had been recorded as predators of *T. absoluta* and can be used to control this pest [2, 15, 136,

The dark paper wasp *Brachygastra lecheguana* (Latreille) is one of predators of *L. coffeella* found in Brazil ^[141]. Adults of *B. lecheguana* had been exposed to some insecticides in order to study its affected efficiency ^[147, 148]. The vespid wasp *B. lecheguana* was recorded preying on *T. absoluta* ^[15, 136, 148].

Morphological and physiological variation between queens and workers of the predacious social wasp *Protonectarina sylveirae* (Saussure) had been investigated ^[149]. The selectivity of 8 insecticides to *P. sylveirae*, among other vespid predators ^[147] and the toxicity of new pyrethroids ^[150] had been evaluated. This wasp was recorded as natural predator on *T. absoluta* ^[15, 136, 147, 148, 151].

As concluded by Prezoto and Braga ^[152], the social wasp *Synoeca cyanea* (Fabr.) has broad distribution throughout Brazil and had been considered as an interesting natural enemy of larvae of *Zaprinus indianus* (Diptera: Drosophilidae) as a dipteran pests of fruit crops. This wasp was recorded as a natural predator on *T. absoluta* in different parts of the world ^[2, 15, 136].

The **Formicidae** (family of ants) are social insects in order Hymenoptera. More than 12,500, out of an estimated total of 22,000 species, have been classified (for more details, see [153-155]).

Bownes [156] conducted a study on the use of ants for controlling crop pests and several studies have demonstrated that ants can be key generalist predators of crop pests in different parts of the world [157-162]

The tropical fire ant *Solenopsis geminata* (Fabricius) (=*Atta geminata* Fabricius) is native to the south-eastern coastal plain of Florida to Texas south through Central America to northern South America ^[163]. It is highly omnivorous feeder ^[164]. The predation of this fire ant on eggs of the apple snail *Pomacea canaliculata* (Gastropoda: Ampullaridae) was studied ^[165]. Way et al. ^[166] discussed the role of *S. geminate* in the BC of tropical upland rice pests. The same fire ant was observed as a pupal predator on *T. absoluta* ^[15, 167]. Another fire ant, *Solenopsis saevissima* (Smith), was recorded as potentially important predator of the sugarcane borer, *Diatraea saccharalis* (Lepidoptera: Crambidae), in sugarcane crops in Brazil ^[168]. The latter fire ant was recorded as a beneficial predator for control *T. absoluta* ^[2, 15, 169].

The ant *Pheidole megacephala* (Fabricius) is used as BC agent against the sweet-potato weevil *Cylas formicarius* (Coleoptera: Brentidae) [170, 171]. Larvae of the rice leaf-folder *Cnaphalocrocis medinalis* (Lepidoptera: Pyralidae) were found attacked by adults of the predatory ant *Pheidole* sp. [172]. *Pheidole* sp. was also recorded as a natural predator for the diamondback moth *Plutella xylostella* (Lepidoptera: Plutellidae) in Brazil [173] and for the sugarcane borer *D. saccharalis* populations [174, 175]. The same ant was reported as pupal natural predator suppressing the populations of *T. absoluta* [15, 167].

2.3. Dermaptera:

Family **Labiduridae**, whose members are known commonly as striped earwigs, is a relatively large family of earwigs in the suborder Forficulinae (order: Dermaptera). The family is mostly cosmopolitan (for some details, see [176, 177]). A number of laboratory studies, focusing on specific orchard pests, demonstrated earwig pest consumption ability of scale insects [177], aphids, [178], spider mites [179], and psyllids [180]. The earwigs are considered voracious predators because they have high ability to attack and feed on different preys, particularly eggs and immature stages of insects of the orders Lepidoptera, Hemiptera, Coleoptera and Diptera [181]. In contrast, some labidurids are considered as plant pests [182, 183].

The Indian earwig (or the tawny earwig) *Labidura riparia* Pallas is polyphagous predator [184]. It mainly preys on cutworms, caterpillars, small grubs, maggots, mealybugs, fleas and other insects [184]. In Brazil, this earwig was associated with commercial poultry ranches as BC agents [185]. In USA, it was recorded as a predator of pupal stage of lepidopterous pests in soybean fields [186], and was the most abundant predator in peanut fields [187]. In Germany, it is a predator of tomato pests [167]. In Egypt, it was recorded affecting the population densities of (Thysanoptera: Thripidae), Empoasca sp. Thrips tabaci (Hemiptera: Cicadellidae) and Compylomma sp. (Hemiptera: Miridae) in corn fields [188]; B. tabaci in cotton fields [189]; Phthorimaea operculella (Lepidoptera: Gelechiidae) in tomato and potato fields [190] and Aphis craccivora (Homoptera: Aphididae) in faba bean fields [191]. Some physiological aspects of L. riparia, such as consumption of prey and nature of carnivory, had been studied [192]. This Indian earwig was reported as a pupal predator for T. absoluta [15, 167].

Forficulidae is a family of earwigs, in the suborder Forficulinae (order: Dermaptera). It is the largest family of earwigs, with 250

species worldwide (for details, see [176, 193, 194]. The characterization of earwig, *Doru lineare* (Eschscholtz), as a predator of larvae of the fall army worm, *Spodoptera frugiperda* (Lepidoptera: Noctuidae), was studied [195, 196]. Zotti *et al.* [197] evaluated the survival of *D. lineare* adults after ingesting eggs of *S. frugiperda* contaminated with some insecticides. This earwig was recorded as a predator for controlling *T. absoluta* [2, 15].

2.4. Coleoptera:

Family **Coccinellidae** (order: Coleoptera) is a well-known beetle family, worldwide distributed and divided into six subfamilies ^[198]. Except for the mycophagous Coccinellinae and the phytophagous Epilachninae, all remaining coccinellids are predators of hemipteran insects from the suborder Sternorrhyncha (e.g. aphids, scales, psyllids and whiteflies), mites and eventually other insect larvae (for detail, see ^[199, 200]). Greater emphasis is needed on evaluation, predator specificity, understanding colonization of new environments, and assessment of community-level interactions to maximize the use of coccinellids in BC ^[201].

Some coccinellid species attracted the attention of BC specialists, particularly against T. absoluta. Herein, a number of the promising biocontrol coccinellids has been reviewed. Cardoso and Lázzar [202] examined the feasibility of use of the spotless ladybird Cycloneda sanguinea (Linnaeus) for the BC of the giant conifer aphids. C. sanguinea is a voracious predator of pests such as aphids, mealybugs, and eggs of Lepidoptera [203]. Nakasu et al. [204] reported C. sanguinea as one of the main predators of non-target pests in Brazilian cotton. This coccinellid predator was recorded as an egg predator for *T. absoluta* [15, 37, 205-207]. Another coccinellid, Chilocorus nigritus (Fabricius), is an important natural enemy against red scale Aonidiella aurantii (Hemiptera: Diaspididae) on citrus in southern Africa [208]. This coccinellid has been widely used for the BC of some pests such as the armored scale insects [209, 210]. The coccinellid predator Chilocorus sp. was observed attacking or used for controlling T. absoluta [211].

The spotted lady beetle Coleomegilla maculata De Geer is a natural enemy of several insect pests. It feeds on pollen and nectar to survive periods when prey is scarce [212]. C. maculata was found as among the most common predators for the soybean aphid, Aphis glycines (Homoptera: Aphididae) in North America [213]. Silva et al. [214] evaluated the development of C. maculata larvae fed on eggs of the Mediterranean flour moth Ephestia (=Anagasta) kuehniella (Lepidoptera: Pyralidae), eggs of S. frugiperda and nymphs of Schizaphis graminum (Hemiptera: Aphididae). Also, larvae of this lady beetle were reared on eggs of E. kuehniella in the laboratory studies [212]. This coccinellid predator had been observed preving on T. absoluta and may be used as a biocontrol agent against this pest [2, 15]. Considering the coccinellid predator *Eriopsis conexa* Germar, it was observed on Artemisia absinthium (Asteraceae) and Foeniculum vulgare (Apiaceae) plants, feeding on aphids [215]. It was reported as a natural predator on the aphids Aphis sp. and Myzus persicae (Homoptera: Aphididae) infesting Papaya tree in Brazil [160]. This coccinellid predator had been observed preying on $T.\ absoluta^{\ [2,\ 15]}.$

Carabidae is a large, cosmopolitan family of beetles (order: Coleoptera). They are commonly known as ground beetles including more than 40,000 species worldwide ^[216]. Carabids are generalist predators in crops and in natural habitats ^[217]. They are cited as being predators of aphids, lepidopteran larvae, slugs, and herbaceous plant seeds ^[218]. Also, many species of these beetles have a role in the natural BC for several lepidopteran pests in

different crops ^[219, 220]. The ground beetle *Calosoma granulatum* (=*Calosoma alternans granulatum*) Perty was recorded as a predator of cotton leafworm *Anticarsia gemmatalis* (Lepidoptera: Noctuidae) in Bolivia ^[221]. Pegoraro and Foerster ^[222] evaluated the abundance and distribution of this predator and the determining conditions in Brazil. Larvae and adults of this predator may prey on relatively high numbers of the cotton leafworm *Alabama argillacea* (Lepidoptera: Noctuidae) pupae ^[219]. This ground beetle was observed as natural predator of *T. absoluta* ^[2, 15]. In addition, the carabids *Lebia concina* and *Selenophorus* sp. were observed as predators of *T. absoluta* ^[2, 15].

2.5. Neuroptera:

Chrysopidae (order: Neuroptera) is a large widespread family of insects comprising about 1,300-2,000 species in 85 genera. They are known as green lacewings. They feed on pollen, nectar and honeydew supplemented with mites, aphids and other small arthropods, and some, namely Chrysopa, are mainly predatory [223]. Therefore, their voracious predacious larvae are colloquially known as "aphid lions" or "aphid wolves". In several countries, millions of voracious Chrysopidae are reared for sale as BC agents of insect and mite pests in agriculture and gardens [224]. The predacious green lacewing Chrysoperla externa (Hagen) has a number of traits that make it amenable to mass-rearing. It. feeds on eggs and young larvae, and so can be utilized in BC programs [225-^{227]}. The influence of adult food on the reproductive potential ^[228], the development [229, 230] and consumption capacity of *C. externa* on eggs and larvae of *S. frugiperda* and eggs of *E. kuehniella* [202] had been evaluated [231]. This lacewing was proved to be a good predator of T. absoluta larvae under laboratory conditions [15, 232]. The role of another Chrysopid species, Chrysopa carnea (Stephens), as a predator of the green peach aphid Myzus persicae (Homoptera: Aphididae) was demonstrated [233]. In Iraq, the biology of the predator Chrysopa mutata, fed either on the aphid Hyalopterus pruni (Homoptera: Aphididae) or the pseudococcid Nipaecoccus vastator (=Nipaecoccus viridis)(Homoptera: Coccidae) was studied in the laboratory [234]. The relationship between the predator Chrysopa carnea and preys (Aphis faba, Aphis gossypii and Myzus persicae, Homoptera: Aphididae) was studied in Egypt [235]. The third instar larvae of C. carnea have considerable potential for the control of *Trialeurodes vaporariorum* (Homoptera: Aleyrodidae) [236]. The chrysopid predator, Chrysopa sp., was observed preying on T. absoluta [2, 15].

2.6. Thysanoptera:

Thrips (Order Thysanoptera) are tiny, slender insects with fringed wings. Thrips species feed on a large variety of plants and animals by puncturing them and sucking up the contents. A large number of thrips species are considered pests, because they feed on plants with commercial value. Some species of thrips feed on other insects or mites and are considered beneficial, while some feed on fungal spores or pollen. So far around 5,000 species have been described (for some details, see [237, 238]).

Thripidae (order: Thysanoptera) is a family of thrips with over 290 genera representing just over two thousand species. Several species are economically significant pests, some of them invasive ^[237, 239, 240]. The bionomics of *Scolothrips sexmaculatus* (Pergande), an insect predator of spider mites, was studied ^[241]. The predation efficiency of this predator on the two-spotted spider mite *Tetranychus urticae* Koch (Acari: Trombidiformes: Tetranychidae) was described ^[242]. The same thrips species was observed preying

on T. absoluta [2, 15].

The Phlaeothripidae is the largest family in order Thysanoptera, with about 3550 species in 460 genera (for systematic details, see [243, 244]). A large number of species are considered pests, because they feed on plants with commercial value while some species feed on other insects or mites and are considered beneficial. Many species are fungivores living in the tropics and feed on fungal spores or pollen [237, 238, 245, 246]. Majority of species in the genus Haplothrips are European, breeding almost exclusively in flowers and many being host-specific. In contrast to the phytophagy of most members of the genus, the thrips Haplothrips subtilissimus (Haliday) is predator on other small arthropods [247]. Some Haplothrips species are known to prey on thrips [248]. For example, Haplothrips victoriensis Bagnall has been reported to be a natural enemy for the spider mite T. urticae [249]. In a mulberry field in Kagoshima, Japan, Haplothrips brevitubus (Karny) adults and larvae had been observed preying on the mulberry thrips, Pseudodendrothrips mori (Thysanoptera: Thripidae) [250]. However, some thrips species of the family Phlaeothripidae were observed preving on T. absoluta [37].

The **Aeolothripidae**, as a family of thrips, are particularly common in the holarctic region, although several occur in the drier parts of the subtropics. While they normally prey on other arthropods, many feed also on flowers [251]. *Franklinothrips* is a pantropical genus of ant-mimicking obligate predators [252]. Various members of the genus are considered to be, and are sometimes marketed as, useful BC agents against pest thrips [253]. *Franklinothrips vespiformis* Crawford has been marketed in Europe as a control agent against pests, such as the green-house thrips, *Heliothrips haemorrhoidalis* [254, 255]. It also feeds on mites, nymbs of a whitefly species and the larvae of an agromyzid fly [256]. Pizzol *et al.* [257] discussed the use of the predator *F. vespiformis* in an integrated pest management program. The same predator thrips was observed preving on *T. absoluta* [15].

3. Predacious arachnids against *T. absoluta* 3.1. Mites

Predatory mites in the family Phytoseiidae (Arachnida: Acari: Mesostigmata) have been successfully used in the BC of numerous agricultural pests worldwide. Many species of the genus Amblyseius are predatory mites feeding on other mites such as red spider mites, and also on thrips. Several species are popular as BC agents to control these pests [258]. In this regard, a population of the phytoseiid mite Amblyseius swirskii (Athias-Henriot) has been identified as a potential predator of the broad mite, Polyphagotarsonemus latus (Acari: Trombidiformes: Tarsonemidae), and is now a candidate for release against this pest on vegetable farms in Benin (Africa) [259, 260]. This mite was observed preying on eggs of *T. absoluta* in the Mediterranean Basin countries ^[15, 62, 74]. Wittmann and Leather ^[261] recorded the mite Amblyseius cucumeris Oudemans (= Neoseiulus cucumeris) as predator of the thrips Frankliniella occidentalis. Also, several studies had been carried out for use this predatory mite to control some species of thrips [262, 263]. Commercially available strains of live predatory mite A. cucumeris were investigated for predating on spider mite eggs and grain mite eggs [264, 265]. This predatory mite was observed attacking T. absoluta in the Mediterranean Basin countries [2,74].

3.2. Spiders.

Conley [266] studied the predation versus resource limitation in

survival of adult burrowing wolf spider *Geolycosa rafaelana* (Araneae: Lycosidae). Different studies had been conducted for the predation of wolf spiders on some insect pests such as three grassland grasshoppers in Nebraska (USA) ^[267]. Monzo *et al.* ^[268] revealed the potential use of DNA markers to track medfly predation by the wolf spider, *Pardosa cribata* Simon in citrus orchards in eastern Spain. Some wolf spiders had been recorded as pupal predators on *T. absoluta* ^[167]. Also, the wolf spider predator *Tanimlanmamis* sp. (Araneae: Lycosidae) was observed attacking *T. absoluta* in Turkey ^[15].

4. An insight for predators as promising biocontrol agents against *T. absoluta* in different parts of the world.

Taking into consideration the predators belonging to insect orders and arachnids, as natural enemies and biocontrol agents, especially against *T. absoluta*, it should be of great importance to compile some of the recently reported efforts in different parts of the world categorized in the most infested parts, the Mediterranean and European countries and South American countries.

4.1. Mediterranean and European countries.

In general, the mirid omnivorous predators *M. pygmaeus*, *N. tenuis* and *D. tamaninii* are considered to be among the principal natural enemies of pests on tomato crops in the Mediterranean region ^[2, 59, 73, 60, 61, 72, 74]. The naturally occurring populations of the mirid predators *Macrolophus* sp. and *Dicyphus* sp. can colonize tomato crops and keep the pest population below the damage threshold if chemical spreyings are reduced ^[269]. The mirid species *M. pygmaeus* is naturally present in Mediterranean area and used to control the populations of *T. absoluta* in protected tomato crops ^[270]. Beside it, the mirid predator *N. tenuis* can regulate *T. absoluta* populations, because it is able to prey efficiently on *T. absoluta* eggs ^[29]. However, the biocontrol of *T. absoluta* by predators in some important tomato-producing countries can be reviewed herein.

In Spain, M. pygmaeus and N. tenuis have already been tested under laboratory conditions to assess their suitability as T. absoluta egg and larval predators [61, 73]. According to these authors, in the tomato fields of Spain where the natural population of their predators were well established, the damage by T. absoluta was significantly lower than in the chemically treated fields. Field data indicated that pest incidence was lower when BC strategies based on the use of these two polyphagous mirid predators were applied instead of conventional chemical control [65, 271]. Molla et al. [60] showed that when M. pygmaeus and N. tenuis were established in tomato crop, they were able to reduce up to 75%-97% of leaflet infestations or 56% -100% of fruit infestations, respectively. In his study, Mollá [16] concluded that N. tenuis is a good BC against T. absoluta in tomato crops and its polyphagous behavior allows it to contribute in the control of other pests. It is possible to establish *N*. tenuis at the starting of the crop season and together a better and efficient control of *T. absoluta*, in tomato crops [272, 273]. A predator of the family Nabidae, N. pseudoferus, is also a native natural enemy usually found on tomato crops in Spain. An experimental work showed that its young nymphs can significantly reduce the numbers of T. absoluta eggs, indicating its potential as a promising candidate for the BC of this pest [110].

In **Italy** (Sardinia), Nannini *et al.* ^[274] evaluated the efficacy of releasing of the predators *M. pygmaeus* or *N. tenuis* per m² for the control of *T. absoluta* infestations, under field conditions. Unfortunately, both predators failed to achieve the levels necessary

for effective pest control due to the application of pesticides. The generalist mirid predator *D. errans* is usually used in the IPM tomato crop of northwestern Italy in tomato infested with *T. absoluta*. This mirid bug preyed mainly on eggs and first instar larvae ^[275]. In tomato plantations infested by *T. absoluta* during the period 2009-2010 in Piedmont, *D. errans* was largely found among nine species of indigenous natural enemies. It revealed to be an effective biocontrol agent preying mainly on eggs and first-instar larvae of *T. absoluta* ^[276]. This mired predator can be considered a promising potential candidate for controlling this pest, and further research is needed to assess its effectiveness under field conditions ^[71]

In **United Kingdom**, the results of BC measures for *T. absoluta* obtained by Morley et al. ^[62] can be summarized as follows. The mirid predator *N. tenuis* is a voracious predator feeding on eggs and larvae. Another mirid predator, *M. pygmaeus*, is closely related to *N. tenuis*. They feed on immature stages of *T. absoluta* and can provide some control of this pest in the absence of harmful insecticide residues. Also, the related species, *M. caliginosus*, has become established on many of UK tomato nurseries, and currently offers the best option for BC. However, according to a report from the UK, *N. tenuis* is problematic because it can attack host plants when prey are in short supply ^[277, 278].

A survey of natural enemies attacking *T. absoluta* was achieved by Biondi et al. ^[69] in Southern **France** during seasons 2011 and 2012. Three species of mirid predators, *M. pygmaeus*, *N. tenuis* and *Dicyphus* sp., were found preying on eggs and young larvae of the pest in both seasons. The obtained results represented the first step towards developing biological and IPM strategies against this pest in France.

In The **Netherlands**, some predators were found on the outdoor tomato plants. The mirid predator *D. errans* occurred regularly during summer and the mirid predator *Heterotoma* sp. was observed occasionally. Dutch growers will continue with the introduction of the predator *M. pygmaeus*, but they are always interested in new developments in BC of *T. absoluta* ^[2].

In **Turkey**, Öztemiz ^[15] reviewed several species of parasitoids and predators as BC agents against *T. absoluta*. He discussed the role of *N. tenuis* and *M. pygmaeus* in BC of this pest.

In **Jordan**, Al-Jboory *et al.* ^[84] conducted biweekly field visits (January-April 2011) to tomato fields (open and plastic houses) heavily infested with *T. absoluta*, in order to survey natural enemies associated with this pest. They recorded the occurrence of the Anthocorid predator *O. albidipennis* and mirid predator *N. tenuis*

In North Africa, Boualem et al. [279] focused on biological study of T. absoluta and the natural mirid predator N. tenuis in natural conditions and in the laboratory in Mostaganem area (Algeria). In the vicinity of this area, Guenaoui et al. [63] recorded some natural enemies such as the mirids N. tenuis, M. caliginosus and D. tamaninii whose predation efficiency had been evaluated under the laboratory conditions. At the next year, Boualem et al. [66] revealed the presence of three mirid predatory species of T. absoluta: N. tenuis, M. pygmaeus (caliginosus) and D. errans. These results had been substantiated because a very recent research work explored the possibilities of the native antagonists, such as two mirid predators: N. tenuis and M. caliginosus, for the control of T. absoluta [68]. With regard to Tunisia, many attempts have been made to biologically control *T. absoluta* using the mirid predator *N*. tenuis in nurseries, greenhouses and open field tomato crops. Abbes et al. [83] carried out few experiments in BC, using the mirid predator *N. tenuis*. The factors of failure of this predator in a protected tomato crop were analyzed by Abbes and Chermiti ^[280]. Recently, Hamdi *et al.* ^[50] suggested a general schema to follow for a successful introduction of a natural enemy into an agroecosystem. They defined this schema based on the example of using the mirid predator *Macrolophus* sp. to control *T. absoluta*. They characterized the predation potential of *M. pygmaeus* under different temperatures to test its effectiveness and its adaptability to the climatic conditions of an introduction area in Tunisia. In **Egypt**, *N. tenuis* was recorded for the first time as associated with *T. absoluta* in tomato plantations in Giza, Qaluobia and Fayoum Governorates. Tabulated data were given on the predator sample collection from different localities in Egypt from May 2011 to October 2012 ^[77].

In connection with arachnids attacking *T. absoluta* or being used to control it in Mediterranean countries, *A. swirskii* and *A. cucumeris* are considered promising egg predators ^[2]. As summarized results of BC measures for *T. absoluta* in UK ^[62], the predatory mite, *A. swirskii*, has been seen feeding on the first instar larvae of *T. absoluta*.

4.2. South American countries.

Although a broad diversity of predators has been found in association with T. absoluta in tomato fields in various South American countries, some predator species have received attention as BC agents for this pest. In Brazil, anthocorid predator Xylocoris sp., C. sanguinea and members of Phlaeothripidae proved to be key predators of both egg and larval T. absoluta stages [37]. As pointed out by several authors [205-207], the occurrence of anthicid predator Anthicus sp., coccinellid predator C. sanguinea, Staphylinidae, Orius sp. and Xylocoris sp. and Formicidae were observed on T. absoluta. The predaceous wasp P. sylveirae is considered as one of key mortality factors of T. absoluta in the spring-summer [58] and caused 29.3-37.4% T. absoluta larval mortality [151]. After a few years, Picanço *et al.* [136] evaluated the importance of social vespid wasps as predators of T. absoluta larvae. They recorded the social vespid predators: B. lecheguana, P. exigua, P. ignobilis, P. scutellaris, P. sylveirae, Polybia fastidiosuscula and S. cyanea. The vespid predators P. scutellaris and P. sylveirae were the most abundant ones. The predatory pentatomid stinkbug P. nigrispinus has received vast scientific attention, with baseline research done on its life history (on *T. absoluta*) [35], its predatory behaviour [122], and its dispersal in the greenhouse environments [80]. The results suggested that P. nigrispinus is able to maintain its population preying only on T. absoluta caterpillars; however, the life table parameters determined individually showed that the pest produces more generations per year and faster population natural growth than the predator. It has the ability to survive on alternative prey, enabling it to maintain a presence in the tomato ecosystem until a primary pest species arrives [35]. In Brazil, also, the green lacewing, C. externa, proved to be a good predator of T. absoluta larvae under laboratory conditions [232]. Lins et al. [97] determined the predation capacity of O. insidiosus on eggs of T. absoluta under laboratory conditions. Bueno et al. [98] evaluated the predation efficacy of two hemipteran predators, G. punctipes and O. insidiosus as biocontrol agents against T. absoluta. Probst et al. [167] hint the importance of pupal predators of T. absoluta such as the labidurid L. riparia, the formicid S. geminata, Pheidole sp. and ground beetles. In Venezuela, the anthocorid, O. insidiosus was reported to be an important predator of T. absoluta eggs and larvae [281]. In Chile, Vargas [282] indicated that the nabid predator Nabis

sp. and the vespid predator *Polistes* sp. are predators of *T. absoluta* in tomato fields. In **Argentina**, larvae of *T. absoluta* were being preyed on by a species of the coccinellid *Chilocorus* sp. ^[211]. The mirid predators *M. pygmaeus* and *N. tenuis* are attacking *T. absoluta*. An additional mirid, *T. cucurbitaceus*, has been recently evaluated as a potential BC agent against *T. absoluta* in Argentina ^[90]

With regard to arachnids attacking *T. absoluta* or being used to control it in some South American countries, Vargas ^[282] reported that spiders are predators of *T. absoluta* in tomato fields in Chile. In Brazil, Miranda et al. ^[37] indicated the presence of Araneidae attacking *T. absoluta*. Oliveira et al. ^[123] indicated that the mite *Pyemotes* sp. feeds on *T. absoluta* larvae, pupae and adults. They described its potential use in BC of the pest. Some wolf spiders were recorded among the important predators attacking *T. absoluta* ^[167]

5. Side-effects of pesticides on predators of *T. absoluta*:

Many indigenous species of native predators are successfully used within IPM programs resulting in high levels of control efficacy against T. absoluta [61]. Unfortunately, occurrence of this pest at increasing population levels led growers to extensively use insecticides, which could cause many side-effects on natural enemies in tomato crops [283-286]. Despite the importance of selective insecticide in preserving the natural BC of pests, little is known about insecticide selectivity in reference to predatory wasps [287]. Although some of the available insecticides used to control T. absoluta are not highly toxic to these predators [40, 288], evaluation of the side-effects of insecticides, widely used to control T. absoluta, attracted the increasing attention of numerous entomologists in different parts of the world. As examples, sideeffects of some insecticides had been assessed on ground beetle C. granulatum [289]. Also, comparative side-effects of some synthetic insecticides and bioinsecticides had been evaluated on the green lacewing C. externa [290-292].

Several worldwide efforts have been reviewed herein aiming to increase the enthusiasm for selecting safer or selective insecticides for the indigenous, or even introduced, predators. Two sulphur formulations, (dustable and wettable powder) are used in Mediterranean basin countries for controlling T. absoluta on tomato under greenhouse and open-field conditions, although its side-effects on N. tenuis should be taken into account [293]. In Spain, Arnó and Gabarra [5] evaluated the lethal and sublethal side-effects of three insecticides most widely used to control T. absoluta (azadirachtin, spinosad and indoxacarb) on the mirid predators M. pygmaeus and N. tenuis. In Italy, the dustable sulphur showed to be moderately harmful (as a fresh residue) for the generalist predator N. tenuis [285]. Also, Biondi et al. [286] assessed the risks of 14 pesticides (synthetic and biopesticides), commonly used in tomato crops, on the generalist predator O. laevigatus. Among the tested biopesticides only Bacillus thuringiensis Berliner (Bacillales: Bacillaceae) proved to be harmless; whereas spinosad, emamectin benzoate and metaflumizone were moderately harmful until 7 days after the treatment. Abamectin was the most noxious and persistent. In Italy, also, abamectin and oxamyl are usually used to control T. absoluta. After treatment with these pesticides, both M. pygmaeus and N. tenuis failed to achieve the levels necessary for effective pest control [274]. In Brazil, dose-response regression lines were estimated for adults of the vespid predator B. lecheguana, used against T. absoluta, after exposing to the insecticides abamectin, cartap, phentoate, and permethrin with or without 0.5%(v/v)

mineral oil. Mineral oil increased selectivity in favor of the predator when in mixture with abamectin, permethrin, and cartap, but had a strong negative effect on phentoate selectivity [148]. Galvan et al. [147] assessed the selectivity of abamectin, acephate, deltamethrin, fenitrothion, fenpropathrin, methyl parathion and trichlorfon to the vespid predators B. lecheguana, P. sylveirae and P. exigua using insecticide concentrations corresponding to 50% and 100% of the current recommended rate for citrus caterpillars. The three predators were highly susceptible to both concentrations of fenitrothion, fenpropathrin and methyl parathion, and the highest concentration of trichlorfon. Very recently, Martinou et al. [294] evaluated the lethal effects of six insecticides and a fungicide on the predator M. pygmaeus nymphs exposed to the pesticides through three routes of exposure: direct, residual and oral. Chlorantraniliprole caused less than 25% mortality to M. pygmaeus and were classified as harmless. In contrast, thiacloprid and metaflumizone caused 100% and 80% mortality, respectively, and were classified as harmful. Indoxacarb resulted in close to 30% mortality to the predator, and was classified as slightly harmful, while the fungicide copper hydroxide caused 58% mortality and was rated as moderately harmful.

Concerning the side-effects of botanical extracts on predators of *T. absoluta*, Oliver and Bringas ^[106] evaluated the effects of neem and rotenone against the conventional synthetic pesticides managed by farmers in Peru, on the population variations of eggs, nymphs and adults of the stilt bug *M. tenellus* in the tomato crop. The results showed rotenone did not affect the developmental stages of *M. tenellus* and neem showed a slight effect while the high-test negative effect was observed after treatment with the conventional pesticides. Also, Iannacone and Lamas ^[295] tested the toxicological effects of extracts of two plants, Peruvian peppertree (*Schinus molle* L.) and lantana (*Lantana camara* L.) on the eggs, first stage larvae and pupae of the chrysopid predator *C. externa* under laboratory conditions in Perú. They discussed the possibility of using these botanicals in an IPM for *T. absoluta*.

6. Supplementation or integration between predators and some other biocontrol agents.

To test if the mirid predator *M. pygmaeus* would show preference between T. absoluta eggs and Trichogramma achaeae-parasitized (Hymenoptera: Trichogrammatidae) T. absoluta eggs, Desneux et al. [2, 296] carried out a choice experiment, and then tested if the predator would reduce the number of T. achaeae-parasitized T. absoluta eggs on whole tomato plants. Their findings suggested the importance of integrating the mirid predator M. pygmaeus presence in the greenhouse tomato crop when using oophagous parasitoids for inundative BC of T. absoluta. Also, it was found that the best BC of T. absoluta was obtained when M. pygmaeus and the parasitoid T. achaeae were released in combination [67, 297]. Considering another mirid bug, N. tenuis, an alternative release method for it and its combination with the eulophid parasitoid Necremnus artynes (Walker) (Hymenoptera: Eulophidae) had been evaluated. It was demonstrated that this alternative release method (pre-plant application) increased the predation capacity of N. tenuis and reduced control costs [298].

On the other hand, the integration of the bacterium *B. thuringiensis* treatments with the release or conservation of some mirid predators may provide a safe strategy to manage *T. absoluta* because *B. thuringiensis* targets larvae and mirids preferentially prey on *T. absoluta* eggs. Therefore, Molla *et al.* ^[29] hypothesized that *B. thuringiensis* treatments applied immediately after the initial

detection of *T. absoluta* on plants did not interfere with mirid establishment in the crop since *T. absoluta* eggs were available. Afterwards, treatments with *B. thuringiensis* could be terminated, allowing the mirid predator *N. tenuis* alone to control the pest. Calvo *et al.* ^[298] developed an early release system for *N. tenuis* aiming to provide a good control of *T. absoluta* in tomato in Spain. *B. thuringiensis* and the parasitic insect *T. achaeae* had been shown to be effective against *T. absoluta* and could be a supplement to the predator *N. tenuis*.

7. Conservation of indigenous natural predators.

Conservation biological control (CBC) is the practice of enhancing the efficacy of natural enemies' assemblages that already exist in the area through modification of the environment, such as biodiversity conservation, landscape aesthetics, provision of clean water, reduce soil water retention or soil erosion [72], or of existing pesticide practices [299, 300]. In spite to this importance, CBC has long been a rather neglected form of BC, but research in this field has increased markedly during the last decade [301-303]. Recent scientific reviews have considered CBC as a component of habitat manipulation [301, 304] or have focused on a part of CBC, e.g., plantprovided food for natural enemies [305]. Several considerations had been discussed [306, 307]. For some details, wild or cultivated flowering plants, including weeds, in or around the field, can increase the diversity of habitats and provide shelter, suitable microclimate and alternative food sources for the natural enemies and can significantly increase the residency times and thus enhance the efficacy of these biocontrol agents [304, 308]. Among natural enemies, the feeding habits of predators largely influence their ecological identities and consequently their effectiveness in BC. For example, omnivorous heteropterous predators can utilize food of different trophic levels (i.e., animal and plant food). Phytophagy constitutes a common trait which can support at an intermediate or low rate the development and reproduction of these omnivorous predators. Its effect in predators' performance is largely determined by the host plant species. Generally, plant feeding may enable them to remain on the crop during periods when their prey is scarce (for details, see: [72, 309,310]).

It is well known that T. absoluta is a dangerous pest in a wide range of pests infesting tomatoes $[^{2}, ^{6}, ^{15}, ^{16}]$. Despite the diverse pest complex $[^{311}]$, their major natural enemies' assemblage is largely constituted by few predatory omnivorous mirid species. In the Mediterranean region, M. pygmaeus, N. tenuis, D. tamaninii and D. errans and in the North America, the mirid predator D. hesperus are considered to be among the principal natural enemies of pests on tomatoes $[^{312-314}]$. Mirids are polyphagous, feeding on almost the entire spectrum of tomato pests. These mirid predators show phytophagous habits that support their development or reproduction to a variable rate depending on the plant substrate $[^{52}, ^{312}, ^{315}]$.

Alomar *et al.* ^[316] and Castañé *et al.* ^[317] showed that the colonization of tomato crops by native predators was higher in fields surrounded by complex agro-environments. Similarly, the environmental complexity may substantially influence the abundance of *M. pygmaeus* and *D. errans* in tomato greenhouses ^[70]. Further developments in CBC should be directed to identify which non-crop host plants support high numbers of these predators early in the season ^[318]. For example, *Solanum nigrum* (Solanaceae) is a major host plant for the predator *M. pygmaeus* in Greece ^[319]. Recent data have shown that *M. pygmaeus* can complete development on *S. nigrum* without prey and can also

reproduce when infested by its main native aphid pest. Therefore, this plant should be evaluated in CBC systems, as a natural reservoir of M. pygmaeus for tomato crop colonization [320]. In greenhouse tomatoes, the use of alternative host plants has been evaluated as a means to introduce the mirid predator, D. hesperus. This predator was invariably found to be more common in greenhouses where the alternative host plant, Verbascum thapsus (Scrophulariaceae), had been introduced, providing also a refuge for the predator when prey on tomato was scarce. In tomato unheated greenhouses, Arnó et al. [321] showed that tobacco plants can serve as a refuge for the mirid predators during winter that may enhance the timely and efficient colonization of the next spring tomato crop. As a measure within the frame of CBC, it has been also suggested that the farmers could collect predators on crop plants at the end of the crop cycle or on major non-crop plants for subsequent release in greenhouses [319, 322, 323]. In addition, the tomato cultivars can affect the efficiency of natural enemies of T. absoluta. As reported by Cabello et al. [85], tomato cultivars were observed to influence the activity of natural enemies, mainly the predator N. tenuis in Spain. This may be because of differences in plant nutrients in different cultivars, which may affect the feeding of omnivorous insects. In contrast, cultivar effects on the eggparasitoid T. achaeae were less apparent or possibly nonexistent. Nevertheless, there was an indirect effect in as much as T. achaeae was favored in cultivars not liked by the predator N. tenuis.

In **conclusion**, different predator species belonging to several insect orders, such as Heteroptera, Hymenoptera, Dermaptera, Coleoptera, Neuroptera and Thysanoptera, as well as some predator species belonging to a number of arachnid families, such as Phytoseiidae and Lycosidae, have been beneficial as biocontrol agents against *T. absoluta*. However, more research should be conducted for the integration between these predators and some other biocontrol agents. Also, conservation of the most efficient indigenous natural predators and selectivity of insecticides should be necessarily studied in detail.

8. Reference

- 1. Gomide EVA, Vilela EF, Picanço M. Comparison of sampling procedures for *Tuta absoluta* (Meyrick) (Lepidoptera: Gelechiidae) in tomato crop. Neotrop. Entomol., 2001; 30 (4): 697-705.
- 2. Desneux N, Wajnberg E, Wyckhuys KAG, Burgio G, Arpaia S, Narváez-Vasquez CA, González-Cabrera J, Ruescas DC, Tabone E, Frandon J. Biological invasion of European tomato crops by *Tuta absoluta*: ecology, geographic expansion and prospects for biological control. J Pest Sci 2010; 83(3): 197-215.
- 3. Gontijo PC, Picanço MC, Pereira EJG, Martins JC, Chediak M, Guedes RC. Spatial and temporal variation in the control failure likelihood of the tomato leaf miner *Tuta absoluta*. Ann Appl Biol 2013; 162(1): 50-59.
- 4. Urbaneja A, Vercher R, Navarro V, Garcia Mari F, Porcuna JL. La polilla del tomate *Tuta absoluta*. Phytoma España 2007; 194: 16-23.
- 5. Arno J, Gabarra R. Side effects of selected insecticides on the *Tuta absoluta* (Lepidoptera: Gelechiidae) predators *Macrolophus pygmaeus* and *Nesidiocoris tenuis* (Hemiptera: Miridae). J Pest Sci 2011; 84(4): 513-520.
- 6. Garcia-Mari F, Vercher R. Description origin and expansion of *Tuta absoluta* (Lepidoptera: Gelechiidae). Phytoma España 2010; 217: 16-20.

- 7. Torres JB, Faria CA, Evangelista WA, Pratissol D. Within-plant distribution of the leaf miner *Tuta absoluta* (Meyrick) immatures in processing tomatoes with notes on plant phenology. Int J Pest Manage 2001; 47: 173-178.
- 8. EPPO (European and Mediterranean Plant Protection Organization). *Tuta absoluta* (GNORAB). http://photos.eppo.org/index.php/search/tuta [Accessed on 10 June 2009].
- 9. Seplyarsky V, Weiss M, Haberman A. *Tuta absoluta* Povolny (Lepidoptera: Gelechiidae), a new invasive species in Israel. Phytoparasitica 2010; 38(5): 445-446.
- Ostrauskas H, and Ivinskis P, Records of the tomato pinworm *Tuta absoluta* (Meyrick, 1917) (Lepidoptera: Gelechiidae) in Lithuania Acta Zool Lituanica 2010; 20(2): 151-155.
- 11. Desneux N, Luna M G, Guillemaud T, Urbaneja A. The invasive South American tomato pinworm *Tuta absoluta* continues to spread in Afro-Eurasia and beyond the new threat to tomato world production J Pest Sci 2011; 84: 403-408.
- 12. Guedes RNC, Picanço MC. The tomato borer *Tuta absoluta* in South America pest status management and insecticide resistance EPPO Bull 2012; 42(2): 211-216.
- 13. Megido RC, Haubruge E, Verheggen FJ. First evidence of deuterotokous parthenogenesis in the tomato leafminer *Tuta absoluta* (Meyrick) (Lepidoptera: Gelechiidae) J Pest Sci 2012; 85(4): 409-412.
- 14. Apablaza J, La polilla del tomate y su manejo Tattersal Pesqui Agropecu Bras 1992; 79:12-13 (In Spanish with English summary).
- 15. Öztemiz S. The tomato leafminer [(*Tuta absoluta* Meyrick (Lepidoptera: Gelechiidae)] and it's biological control KSU J Nat Sci 2012; 15(4): 47-57. (In Turkish with English summary).
- 16. Mollá HÓ. Control biológico de la polilla del tomate *Tuta absoluta* (Lepidoptera: Gelechiidae) mediante la gestión de míridos depredadores Ph.D. Thesis Universitat de València España, 2013, 214. (In Spanish with English summary).
- 17. Garzia GT, Siscaro G, Biondi A, Zappalà L. *Tuta absoluta* a South American pest of tomato now in the EPPO region biology distribution and damage. EPPO Bull 2012; 42(2): 205-210.
- 18. Collavino MD, Giménez RA. Efficacy of imidacloprid to control the tomato borer (*Tuta absoluta* Meyrick). IDESIA (Chile) 2008; 26(1): 65-72.
- 19. Ortega F, Astor E, Scals D. El control de la polilla del tomate (*Tuta absoluta*). Hortic Int 2008; 64: 30-31.
- 20. Estay P, Bruna A. Insectos y acaros asociados al tomate en Chile In (Estay P, Bruna A., eds.) "Insectos, acaros y enfermed-ades asociados al tomate en Chile". Centro regional de Investigacion INIA La Platina, Santiago, Chile, 2002, 9-22.
- 21. Siqueira HAA, Guedes RNC, Picanço MC. Insecticide resistance in populations of *Tuta absoluta* (Lepidoptera: Gelechiidae). Agric Forest Entomol 2000; 2(2): 147-153.
- 22. Siqueira HAA, Guedes RNC, Picanço MC. Cartap resistance and synergism in populations of *Tuta absoluta* (Lep., Gelechiidae). J Appl Entomol 2000; 124(5-6): 233-238.
- 23. Lietti MMM, Botto E, Alzogaray RA. Insecticide

- resistance in Argentine populations of *Tuta absoluta* (Meyrick) (Lepidoptera: Gelechiidae). Neotrop Entomol 2005; 34(1):113-119.
- 24. Silva GA, Picanço MC, Bacci L, Crespo ALB, Rosado JF, Guedes RNC. Control failure likelihood and spatial dependence of insecticide resistance in the tomato pinworm, *Tuta absoluta*. Pest Manage Sci 2011; 67(8): 913-920.
- 25. Siqueira HAA, Guedes RNC, Fragoso DB, Magalhaes LC. Abamectin resistance and synergism in Brazilian populations of *Tuta absoluta* (Meyrick) (Lepidoptera: Gelechiidae). Int J Pest Manage, 2001; 47(4): 247-251.
- 26. Bielza P. Resistance to insecticides in *Tuta absoluta* (Meyrick). Phytoma España 2010; 217: 103-106.
- 27. Batalla-Carrera L, Morton A, Garcia-del-Pino F. Efficacy of entomopathogenic nematodes against the tomato leafminer *Tuta absoluta* in laboratory and greenhouse conditions. BioControl 2010; 55:523-530.
- 28. Gonzalez-Cabrera J, Molla O, Monton H, Urbaneja A. Efficacy of *Bacillus thuringiensis* (Berliner) in controlling the tomato borer *Tuta absoluta* (Meyrick) (Lepidoptera: Gelechiidae). BioControl 2011; 56:71-80.
- 29. Molla O, Gonzalez-Cabrera J, Urbaneja A. The combined use of *Bacillus thuringiensis* and *Nesidiocoris tenuis* against the tomato borer *Tuta absoluta*. BioControl 2011; 56: 883-891.
- 30. Vacas S, Alfaro C, Primo J, Navarro-Llopis V. Studies on the development of a mating disruption system to control the tomato leafminer *Tuta absoluta* Povolny (Lepidoptera: Gelechiidae). Pest Manage Sci 2011; 67(11): 1473-1480.
- 31. Zappala L, Bernardo U, Biondi A, Cocco A, Deliperi S, Delrio G, Giorgini M, Pedata P, Rapisarda C, Garzia GT, Siscaro G. Recruitment of native parasitoids by the exotic pest *Tuta absoluta* in Southern Italy. Bull Insectol 2012; 65 (1): 51-61.
- 32. Savino V, Coviella CE, Luna MG. Reproductive biology and functional response of *Dineulophus phtorimaeae* a natural enemy of the tomato moth *Tuta absoluta*. J Insect Sci 2012; 12: 1-14.
- Greathead DJ. Parasitoids in classical biological control In (Waage, J. and Greathead, D., eds.) "Insect Parasitoids".
 13th Symposium of Royal Entomological Society of London, 18-19. Sept. 1985. Academic Press, London, 1986, 289-318.
- 34. Öztemiz S. Population of *Tuta absoluta* and natural enemies after releasing on tomato grown greenhouse in Turkey. African J Biotechnol 2013; 12(15): 1882-1887.
- 35. Vivan LM, Torres JB, Veiga AFSL. Development and reproduction of a predatory stinkbug *Podisus nigrispinus* in relation to two different prey types and environmental conditions. Biocontrol 2003; 48:155-168.
- 36. Bale JS, van Lenteren JC, Bigler F. Biological control and sustainable food production. Philos Trans Royal Society B 2008; 363: 761-776.
- 37. Miranda MMM, Picanco M, Zanuncio JC, Guedes RNC. Ecological life table of *Tuta absoluta* (Meyrick) (Lepidoptera: Gelechiidae). Biocontrol Sci Technol 1998; 8: 597-606.
- 38. Marchiori CH, Silva CG, Lobo AP. Parasitoids of *Tuta absoluta* (Meyrick, 1917) (Lepidoptera: Gelechiidae) collected on tomato plants in lavras State of Minas Gerais

- Brazil. Braz Brazilian J Biol 2004; 64(3A): 552-554.
- 39. Medeiros MA, Vilela NJ, França FH. Technical and economic efficiency of biological control of the South American tomato pinworm in protected environment Horticultura. Brasileira 2006; 24: 180-184.
- 40. Urbaneja A, González-Cabrera J, Arnó J, Gabarra R. Prospects for the biological control of *Tuta absoluta* in tomatoes of the Mediterranean basin. Pest Manage Sci 2012; 68(9): 1215-1222.
- 41. Van Driesche R, Bellows TS. Jr Biological Control Springer New York, 1996, 539.
- 42. Smee D. Species with a large impact on community structure Nature Education Knowledge 2012; 3(10): 40.
- 43. Botkin D, Keller E. Environmental Science Earth as a living planet. 7th ed, John Wiley & Sons Inc, 2010, 752.
- 44. Snyder WE, Wise DH. Predator interference and the establishment of generalist predator populations for biocontrol. Biological Control 1999; 15: 283-292.
- 45. Hassell MP. The dynamics of arthropod predator-prey systems. Princeton University, New Jersey, 1978, 23.
- 46. Rosenheim JA, Wilhoit LR, Armer CA. Influence of intraguild predation among generalist insect predators on the suppression of an herbivore population. Oecologia 1993; 96: 439-449.
- 47. Tscharntke T, Steffan-Dewenter I, Kruess A, Thies C. Characteristics of insect populations on habitat fragments A mini review. Ecol Res 2002; 17(2): 229-239.
- 48. Alomar O. Facultative predation as a biological control In (Pimentel, D., ed). Encylopedia of Pest Management. Marcel Dekker Inc, New York, 2002, 1-3.
- 49. Cassis G, Schuh RT. Systematics Biodiversity Biogeography and Host Associations of the Miridae (Insecta: Hemiptera: Heteroptera: Cimicomorpha). Annu Rev Entomol 2012; 57: 377-404.
- 50. Hamdi F, Chermiti B, Gauthier N, Streito J-C, Bonato O. *Macrolophus pygmaeus* Wagner (Heteroptera: Miridae) a promising biological control agent for protected tomato crop in Tunisia. International Symposium "Crop Protection Management in Mediterranean Organic Agriculture" May 14-16, 2013, Sousse (Tunisia). Book of Abstracts, p19.
- 51. Sanchez JA, Gillespie DR, McGregor RR, The effects of mullein plants (*Verbascum thapsus*) on the population dynamics of *Dicyphus hesperus* (Heteroptera: Miridae) in tomato greenhouses. Biological Control 2003; 28(3): 313-319
- 52. Perdikis D, Lykouressis P. Thermal requirement for development of the polyphagous predator *Macrolophus pygmaeus* (Hemiptera: Miridae). Environ Entomol 2002; 31: 661-667.
- 53. Calvo J, Urbaneja A. *Nesidiocoris tenuis* Reu (Het.: Miridae) en tomate Amigo o Enemigo. Almeria Verde 2003; 4: 21-23.
- 54. Kerzhner JM, Josifov M. Catalogue of the Heteroptera of the Palaearctic Region. Miridae In Aukema, B. and Rieger, Ch., eds, Vol. 3, i-xiv, The Netherlands Entomol Soc, Amesterdam, 1999, 1-577.
- 55. Carapezza A. The Specific Identities of *Macrolophus melanotoma* (Costa, A. 1853) and *Stenodema curticolle* (Costa, A. 1853) (Insecta Heteroptera, Miridae). Natural Siliciano S IV 1995; 19: 295-298.

- 56. Constant B, Grenier S, Bonnot G. Artificial substrate for egg laying and embryonic development by the predatory bug *Macrolophus caliginosus* (Heteroptera: Miridae). Biological Control 1996; 7(2): 140-147.
- 57. Malausa JC, Trottin CY. Adavances in the strategy of use of the predaceous bug *Macrolophus caliginosus* (Heteroptera: Miridae) in glasshouse crops Thomas Say. Publ Entomol Lanham MD 1996, 178-189.
- 58. Bacci L. Factors determining the attack of *Tuta absoluta* on tomato M.Sc Universidade Federal de Viçosa Minas Gerais Brazil, 2006.
- 59. Arno J, Mussoll A, Gabarra R, Sorribas R, Prat M, Garreta A, Gomez A, Matas M, Pozo C, Rodriguez D. *Tuta absoluta* una nueva plaga en los cultivos de tomate Phytoma. Espana 2009; 211: 16-22. (In Spanish with English summary).
- 60. Mollá Ó, Montón H, Vanaclocha P, Beitia F, Urbaneja A. Predation by the mirids *Nesidiocoris tenuis* and *Macrolophus pygmaeus* on the tomato borer *Tuta absoluta* Proceedings of the IOBC/WPRS Working Group. "Integrated Control in Protected Crops" Crete Greece 6-11 September 2009 IOBC/WPRS Bull 49: 209-214.
- 61. Urbaneja A, Monton H, Molla O. Suitability of the tomato borer *Tuta absoluta* as prey for *Macrolophus caliginosus* and *Nesidiocoris tenuis*. J Appl Entomol 2009; 133: 292-296.
- 62. Morley P, Jacobson R, Challinor P. Organic tomato Phase 1 of contingency plans for the control of *Tuta absoluta* and *Nesidiocoris tenuis* Final report of the project at Horticilha Cilha Queimada Alcochete Portugal Wight Salads Group Arreton Isle of Wight PO30 3AR RJC Ltd Bramham W Yorks LS23 6TH 2010 48pp.
- 63. Guenaoui Y, Bensaad R, Ouezzani K. Importance of native polyphagous predators able to prey on *Tuta absoluta* Meyrick (Lepidoptera: Gelechiidae) on tomato crop (Algeria) EPPO/IOBC/FAO/NEPPO Joint International symposium on management of *Tuta absoluta*-Agadir-Morocco November 16-18 2011.
- 64. Pumariño L, Alomar O, Agustí N. Development of specific ITS markers for plant DNA identification within herbivorous insects. Bull Entomol Res 2011; 101(3): 271-276.
- 65. Straten M.J, Potting RPJ, Linden A. Introduction of the tomato leafminer *Tuta absoluta* into Europe. Proc Neth Entomol Soc Meet 2011; 22: 23-30.
- 66. Boualem M, Allaoui H, Hamadi R, Megahed M. Biology and complex of natural enemies of *Tuta absoluta* in Mostaganem (Algeria). EPPO Bull 2012; 42 (2): 268-274.
- 67. Chailleux A, Bearez P, Pizzol J, Amiens-Desneux E, Ramirez-Romero R, Desneux N. Potential for combined use of parasitoids and generalist predators for biological control of the key invasive tomato pest *Tuta absoluta*. J Pest Sci 2013; 86(3): 533-541.
- 68. Dahliz A, Lakhdari W, Soud A, Hammi H, Bouchekima H, Belaidi M. Complex of natural enemies and control methods of the exotic invasive pest *Tuta absoluta* (Lepidoptera: Gelechiidae) in Southern Algeria CTAB-ISOFAR-MOAN Symposium 2013 Crop protection management in Mediterranean Organic Agriculture 14-16 May 2013 in Sousse Tunisia (Ben Kheder, M. and Neuhoff, D.; eds.) (Book of Abstracts) p.48.

- 69. Biondi A, Chailleux A, Lambion J, Han P, Zappalà L, Desneux N. Indigenous natural enemies attacking *Tuta absoluta* (Lepidoptera: Gelechiidae) in Southern France. Egypt J Biol Pest Control 2013; 23(1): 117-121.
- Ingegno BL, Pansa MG, Tavella L. Tomato colonization by predatory bugs (Heteroptera: Miridae) in agroecosystems of NW Italy IOBC/WPRS Bull 2009; 49: 287-291.
- 71. Ingegno BL, Ferracini C, Gallinotti D, Alma A, Tavella L. Evaluation of the effectiveness of *Dicyphus errans* (Wolff) as predator of *Tuta absoluta* (Meyrick). Biological Control 2013; 67(2): 246-252.
- 72. Perdikis D, Fantinou A, Lykouressis D. Enhancing pest control in annual crops by conservation of predatory Heteroptera. Biological Control 2011; 59: 13-21.
- 73. Arno J, Sorribas R, Prat M, Matas M, Pozo C, Rodríguez D, Garreta A, Gómez A, Gabarra R. *Tuta absoluta* a new pest in IPM tomatoes in the northeast of Spain IOBC/WPRS Bull 2009; 49: 203-208.
- 74. Molla Ó, Alonso M, Monton H, Beitia F, Verdu MJ, Gonzalez-Cabrera J, Urbaneja A. Biological control of *Tuta absoluta* Survey of natural enemies and role of mirid predators as biocontrol agents Phytoma España 2010; 217: 42-47. (In Spanish with English summary).
- Van der Blom J. La introduccion artificial de la fauna auxiliar en cultivos agricolas Boletin de Sanidad Vegetal. Plagas 2002; 28:109-120 (In Spanish with English summary).
- 76. Sanchez JA, Martinez JI, Lacasa A. Distribution and abundance of mirids in horticultural crops in the region of Murcia (Spain) In 3rd International *Bemisia* Workshop Barcelona 17-20 March 2003 Barcelona Spain Solsoloy AD Domi.
- 77. El-Arnaouty SA, Kortam MN. First record of the mired predatory species *Nesidiocoris tenuis* Reuter (Heteroptera: Miridae) on the tomato leafminer *Tuta absoluta* (Meyrick) (Lepidoptera: Gelechiidae) in Egypt. Egypt J Biol Pest Control 2012; 22(2): 223-224.
- 78. Carnero A, Diaz S, Amador S, Hernández M, Hernández E. Impact of *Nesidiocoris tenuis* Reuter (Hemiptera: Miridae) on whitefly population in protected tomato crops IOBC/WPRS. Bull 2000; 23 (1): 259.
- 79. Devi PK, Yadav DN, Anand J. Role of *Nesidiocoris tenuis* Reuter (Hemiptera: Miridae) in natural suppression of tomato fruit borer *Helicoverpa armigera* (Hubner) (Lepidoptera: Noctuidae) Pest Manage. Horticul Ecosys 2002; 8: 109-113.
- 80. Torres JB, Evangelista WS, Barras R, Guedes RNC. Dispersal of *Podisus nigrispinus* (Het., Pentatomidae) nymphs preying on tomato leafminer effect of predator release time density and satiation level. J Appl Entomol 2002; 126: 326-332.
- 81. García M, Strassera ME, Luna MG, Polack LA, Mezquiriz N. Monitoreo de plagas. Boletín Hortícola 2005; 10(31): 31-37 (In Spanish with English summary).
- 82. Urbaneja A, Monton H, Vanaclocha P, Molla O, Beitia F. La polilla del tomate *Tuta absoluta* una nueva presa para los miridos *Nesidiocoris tenuis* y *Macrolophus pygmaeus*. Agricola Vergel 2008; 320: 361-367 (In Spanish with English summary).
- 83. Abbes K, Harbi A, Chermiti B. The tomato leafminer Tuta

- absoluta (Meyrick) in Tunisia current status and management strategies. EPPO Bull 2012; 42(2): 226-233.
- 84. Al-Jboory IJ, Katbeh-Bader A, Shakir A. First observation and identification of some Natural enemies collected from heavily infested tomato by *Tuta absoluta* (Meyrick) (Lepidoptera: Gelechiidae) in Jordan Middle-East. J Sci Res 2012; 11 (6): 787-790.
- 85. Cabello T, Gallego JR, Fernandez FJ, Gamez M, Vila E, Pimo MD, Hernandez-Suarez E. Biological control trategies for the South American tomato moth (Lepidoptera: Gelechiidae) in greenhouse tomatoes. J Econ Entomol 2012; 105(6): 2085-2096.
- 86. Durán CL. Evaluacion de *Trichogramma cacoeciae* como parasitoide de *Tuta absoluta* M.Sc. Thesis Universidad de Almeria Escuela Politecnica Superior y Facultad de Ciencias. Experimentales, 2013, 75. (In Spanish with English summary).
- 87. Biondi A. Combining natural enemies and selective pesticides in IPM programmes of exotic pests the *Tuta absoluta* (Lepidoptera: Gelechiidae) case Ph.D. Thesis Archivio istituzionale dell'Universita' di Catania Italy 2013 (In Italian with English summary).
- 88. Muñoz O, A Villalba Velásquez, V.; López, S.N. Development of *Tupiocoris cucurbitaceus* (Hemiptera: Miridae) on *Bemisia tabaci* (Hemiptera: Aleyrodidae) in several vegetables. Fitosanidad, 2012; 16(3): 147-153.
- 89. López SN, Rojas FA, Velásquez VV, Cagnotti C. Biology of *Tupiocoris cucurbitaceus* (Hemiptera: Miridae) a predator of the greenhouse whitefly *Trialeurodes vaporariorum* (Hemiptera: Aleyrodidae) in tomato crops in Argentina. Biocontrol Sci Technol 2012; 22(10): 1107-1117.
- 90. Lopez SN. Evaluación de atributos biológicos de *Tupiocoris cucurbitaceus* (Hemiptera: Miridae) chinche predadora de moscas blancas en cultivos hortícolas INTA-Instituto de Microbiología y Zoología Agrícola-Boletin MIP 2010; 15 (In Spanish with English summary).
- 91. Messing RH, Aliniazee MT. Impact of predaceous insects on filbert aphid *Myzocallis coryli* (Homoptera: Aphididae). Environ Entomol 1986; 15(5): 1037-1041.
- 92. Alhmedi A, Haubruge E, Bodson B, Francis F. Aphidophagous guilds on nettle (*Urtica dioica*) strips closeto fields of green pea rape and wheat. Insect Science 2007; 14: 419-424.
- 93. Guidone L, Loru L, Marras PM, Fois X, Pantaleoni RA, Tavella L. Predatory bugs in hazelnut orchards of Piedmont and Sardinia (Italy). Bull Insectol 2008; 61(1): 207-208.
- 94. Lattin JD. Bionomics of the Anthocoridae Annu Rev Entomol 1999; 44: 207-231.
- 95. Kiman ZB, Yeargan KV. Development and reproduction of the predator *Orius insidiosus* (Hemiptera: Anthocoridae) reared on diets of selected plant material and arthropod prey. Ann Entomol Soc Amer 1985; 78: 464-467.
- 96. Ghahari H, Carpintero DL, Ostoran H. An annotated catalogue of the Iranian Anthocoridae (Hemiptera: Heteroptera: Cimicomorpha) Acta Entomol Musei. Nationalis Pragae 2009; 49(1): 43-58.
- 97. Lins Jr JC, Bueno VHP, Silva DB, van Lenteren JC, Calixto AM Sidney LA. *Tuta absoluta* egg predation by

- Orius insidiosus. IOBC/wprs Bull 2011; 68: 101-104.
- 98. Bueno VHP, Montes FC, Pereira AMC, Lins JC Jr, van Lenteren JC. Can recently found Brazilian hemipteran predatory bugs control *Tuta absoluta*. IOBC-WPRS Bull 2012; 80: 63-67.
- 99. Puysseleyr V, de Höfte M, Clercq P. de 2014 Continuous rearing of the predatory anthocorid *Orius laevigatus* without plant materials. J Appl Entomol 138(1-2) 45-51.
- 100.Pericart J. Family Anthocoridae Fieber 1836-flower bugs minute pirate bugs Pp 108-140 In (Aukema, B. and Rieger, Ch., eds.) "Catalogue of the Heteroptera of Palaearctic region" Vol 2, Cimicomorpha 1. The Netherlands Entomol Soc Amesterdam 1996, xiv+361.
- 101.Hamdan AJ, Abu-Awad I. Effect of host plants on predatory-prey relationship between predatory bug *Orius laevigatus* (Fiber) (Hemiptera: Anthocoridae) and tobacco whitfly *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae). An_Najah Univ Res J (N.Sc.) 2007; 21: 85-99.
- 102. Sobhy IS, Sarhan AA, Shoukry AA, El-Kady GA, Mandour NS, Reitz SR. Development consumption rates and reproductive biology of *Orius albidipennis* reared on various prey. BioControl, 2010; 55(6): 753-765.
- 103.Sayed SM, Montaser MM, Elsayed G, Amer SAM. Preliminary molecular identification of a predatory bug *Orius albidipennis* collected from ornamental plants. J Insect Sci 2013; 13:11. Available online: http://www.insectscience.org/13.11
- 104.Papavizas GC. Biological control in crop production Allanheld Osmum & Co Publisher Inc New Jersey 1981 461pp.
- 105.Henry TJ. Biodiversity of the Heteroptera In (Foottit R.G. and Adler P.H., eds.) "Insect biodiversity: Science and society" Wiley-Blackwell Oxford 2009, 223-263.
- 106.Oliver JAI, Bringas YM. Effects on the populations of the predator *Metacanthus tenellus* (Heteroptera: Berytidae) by the botanic insecticides rotenone and neem on tomato crop in Peru. Rev Colomb Entomol 2000: 26(3/4): 89-97.
- 107.Berskaya EA, Sharov AA, Izhevskii SS. Predators of the American white web-worm moth (*Hyphantria cunea*) in the south of the European part of the USSR. Zool Zh 1985; 64: 1384-91 (In Russian with English summary).
- 108.Irwin ME, Shepard M. Sampling predaceous Hemiptera on soybean In (Kogan, M. and Herzog, D.C. eds.) "Sampling Methods in Soybean Entomology" New York/Heidelberg/ Berlin Springer-Verlag 1980, 505-31.
- 109.Gámez MZ, Sebestyén Z, Varga J, Garay JR, Gallego FJ, Fernández T. Cabello Multi-stage dynamic model for a prey-predator interaction Application to *Spodoptera exigua* (Hubner) (Lep.: Noctuidae) and *Nabis pseudoferus* Remane (Hem.: Nabidae) in greenhouse conditions IOBC-WPRS-Working Group Conference on Integrated Control in Protected Crops-Mediterranean Climate Catania (Italy), 9-12 October 2012.
- 110.Cabello T, Gallego JR, Fernandez FJ, Soler A, Beltran D, Parra A, Vila E. The damsel bug *Nabis pseudoferus* (Hem.: Nabidae) as a new biological control agent of the South American tomato pinworm *Tuta absoluta* (Lep.: Gelechiidae) in tomato crops of Spain OIBC/WPRS Bull 2009; 49: 219-223.
- 111. Cabello T, Gallego JR, Vila E, Soler A, Parra A,

- Fernandez FJ. Biological control of *Tuta absoluta* with egg parasitoids in Spain. Phytoma España 2010; 217: 53-59 (In Spanish with English summary).
- 112. Schuh RT, Slater JA. True Bugs of the World (Hemiptera: Heteroptera) Classification and Natural History Cornell University Press, Ithaca, 1995, 336.
- 113.Bell KO, Whitcomb WH. Field studies on egg predators of the bollworm *Heliothis zea* (Boddie) Florida Entomologist 1964; 47: 171-180.
- 114.Orhanides GM, Gonzalez D, Bartlett BR. Identification and evaluation of pink bollworm predators in southern California. J Econ Entomol 1971; 64: 421-424.
- 115.Olson DM, Wäckers FL. Management of field margins to maximize multiple ecological services. J Appl Ecol 2007; 44: 13-21.
- 116.Zanuncio TV, Freitas FA de, Tavares WS, Menezes CWG de, Serrão JE, Zanuncio JC. *Podisus nigrispinus* (Heteroptera: Pentatomidae) and IAC-24 soybean cultivar are compatible Sustain. Agric Res 2012; 1(2): 41-50.
- 117.Swanson DR. An updated synopsis of the Pentatomoidea (Heteroptera) of Michigan Great Lakes. Entomologist 2012; 45: 263-311.
- 118.Torres J B, Zanuncio J C, Moura MA. The predatory stinkbug *Podisus nigrispinus* biology ecology and augmentative releases for lepidoperan larval control in *Eucalyptus* forests in Brazil Perspectives in Agriculture. Veterinary Science Nutrition and Natural Resources 2006; 1(15): 1-18.
- 119.Rodrigues AR, Torres JB, Siqueira HA, Teixeira VW. *Podisus nigrispinus* (Dallas) (Hemiptera: Pentatomidae) requires long matings for successful reproduction. Neotrop Entomol 2009; 38(6): 746-53. (In Portuguese with English summary).
- 120.Malaquias JB, Ramalho FS, Fernandes FS, Nascimento Júnior JL, Correia ET, Zanuncio JC. Effects of photoperiod on reproduction and longevity of *Podisus nigrispinus* (Heteroptera: Pentatomidae). Ann Entomol Soc Amer 2010; 103(4): 603-610.
- 121. Vivan LM, Torres JB, Barros R, Veiga AFSL. Population growth rate of the depredating *Podisus nigrispinus* (Heteroptera: Pentatomidae) and of the *Tuta absoluta* (Leptoptera: Gelechiidae) in wintering place. Rev Biol Trop 2002; 50(1): 145-153.
- 122. Vivan LM, Torres JB, Veiga AFDL, Zanuncio JC, Predatory behavior and food conversion of *Podisus nigrispinus* preying on tomato leafminer Pesqui. Agropecu Bras 2002; 37: 581-587.
- 123.Oliveira CRF, Matos CHC, Hatano E. Occurrence of *Pyemotes* sp on *Tuta absoluta* (Meyrick). Brazilian Arch Biol Technol 2007; 50(6): 929-932.
- 124.Picker M, Griffiths CL, Weaving A. Field Guide to Insects of South Africa New Holland Publishers Ltd, 2004, 444.
- 125. Weirauch C, Munro JB. Molecular phylogeny of the assassin bugs (Hemiptera: Reduviidae) based on mitochondrial and nuclear ribosomal genes. Molec Phylogen Evol 2009; 53(1): 287-299.
- 126.Sahayaraj K, Kanna AV, Kumar SM. Gross Morphology of Feeding Canal Salivary Apparatus and Digestive Enzymes of Salivary Gland of *Catamirus brevipennis* (Servile) (Hemiptera: Reduviidae). J Entomol Res Soc

- 2010; 12 (2): 37-50.
- 127. Duncan I. Second review of progress on cashew production processing and marketing in cooperative de reforma agrarian la maranonera de RL Coralama U.S. Agency for International Development (USAID) Washington D.C, 1999.
- 128.Sühs RB, Somavilla A, Putzke J, Köhler A. Pollen vector wasps (Hymenoptera, Vespidae) of *Schinus terebinthifolius* Raddi (Anacardiaceae) Santa Cruz do Sul RS Brazil. Brazilian J Biosci 2009; 7(2): 138-143.
- 129.Raveret RM. Social wasp (Hymenoptera: Vespidae) foraging behavior. Annu Rev Entomol 2000; 45:121-150.
- 130.Gonring AHR, Picanço MC, Zanuncio JC, Puiatti M, Semeão AA. Natural biological control and key mortality factors of the pickleworm *Diaphania nitidalis* Stoll (Lepidoptera: Pyralidae) in cucumber. Biol Agric Hortic 2003; 20: 365-380.
- 131.Gonring AHR, Picanço MC, Guedes RNC, Silva EM. Natural biological control and key mortality factors of *Diaphania hyalinata* (Lepidoptera: Pyralidae) in cucumber. Biocontrol Sci Technol 2003; 13: 361-366.
- 132. Prezoto F, Santos-Prezoto HH, Machado VLL, Zanuncio JC. Prey captured and used in *Polistes versicolor* (Olivier) (Hymenoptera: Vespidae) nourishment. Neotrop Entomol 2006; 35(5): 707-709.
- 133.Pereira EJG, Picanço MC, Bacci L, Crespo ALB, Guedes RNC. Seasonal mortality factors of the coffee leafminer *Leucoptera coffeella*. Bull Entomol Res 2007; 97: 421-432.
- 134. Pereira EJG, Picanço MC, Bacci L, Della Lucia TMC, Silva EM, Fernandes FL. Natural mortality factors of *Leucoptera coffeella* (Lepidoptera: Lyonetiidae) on *Coffea arabica*. Biocontrol Sci Technol 2007; 17: 545-553.
- 135.Rayor LS. Effects of monarch larval host plant chemistry and body size on *Polistes* wasp predation In (Oberhauser, K.S. and Solensky, M.J., eds.) "The Monarch Butterfly: Biology and Conservation" Cornell University Press Ithaca NY, 2004, 39-46.
- 136.Picanco MC, Bacci L, Queiroz RB, Silva GA, Miranda MMM, Leite GLD, Suinaga FA. Social wasp predators of *Tuta absoluta*. Sociobiology 2011; 58(3): 621-633.
- 137.Drees BM, Jackman J. Field Guide to common Texas Insects. Taylor Trade Publishing, 1998, 352.
- 138.Gould WP, Jeanne RL. Polistes wasps (Hymeoptera: Vespidae) as control agents of lepidopterous cabbage pests. J Econ Entomol 1984; 13:150-156.
- 139.Stamp NE, Bowers MD. Direct and indirect effects of predatory wasps (*Polistes* sp Vespidae) on gregarious caterpillars (*Hemileuca lucina*: Saturniidae). Oecologia 1988; 75(4): 619-624.
- 140.Melo EL. Potencial de Control Biológico en el Manejo de las Plagas de la Yuca In (Ospina, B. and Ceballos, H. eds.) "La Yuca en el Tercer Milenio Sistemas Modernos de Producción Procesamiento Utilización y Comercialización" CIAT Publicación, 2003, 234-249 (In Spanish).
- 141.Reis PR, Souza JC, Zacarias MS. Alerta para o bichomineiro Cultivar 2006; 8: 13-16.
- 142.Prezoto F, Lima MAP, Machado VLL. Survey of preys captured and used by *Polybia platycephala* (Richards) (Hymenoptera: Vespidae: Epiponini). Neotrop Entomol

- 2005; 34: 849-851.
- 143.Bichara CC, Santos GMD, Resende JJ, Da Cruz JD, Gobbi N, Machado VLL. Foraging behavior of the swarm-founding wasp *Polybia* (*Trichothorax*) *sericea* (Hymenoptera, Vespidae) prey capture and load capacity. Sociobiology 2009; 53: 61-69.
- 144. Souza GK, Pikart TG, Jacques GC, Castro AA, Souza MM de, Serrão JE, Zanuncio JC. Social wasps on *Eugenia uniflora* Linnaeus (Myrtaceae) plants in an urban area. Sociobiology 2013; 60(2): 204-209.
- 145.West-Eberhard MJ, Carpenter JM, Hanson PE, Familia Vespidae. In (Hanson PE and Gauld ID eds) "Hymenoptera de la region tropical" Gainesville Memoirs of the American. Entomological Institute 2006; 77: 617-644.
- 146. Torres C, Dambolena JS, Zunino MP, Galetto L. Nectar characteristics and pollinators for three native, co-occurring insect pollinated *Passiflora* (Passifloraceae) from central Argentina. Int J Plant Reprod Biol 2012; 4: 121-126.
- 147.Galvan TL, Picanço MC, Bacci L, Pereira EJG, Crespo ALB. Selectivity of eight insecticides to predators of citrus caterpillars. Pesq agropec bras Brasília 2002; 37(2): 117-122.
- 148.Leite GLD, Picango M, Guedes RNC, Gusmáo MR. Selectivity of insecticides with and without mineral oil to *Brachygastra lecheguana* (Hymenoptera: Vespidae) a predator of *Tuta absoluta* (Lepidoptera: Gelechiidae). Ceiba 1998; 39(2): 191-194.
- 149. Tanaka Junior GM, Soleman RA, Noll FB. Morphological and physiological variation between queens and workers of *Protonectarina sylveirae* (de Saussure) (Hymenoptera, Vespidae Epiponini). Rev Bras Entomol 2010; 54(1): 104-109.
- 150.Moreno SC, Picanço MC, Silvério FO, Alvarenga ES, de Carvalho GA. Toxicity of new pyrethroids to the social insects *Protonectarina sylveirae Solenopsis saevissima* and *Tetragonisca angustula*. Sociobiology 2009; 54(3): 893-906.
- 151.Bacci L, Picando MC, Sousa FF, Silva EM, Campos MR, Tome HVT. (2008) Inimigos naturais da traca do tomateiro. Hortic Bras 26: 2808-2812.
- 152.Prezoto F, Braga N. Predation of *Zaprinus indianus* (Diptera: Drosophilidae) by the Social Wasp *Synoeca cyanea* (Hymenoptera: Vespidae). Florida Entomologist 2013; 96(2): 670-672.
- 153.Keller L. Queen life-span and colony characteristics in ants and termites. Insectes Sociaux 1998; 45 (3): 235-246.
- 154. Chapman RF. The Insects Structure and Function. Edn 4, Cambridge University Press, 1998, 600.
- 155.Detrain C, Deneubourg JL, Pasteels JM. Information processing in social insects. Birkhäuser Basel, 1999, 415.
- 156.Bownes A. The structure of ant communities and their impact on soil-pupating pests in citrus orchards in the Grahamstown area of the Eastern Cape M.Sc. Thesis Rhodes University, South Africa, 2002, 176.
- 157. Symondson WOC, Sunderland KD, Greenstone MH. Can generalist predators be effective biocontrol agents. Annu Rev Entomol 2002; 47: 561-594.
- 158.Radeghieri P. *Cameraria ohridella* (Lepidoptera: Gracillariidae) predation by *Crematogaster scutellaris*

- (Hymenoptera Formicidae) in Northern Italy (Preliminary note). Bull Insectol 2004; 57(1): 63-64.
- 159.Masuko K. Studies on the predatory biology of oriental dacetine ants (Hymenoptera: Formicidae) II Novel prey specialization in *Pyramica benten*. J Nat Hist 2009; 43(13-14): 825-841.
- 160. Fernandes FL, Picanco MC, Fernandes MES, Xavier VM, Martins JC, Salva VF. Controle biologic natural de pragas e interações ecológicas com predadores e parasitóides em feijoeiro. Biosci J 2010; 26: 6-14.
- 161.Sanders D, van Veen FJF. Ecosystem engineering and predation the multi-trophic impact of two ant species. J Anim Ecol 2011; 80: 569-576.
- 162.Rabeling C, Verhaagh M, Garcia MVB. Observations on the specialized predatory behavior of the pitchforkmandibled Ponerine ant *Thaumatomyrmex paludis* (Hymenoptera: Formicidae). Breviora 2012; 533: 1-8.
- 163.Trager JC. A revision of the fire ants *Solenopsis geminata* group (Hymenoptera: Formicidae: Myrmicinae). J New York Entomol Soc 1991; 99: 141-198.
- 164. Carroll CR, Risch SJ. Tropical annual cropping systems Ant ecology Environ Manage 1983; 7(1): 51-57.
- 165. Yusa Y. Predation on eggs of the apple snail *Pomacea canaliculata* (Gastropoda: Ampullaridae) by the fire ant *Solenopsis geminate*. J Molluscian Studies London 2001; 67: 275-279.
- 166. Way MJ, Javier G, Heong KL. The role of ants especially the fire ant *Solenopsis geminata* (Hymenoptera: Formicidae) in the biological control of tropical upland rice pests. Bull Entomol Res 2002; 92(5): 431-437.
- 167. Probst K, Pulschen L, Sauerborn J, Zebitz CPW. Influencia de varios regimenes de uso de plaguicidas sobre la entomofauna de tomate en las tierras altas de. Ecuador Manejo Integrado de Plagas CATIE Costa Rica 1999; 54: 53-62 (In Spanish).
- 168.Oliveira RF, Almeida LC, Munhae CB. Bueno OC. Morini MSC. Ant diversity (Hymenoptera: Formicidae) and predation by ants on the different stages of the sugarcane borer life cycle *Diatraea saccharalis* (Lepidoptera: Crambidae). Eur J Entomol 2012; 109(3): 381-387.
- 169.Moreno SC, Carvalho GA, Picanço MC, Morais EG, Pereira RM. Bioactivity of compounds from *Acmella oleracea* against *Tuta absoluta* (Meyrick) (Lepidoptera: Gelechiidae) and selectivity to two non-target species. Pest Manag Sci 2012; 68(3): 386-393.
- 170.Sutherland JAA. review of the biology and control of sweetpotato weevil *Cylas formicarius* (Fab.). Trop Pest Manage 1986; 32: 304-315.
- 171. Amalin DM, Vasquez EA. A handbook on Philippine sweet potato pests and their natural enemies. International Potato Center (CIP) Los Baños Philippines, 1993, 82.
- 172.Das NM, Abraham CC, Mathew KP. New record of *Pheidole* sp Hymenoptera Formicidae as a predator of the rice leaf folder *Cnaphalocrocis medinalis* Guen. Current Science 1974; 43(23): 767-768.
- 173.Silva-Torres CS, Pontes IV, Torres JB, Barros R. New records of natural enemies of *Plutella xylostella* (L.) (Lepidoptera: Plutellidae) in Pernambuco Brazil. Neotrop Entomol 2010; 39(5): 835-838.
- 174. Pereira JA, Bento A, Cabanas JE, Torres LM, Herz A,

- Hassan SA. Ants as predators of the egg parasitoid *Trichogramma* cacoeciae (Hymenoptera: Trichogrammatidae) applied for biological control of the olive moth *Praysoleae* (Lepidoptera: Plutellidae) in Portugal. Bioc Sci Technol 2004; 14: 653-664.
- 175. Schatz B, Kjellberg F, Nyawa S, Hossaert-Mckey M. Fig wasps a staple food for ants on *Ficus*. Biotropica 2008; 40: 190-195.
- 176.Gillott C. Entomology, Edn 3, Springer Dordrecht, 2005, 175-179.
- 177.Marshall SA. Insects their natural history and diversity with a photographic guide to insects of eastern North America. Edn 4, Firefly Books, Buffalo, NY, Richmond Hill Ontario, 2006, 63-64.
- 178. Karsemeijer MMD. Observations on the enemies of the oyster shell scale *Lepidosaphes ulmi* on apple in. The Netherlands J Plant Pathol 1973; 79: 122-124.
- 179.Romeu-Dalmau C, Piñol J, Agustí N. Detecting aphid predation by earwigs in organic citrus orchards using molecular markers. Bull Entomol Res 2012; 14:1-7.
- 180.Phillips ML. The ecology of the common earwig *Forficula auricularia* in apple orchards Ph.D. Thesis University of Bristol, 1981, 241.
- 181.Lenfant C, Lyoussoufi A, Chen X, D'Arcier FF, Sauphanor B. Potentialités prédatrices de Forficula auriculariasur le psylle du poirier Cacopsylla pyri. Entomol Exper Appl 1994; 73: 51-60.
- 182.Redoan ACM, Carvalho GA, Cruz I, Figueiredo M, de LC, Silva RB. da Physiological selectivity of insecticides to adult of *Doru luteipes* (Scudder, 1876) (Dermaptera: Forficulidae). Rev Ciência Agron 2013; 44(4): 842-850.
- 183.Kuthe K. The comman earwig (*Forficula auricularia*) an occasional pest of sugarbeet. Ges Pfl 1996; 48: 55-57.
- 184. Cooper R. Earwigs an IPM approach to controlling this common household pest. Pest Cont Tech 1997; 25: 54-48.
- 185.Kharboutli MS, Mack TP. Effect of temperature humidity and prey density on feeding rate of the striped earwig (Dermaptera: Labiduridae). Env Ent 1993; 22: 1134-1139.
- 186.Dugard R. Orange County vector control district Pest Control Bull 13 13001 Garden Grove Boulevard CA 92843, 2006, www.ocvcd.org.
- 187.Guimaraes J.H, Tocci EC, Gomes JPC. Dermaptera (Insecta) associated with commercial poultry ranches in Sao Paulo State and their importance as biological control agents of poultry pests. Rev Brasil Entomol 1992; 36 (3): 527-534.
- 188.Domiciano NL, Herzog DC. Population dynamics of pests and some of their predators in soybean field under influence of herbicidal applications of Toxaphene. Resquisia Agropecuaria Brasileria 1990; 25(2): 253-273.
- 189.Kharboutli MS, Mack TP. Relative and seasonal abundance of predaceous arthropods in Alabama peanut fields as indexed by Pitfall traps. J Econ Entomol 1991; 84 (3): 1015-1023.
- 190. Ahmed SA, Darwish YA. Influence of weed control on sucking insect pests and natural enemies in a corn field in Assiut. Assiut J Agric Sci 1991; 22 (2): 3-13.
- 191.Darwish YA, Farghal AI. (1990) Evaluation of certain pesticides activity against the cotton whitefly *Bemisia* tabaci and associated natural enemies on cotton plants

- under field conditions in Assiut. Assiut J Agric Sci 21 (5): 331-339.
- 192. Abbas MST, Abou-Zeid NA, Megahed MM. On the natural enemies of the potato tuber moth *Phthorimaea operculella* in Egypt. Egyptian J Agric Res 1993; 71(4): 943-950.
- 193.El-Defrawi GM, Emam AK, Marzouk IA, Rizkala L. Population dynamics and seasonal distribution of *Aphis craccivora* K and associated natural enemies in relation to virus disease incidence in faba bean field. Egyptian J Agric Res 2000; 78 (2): 627-641.
- 194.Bassal TM, El-Naggar ME, Fahmy NM, Dorrah MAA, Sallam MH, Salama MS. Carnivory rate of digestion and prey consumption by *Labidura riparia* (Dermaptera: Labiduridae). Efflatounia 2001; 1: 13-19.
- 195.Engel MS, Haas F. Family-group names for earwigs (Dermaptera). American Museum Novitates, 2007, 3567, 20.
- 196.Powell JA. Dermaptera In (Resh, V.H. and Cardé, R.T., eds) "Encyclopedia of Insects" Academic Press, 2009, 1132.
- 197. Pasini A, Postali Parra JR, Nava DE, Butnariu AR. Thermal requirements of *Doru lineare* Eschs and *Doru luteipes* Scudder in the laboratory Ciência Rural Santa Maria 2010; 40(7): 1562-1568.
- 198.Butnariu AR, Pasini A, Reis FS, Bessa E. Maternal Care by the Earwig *Doru lineare* Eschs (Dermaptera: Forficulidae). J Insect Behavior 2013; 26(5): 667-678.
- 199.Zotti M.J, Grutzmacher AD, Grutzmacher DD, Castilhos RV, Martins JFS. Seletividade de inseticidas usados cultura do milho para ovos e ninfas do predador *Doru lineare* (Eschcholtz, 1822) (Dermaptera: Forficulidae). Arquivos do Instituto Biológico São Paulo 2010; 77(1): 111-118.
- 200. Vandenberg NJ. Coccinellidae Latreille 1807 In (Arnett, RH and Thomas, M.C. eds.) "American Beetles" Vol 2, "Polyphaga: Scarabaeoidea through Curculionoidea" pp 371-389 Boca Raton CRC Press, 2002, 861.
- 201.Dixon AFG. Insect predator–prey dynamics Ladybirds beetles & biological control London Cambridge University Press, 2000, 257.
- 202.Hodek I, Honek A, van Emden HF. Ecology and behaviour of the ladybird beetles (Coccinellidae). John Wiley & Sons, 2012, 604.
- 203. Obrycki JJ, Kring TJ. Predaceous Coccinellidae in biological control. Annu RevEntomol 1998; 43: 295-321.
- 204. Cardoso JT, Lázzar SMN. Comparative biology of *Cycloneda sanguinea* (Linnaeus, 1763) and *Hippodamia convergens* Guérin-Méneville 1842 (Coleoptera, Coccinellidae) focusing on the control of *Cinara* spp (Hemiptera, Aphididae). Rev Bras entomol 2003; 47 (3): 443-446.
- 205. Heit GE, Sardoy P, Cohen GR, Mareggiani G. Locomotor activity of *Cycloneda sanguinea* (Coleoptera: Coccinellidae) exposed to volatile semiochemicals and to direct contact with the odour source. Rev Soc Entomol Argent 2007; 66(3-4): 197-203.
- 206.Nakasu YET, Dias SC, Pires CSS, Andow DA, Paula DP, Togni PHB, Macedo TR, Sujii ER, Sá MFG de. Fontes EMG. Bitrophic toxicity of Cry1Ac to *Cycloneda sanguinea* a predator in Brazilian cotton. Entomol Exper

- Appl 2013; 148: 105-115.
- 207.Bergmann EC, Imenes SDL, Campos TB, Hojo H, Takematsu AP, Macellaro MIFS. Levantamento da entomofauna em cultura de tomate (*Lycopersicon esculentum*) O Biologico Sao Paulo 1984; 50: 229-236. (In Portugese with English summary).
- 208.Bergmann EC, Imenes SDL, Campos TB, Hojo H, Takematsu AP. Contribuic, ao conhecimento da entomofauna em cultura de tomate (*Lycopersicum esculentum*) atraves de arma-dilhas de agua Anais da. Sociedade Entomologica do Brasil Jaboticabal 1988; 17:19-40. (In Portugese with English summary).
- 209. Miranda MMM, Picanco MC, Zanuncio JC, Bacci L, Silva EM da. Impact of integrated pest management on the population of leafminers fruit borers and natural enemies in tomato. Cienc Rural 2005; 35: 204-208.
- 210.Samways MJ, Wilson SJ. Aspects of the feeding behaviour of *Chilocorus nigritus* (F.) (Col., Coccinellidae) relative to its effectiveness as a biocontrol agent. J Appl Entomol 1988; 106: 177-182.
- 211.Omkar P. A Ecology and Biocontrol Potential of a Scale-Predator *Chilocorus nigritus*. Biocontrol Sci Technol 2003; 13(4): 379-390.
- 212.Boothe RA, Ponsonby DJ. Searching behaviour in *Chilocorus nigritus* (F.) (Coleoptera: Coccinellidae). Commun Agric Appl Biol Sci 2006; 71(2 Pt A): 253-261.
- 213. Vasicek AL. Natural enemies of *Scrobipalpula absoluta* Meyr (Lep.: Gelechidae) Revista de la Facultad de. Agronomia Universidad Nacional de la Plata 1983; 59(1/2): 199-200.
- 214.Santos-Cividanes TM, Anjos ACR dos, Cividanes FJ, Dias PC. Effects of food deprivation on the development of *Coleomegilla maculata* (De Geer) (Coleoptera: Coccinellidae). Neotrop Entomol 2011; 40(1): 112-116.
- 215.Ragsdale DW, Landis DA, Brodeur J, Heimpel GE, Desneux N. Ecology and management of the soybean aphid in North America. Annu Rev Entomol 2011; 56: 375-99.
- 216. Silva RB da, Cruz I, Figueiredo MA de LC. Tavares W de S. Development of *Coleomegilla maculate* de Geer (Coleoptera: Coccinellidae) with prey and artificial diet. Revista Brasileira de Milho e Sorgo, 2010; 9(1): 13-26.
- 217.Leite GLD, Araujo CBO, Amorim CAD, Martins ER. Weather factors affecting arthropods abundance in medicinal plants in the Minas Gerais State Brazil. Rev Bras Pl Med Botucatu 2006; 8(3): 43-51.
- 218.Kromp B. Carabid beetles in sustainable agriculture a review on pest control efficacy cultivation aspects and enhancement. Agri Ecosys Environ 1999; 74 (1-3): 187-228.
- 219. Fournier E Loreau M. Effects of newly planted hedges on ground-beetle (Coleoptera: Carabidae) in an agricultural landscape. Ecography 1999; 22: 87-97.
- 220. Tooley J, Brust GE. Weed predation by carabid beetles In Edn Holland, J.M. "The agroecology of carabid beetles" Intercept Andover United Kingdom, 2002, 215-229.
- 221. Chocorosqui VR, Pasini A. Predação de pupas de *Alabama argillacea* (Hübner) (Lepidoptera: Noctuidae) por larvas e adultos de *Calosoma granulatum* Perty (Coleoptera: Carabidae) em laboratório. An Soc Entomol Bras 2000; 29(1): 65-70.

- 222. Wyckhuys KAG, O'Neil RJ. Population dynamics of *Spodoptera frugiperda* Smith (Lepidoptera: Noctuidae) and associated arthropod natural enemies in Honduran subsistence maize. Crop Prot 2006; 25: 1180-1190.
- 223. Allen RT. *Calosoma* (*Castrida*) alternans granulatum Perty a predator of cotton leaf worms in Bolivia (Coleoptera: Carabidae: Carabini). Coleopterist Bull 1977; 31: 73-76.
- 224.Pegoraro RA, Foerster LA. Abundance and distribution of larvae and adults of *Calosoma granulatum* Perty 1830 (Coleoptera: Carabidae) in cultivars of soyabean sown at different dates. Anais da Sociedade Entomológica do Brasil, 1988; 17(2): 237-248.
- 225.Engel MS, Grimaldi DA. (2007) The neuropterid fauna of Dominican and Mexican amber (Neuropterida, Megaloptera, Neuroptera). American Museum Novitates 3587: 1-58.
- 226.New TR. Prospects for extending the use of Australian lacewings in biological control. Acta Zool Acad Sci Hungaricae 48(Supplement 2): 2002, 209-216.
- 227. Auad AM, Carvalho CF, Souza B, Barbosa LR. Biological aspects of *Chrysoperla externa* (Hagen, 1861) (Neuroptera: Chrysopidae) fed on eggs and larvae of *Spodoptera frugiperda* (JE Smith, 1797) (Lepidoptera: Noctuidae). Rev Bras Milho Sorgo 2003; 2: 106-111
- 228. Hoballah ME, Degen T, Bergvinson D, Savidan A, Tamò, C, Turlings TCJ. Occurrence and direct control potential of parasitoids and predators of the fall armyworm (Lepidoptera: Noctuidae) on maize in the subtropical lowlands of Mexico. Agr Forest Entomol 2004; 6: 83-88.
- 229. Hagerty AM, Kilpatrick AL, Turnipssed SG, Sullivan MJ. Bridges WC. Predaceous arthropods and lepidopteran pests on conventional Bollgard and Bollgard II cotton under untreated and disrupted conditions. Environ Entomol 2005; 34: 105-114.
- 230.Ribeiro LJ, Freitas S. de Influence of food on *Chrysoperla externa* (Hagen) (Neuroptera: Chrysopidae) reproductive potential. Rev Agric (Piracicaba) 2000; 75(2): 187-196.
- 231.Silva CG, Souza B, Auad AM, Bonani JP, Torres LC, Carvalho CF, Ecole CC. Development of immature tages of *Chrysoperla externa* fed on nymphs of *Bemisia tabaci* biotype B reared on three hosts Pesqui Agropecu Bras 2004; 39: 1065-1070.
- 232. Souza B, Costa RIF, Tanque RL, Oliveira OS, Santos FA. Predation among *Chrysoperla externa* (Hagen, 1861) and *Ceraeochrysa cubana* (Hagen, 1861) (Neuroptera: Chrysopidae) larvae under laboratory conditions Ciênc Agrotec, 2008; 32: 712-716.
- 233. Tavares WS, Cruz I, Silva RB, Serrão JE, Zanuncio JC. Prey consumption and development of *Chrysoperla externa* (Neuroptera: Chrysopidae) on *Spodoptera frugiperda* (Lepidoptera: Noctuidae) eggs and larvae and *Anagasta kuehniella* (Lepidoptera: Pyralidae) eggs. Maydica 2011; 56(3): 283-289.
- 234. Carneiro JR, Medeiros MA. Potencial de consumo de *Chrysoperla externa* (Neuroptera: Chrysopidae) utilizando ovos de *Tuta absoluta* (Lepidoptera: Gelechiidae) In: "Congresso Brasileiro de Entomologia" (Abstracts). vol 16, Salvador Brazil, 1997, 117-118.
- 235.Hassan SA, Klingauf F, Shahin F. Role of *Chrysopa* carnea as an aphid predator on sugar beet and the effect of

- pesticides. Zeitsch Ange Entomol 1985; 100(1-5): 163-174.
- 236. Abid MK, Al-Rubeae JK, Hussien AK. Biological studies on the predator *Chrysopa mutata* McLachlan (Chrysopidae Neuroptera) in Iraq. J Agric Water Resour Res 1985; 4(1): 153-160.
- 237.Aly MZY, Osman KSM, Mohamed IEE, Wahballah NM. The relationship between some insect predators and aphid insects in Qena Governorate Egypt. Acad J biolog Sci 2011; 4 (1): 131- 144.
- 238. Flores JL, Guevara Acevedo LP, Aguirre Uribe LA, Chavez EC, Badii Zabeh MH, Ochoa Fuentes YM. Functional Response of *Chrysoperla carnea* (Neuroptera: Chrysopidae) on *Trialeurodes vaporariorum* (Hemiptera: Aleyrodidae) Eggs. Southwestern Entomologist 2013; 38(2): 345-352.
- 239. Mound LA. Thysanoptera diversity and interactions. Annu Rev Entomol 2005; 50: 247- 269.
- 240. Tipping C. Thrips (Thysanoptera) In (Capinera, J.L. ed.) "Encyclopedia of Entomology" Springer, New York, 2008, 3769-3771.
- 241.Mound LA, Morris DC. Thysanoptera Phylogeny-the morphological background. Acta PhytopatholEntomol Hungarica 2004; 39: 101-113.
- 242.Mirab-balou M, Minaei K, Chen X.-X. An illustrated key to the genera of Thripinae (Thysanoptera, Thripidae) from Iran. Zookeys, 2013; 317: 27-52.
- 243.Gilstrap FE, Oatman ER. (1976) The bionomics of *Scolothrips sexmaculatus* (Pergande) (Thysanoptera: Thripidae) an insect predator of spider mites. Hilgardia 44(2): 27-59.
- 244. Naher N, Islam W, Haque MM. Predation of three predators on two-spotted spider mite *Tetranychus Urticaekoch* (Acari: Tetranychidae). J Life Earth Science 2005; 1(1): 1-4.
- 245.Mound LA. Order Thysanoptera Haliday 1836. In: (Edn Zhang, Z. Q.) "Animal biodiversity: An outline of higher-level classification and survey of taxonomic richness" Zootaxa 3148, 2011, 1-237.
- 246.Buckman RS, Mound LA, Whiting MF. Phylogeny of thrips (Insecta: Thysanoptera) based on five molecular loci. System Entomol 2013; 38(1): 123-133.
- 247.Moritz G, Morris D, Mound L. Thrips ID pest thrips of the world ACIAR and CSIRO Publishing Collingwood VIC. Australia, 2001.
- 248.Morse JG, Hoddle MS. Invasion biology of thrips. Annu Rev Entomol 2005; 51: 67-89.
- 249.Mound LA, Morison GD, Pitkin BR, Palmer JM. Thysanoptera. Hdbks Ident Br Ins 1976; I: 1-79.
- 250.Riudavets J. Predators of *Frankliniella occidentalis* (Perg.) and *Thrips tabaci* Lind a review. Wageningen Agricultural University Papers, The Netherlands 1995; 95-1: 43-87.
- 251.Bailey P, Caon G. Predation on twospotted mite *Tetranychus urticae* Koch (Acarina: Tetranychidae) by *Haplothrips victoriensis* Bagnall (Thysanoptera: Phlaeothripidae) and *Stethorus nigripes* Kapur (Coleoptera: Coccinellidae) on seed lucerne crops in South Australia. Aust J Zool 1986; 34: 515-525.
- 252.Kakimoto K, Inoue H, Hinomoto N, Noda T, Hirano K, Kashio T, Kusigemati K, Okajima S. Potential of

- Haplothrips brevitubus (Karny)(Thysanoptera: Phlaeothripidae) as a predator of mulberry thrips Pseudodendrothrips mori (Niwa) (Thysanoptera: Thripidae). Biological Control 2006; 37: 314-319.
- 253.Mound LA. A new genus of Aeolothripidae (Thysanoptera) from New Zealand and New Caledonia New Zealand. J Zool 1977; 4:149-152.
- 254. Araraki N, Okajima S. Notes on the biology and morphology of a predatory thrips *Franklinothrips vespiformis* (Crawford) (Thysanoptera: Aeolothripidae) first record from Japan. Entomol Sci 1998; 1: 359-363.
- 255.Loomans AJM, Heijboer A. *Franklinothrips vespiformis* (Crawford) a polyphagous predator preying on thrips eggs Proceedings of the section Experimental and Applied Entomology of the Netherlands. Entomological Society (N.E.V.) 1999; 10: 143-150.
- 256.Hoddle MS. Predation behaviors of Franklinothrips orizabensis (Thysanoptera: Aeolothripidae) towards Scirtothrips perseae and *Heliothrips haemorrhoidalis* (Thysanoptera: Thripidae). Biological Control 2003; 27: 323-328.
- 257.Larentzaki E. (2004) Evaluation of 'Franklinothrips vespiformis' (Crawford) (Thysanoptera: Aeolothripidae) as a natural enemy of glasshouse pests Ph.D. Thesis Imperial College London University of London. 244.
- 258.Mound LA, Reynaud P. *Franklinothrips* a pantropical Thysanoptera genus of ant-mimicking obligate predators (Aeolothripidae). Zootaxa 2005; 864: 1-16.
- 259.Pizzol J, Nammour D, Hervouet P, Poncet C, Desneux N, Maignet P. Population dynamics of thrips and development of an integrated pest management program using the predator *Franklinothrips vespiformis*. Acta Horticu 2012; 927: 219-226.
- 260.Moraes GJ de, McMurtry JA, Denmark HA, Campos CB A revised catalog of the mite family Phytoseiidae. Zootaxa 2004; 434: 1-494.
- 261.Maanen van R, Vila E, Sabelis MW, Janssen A. Biological control of broad mites (*Polyphagotarsonemus latus*) with the generalist predator *Amblyseius swirskii*. Exper Appl Acarol 2010; 52: 29-34.
- 262.Onzo A, Houedokoho AF, Hanna R. Potential of the predatory mite *Amblyseius swirskii* to suppress the broad mite *Polyphagotarsonemus latus* on the gboma eggplant *Solanum macrocarpon* J Insect Sci 2012; 12:7 Available online insectscience.org/12.7
- 263. Wittmann EJ, Leather SR. Compatibility of *Orius laevigatus* Fieber (Hemiptera: Anthocoridae) with *Neoseiulus Amblyseius cucumeris* Oudemans (Acari: Phytoseiidae) and *Iphiseius Amblyseius degenerans* Berlese (Acari: Phytoseiidae) in the biocontrol of *Frankliniella occidentalis* Pergande (Thysanoptera: Thripidae). Exper Appl Acarol 1997; 21(8): 523-538.
- 264. Houten YM, van Stratum P. van Biological control of western flower thrips in greenhouse sweet pepper using non- diapausing predatory mites. Proceedings of the section Experimental and Applied Entomology of the Netherlands Entomological Society (N.E.V.), 1993; 4: 229-234.
- 265. Houten YM, van Stratum P, van Bruin J, Veerman A. Selection for non-diapause in *Amblyseius cucumeris* and *Amblyseius barkeri* and exploration of the

- effectiveness of selected strains for thrips control. Entomol Experi Appl 1995; 77: 289-295.
- 266.Kim JH, Broadbent AB, Lee SG. Quality control of the mass-reared predatory mite, *Amblyseius cucumeris* (Acarina: Phytoseiidae). J Asia-Pacific Entomol 2001; 4(2): 175-179.
- 267. Zhang HY, Ma M, Dong T, Liu X.Y, Zhang K, Wang FL. Biological control efficiency of *Amblyseius cucumeris* (Oudemans) on *Panonychus ulmi* (Koch)] Ying Yong Sheng Tai Xue Bao 2010; 21(1): 191-196. (In Chinese with English summary)
- 268. Conley MR. Predation versus resource limitation in survival of adult burrowing wolf spiders (Araneae: Lycosidae). Oecologia 1985; 67: 71-75.
- 269.Oedekoven MA. Stage-based mortality of grassland grasshoppers (Acrididae) from wandering spider (Lycosidae) predation. Acta Oecologica 1998; 19(6): 507-515
- 270.Monzo C, Sabater-Munoz B, Urbaneja A, Castanera P. Tracking medfly predation by the wolf spider *Pardosa cribata* Simon in citrus orchards using PCR-based gutcontent analysis. Bull Entomol Res 2010; 100: 145-152.
- 271.Albajes R, Alomar O. Current and potential use of polyphagous predators In. Edn Albajes, R.; Gullino, M.L.; Van Lenteren, J.C.; Elad, Y. "Integrated Pest Disease Management in Greenhouse Crops" Kluwer Academic Publishers Dordrecht The Netherlands, 1999, 265-275.
- 272. Hamdi F, Bonato O. Study of the temperature influence on the functional response of the biological control agent *Macrolophus pygmaeus* Les Cochenilles ravageur principal ou secondaire 9ème Conférence Internationale sur les Ravageurs en Agriculture SupAgro Montpellier France 25-27 octobre, 2011, 267-272.
- 273.Gabarra R, Protecció V, Arnó J. Results of tomato leaf miner biological control experiments in greenhouse and open air crops in Catalonia (Spain). Phytoma España 2010; 217: 66-68.
- 274.Belda JE, Calvo FJ, Giménez A. Biological control strategy for *Tuta absoluta* in tomato by release of *Nesidiocoris tenuis* in pre-transplant. Phytoma España 2010; 217: 48-52.
- 275.Calvo FJ, Gimenez A. Belda JE. Biological control strategy for *Tuta absoluta* in tomato by release of *Nesidiocoris tenuis* in pre-transplant Encuentro Internacional sobre *Tuta absoluta* La Polilla del Tomate un Grave Problema en Expansión1 Valencia (España) 23-24 Mar. 2010.
- 276.Nannini M, Atzori F, Murgia G, Pisci R. Sanna F. Use of predatory mirids for control of the tomato borer *Tuta absoluta* (Meyrick) in Sardinian greenhouse tomatoes. EPPO Bull 2012; 42(2): 255-259.
- 277. Tavella L, Ferracini C, Ingegno BL. Adaptation of native natural enemies the parasitoids of the genus *Necremnus* and the predator *Dicyphus errans*. Atti Accademia Nazionale Italiana di Entomologia 2012; LX: 95-102.
- 278.Ferracini C, Ingegno BL, Mosti M, Navone P, Tavella L. Alma A. Promising native candidates for biological control of *Tuta absoluta* in Italy. IOBC-WPRS Bull 2012; 80: 51-55.
- 279. Sanchez JA, López JCE, Spina M, Mengual La, Monserrat M, Lacasa A. Benefits and damage of *Nesidiocoris tenuis*

- in tomato crops In. Edn Mason, P.G.; Gillespie, D.R.; Vincent, C. Proceedings of ISBCA 3 (International Symposium on Biological Control of Arthropods) Christchurch New Zealand, 2008.
- 280.Call for integrated pest management as Mediterranean tomato pests spread to UK Horticulture Week (October 9, 2009).
 - http://www.hortweek.com/news/search/943628/Call-integrated-pest-management-Mediterranean-tomato-pests-spread-UK/. 5 Nov, 2009.
- 281. Boualem M, Allaoui H, Hamadi R. Biology of *Tuta absoluta* (Meyrick) and the natural predator *Nesidiocoris tenuis* (Reuter) in Mostaganem area (Algeria) Les Cochenilles ravageur principal ou secondaire 9ème Conférence Internationale sur les Ravageurs en Agriculture SupAgro Montpellier France 25-27 Octobre, 2011, 299-307.
- 282. Abbes K, Chermiti B. Failure of the biological control of *Tuta absoluta* using the predator *Nesidiocoris tenuis* in a protected tomato crop analysis of factors. IOBC-WPRS Bull 2012; 80: 231-236.
- 283. Salas J. *Orius insidiosus* (Hemiptera: Anthocoridae) su presencia en la region centro occidental de Venezuela. Agron Trop 1995; 45: 637-645.
- 284. Vargas CH. Observations on the bionomics and natural enemies of the tomato moth *Gnorimoschema absoluta* (Meyrick) (Lep. Gelechiidae). Depto Agricultura Universidad del Norte-Arica 1970; 1: 75-110.
- 285.Croft BA. Arthropod biological control agents and pesticides John Wiley & Sons New York, 1990, 723.
- 286.Desneux N, Decourtye A, Delpuech JM. The sub-lethal effects of pesticides on beneficial arthropods. Annu Rev Entomol 2007; 52: 81-106.
- 287.Biondi A, Desneux N, Siscaro G, Garzia GT, Amiens-Desneux E, Zappalà L. Side effects of bioinsecticides used to control *Tuta absoluta*. IOBC-WPRS Bull 2012; 80: 211-216.
- 288.Biondi A, Desneux N, Siscaro G, Zappalà L. Using organic-certified rather than synthetic pesticides may not be safer for biological control agents selectivity and side effects of 14 pesticides on the predator *Orius laevigatus*. Chemosphere 2012; 87: 803-812.
- 289. Picanco M, Ribeiro LJ, Leite GLD, Gusmao MR. (1998) Selectivity of insecticides to *Polybia ignobilis* (Haliday) (Hymenoptera: Vespidae) predator of *Ascia monuste orseis* (Godart) (Lepidoptera: Pieridae). An Soc Entomol Bras 27(1): 85-90.
- 290. Gianessi L, Williams A. Insecticides have key role in controlling *Tuta absoluta* in Mediterranean tomatoes International Pesticide Benefits CropLife Foundation Washington DC 2012 Case Study No 67 1p.
- 291. Pasini A, Foerster LA. Effect of insecticides on *Calosoma* granulatum P (Coleoptera: Carabidae). Anais da sociedade entomologica do brasil 1994; 23: 455-460.
- 292. Cordeiro EM, Corrêa AS, Venzon M, Guedes RN. Insecticide survival and behavioral avoidance in the lacewings *Chrysoperla externa* and *Ceraeochrysa cubana*. Chemosphere 2010; 81(10): 1352-1357.
- 293. Silva RA, Carvalho GA, Carvalho CF, Silva DB. Effects of pesticides on eggs of *Chrysoperla externa* (Neuroptera: Chrysopidae) and consequences on subsequent

- development. Rev ColombEntomol 2012; 38(1): 58-63.
- 294.Zotti MJ, Grutzmacher A.D, Lopes IH, Smagghe G. Comparative effects of insecticides with different mechanisms of action on *Chrysoperla externa* (Neuroptera: Chrysopidae): Lethal sublethal and doseresponse effects. Insect Sci 2013; 20(6): 743-752.
- 295.Zappala L, Siscaro G, Biondi A, Mollá O, González-Cabrera J, Urbaneja A. Efficacy of sulphur on *Tuta absoluta* and its side effects on the predator *Nesidiocoris tenuis*. J Appl Entomol 2012; 136(6): 401-409.
- 296.Martinou AF, Seraphides N, Stavrinides MC. Lethal and behavioral effects of pesticides on the insect predator *Macrolophus pygmaeus*. Chemosphere 2014; 96: 167-173.
- 297.Iannacone OJ, Lamas MG. Toxicological effects of extracts of Peruvian peppertree (*Schinus molle*) and lantana (*Lantana camara*) on *Chrysoperla externa* (Neuroptera: Chrysopidae) *Trichogramma pintoi* (Hymenoptera: Trichogrammatidae) and *Copidosoma koehleri* (Hymenoptera: Encyrtidae) in Perú. Agricultura Técnica 2003; 63(4): 347-360.
- 298.Desneux N, Pizzol J, Thomas C, Pautrat E, Bearez P, Poncet C, Tabone E, Kabiri F, Frandon J. Potential for direct interference between natural enemies of *Tuta absoluta* on tomato. Acta Horticulturae 2011; 917: 31-37.
- 299. Trottin-Caudal Y, Baffert V, Leyre J-M, Hulas N. Experimental studies on *Tuta absoluta* (Meyrick) in protected tomato crops in France biological control and integrated crop protection. EPPO Bull 2012; 42(2): 234-240.
- 300.Calvo FJ, Lorente MJ, Stansly PA, Belda JE. Preplant release of *Nesidiocoris tenuis* and supplementary tactics for control of *Tuta absoluta* and *Bemisa tabaci* in greenhouse tomato. Entomol Exper Appl 2012; 143(2): 111-119
- 301. Eilenberg J, Hajek A, Lomer C. Suggestions for unifying the terminology in biological control. BioControl 2001; 46: 387-400.
- 302.Barbosa P. Conservation Biological Control Academic Press San Diego CA 2003.
- 303.Gurr GM, Scarratt SL, Wratten SD, Berndt L, Irvin N. Ecological engineering habitat manipulation and pest management In. Edn Gurr, G.M.; Wratten, S.D.; Altieri, M.A. "Ecological Engineering for Pest Management: Advances in Habitat Manipulation for Arthropods" Cornell University Press New York NY, 2004, 1-12.
- 304. Wilkinson TK, Landis DA. Habitat diversification in biological control the role of plant resources In. Edn Wackers, F.L.; van Rijn, P.C.J.; Bruin, J. "Plant Provided Food and Plant-Carnivore Mutualism" Cambridge University Press Cambridge UK, 2005, 305-325.
- 305. Wade MR, Gurr GM, Wratten SD. Ecological restoration of farmland progress and prospects. Philos Trans Royal Soc B Biol Sci 2008; 363: 831-847.
- 306.Landis DA, Wratten SD, Gurr GM. Habitat management to conserve natural enemies of arthropod pests in agriculture. Annu RevEntomol 2000; 45: 175-201.
- 307. Wackers FL, van Rijn PCJ, Bruin J. (Eds.) Plant-provided food for carnivorous insects a protective mutualism and its applications Cambridge University Press Cambridge UK, 2005.
- 308. Jonsson M, Wratten SD, Landis DA, Gurr GM. Recent

- advances in conservation biological control of arthropods by arthropods. Biological Control 2008; 45: 172-175.
- 309.Balzan MV, Moonen AC. Management strategies for the control of *Tuta absoluta* (Lepidoptera: Gelechiidae) damage in open-field cultivations of processing tomato in Tuscany (Italy). OEPP/EPPO Bull 2012; 42(2): 217-225.
- 310.Berndt LA, Wratten SD. Effects of alyssum flowers on the longevity fecundity and sex ratio of the leafroller parasitoid *Dolichogenidea tasmanica*. Biological Control 2005; 32: 65-69.
- 311.Eubanks MD, Styrsky JD. Effects of plant feeding on the performance of omnivorous "predators" In. Edn Wäckers, F.L.; van Rijn, P.C.J.; Bruin, J. "Plant-provided food and herbivore-carnivore Interactions" Cambridge University Press, Cambridge, 2005, 148-177.
- 312.Lundgren JG. Relationships of Natural Enemies and Nonprey Foods Springer International Dordrecht The Netherlands 2009.
- 313.Lange WH, Bronson L. Insects pests of tomatoes Annu Rev Entomol 1981; 26: 345-372.
- 314.Perdikis D, Lykouressis D. Effects of various items host plants and temperatures on the development and survival of *Macrolophus pygmaeus* (Rambur) (Hemiptera: Miridae). Biological Control 2000; 17: 55-60.
- 315. Fantinou AA, Perdikis DCh, Labropoulos P, Maselou D. Preference and consumption of *Macrolophus pygmaeus* preying on mixed instar assemblages of *Myzus persicae*. Biol Control 2009; 51: 76-80.
- 316.Urbaneja A, Tapia G, Stansly P. Influence of host plant and prey availability on developmental time and survivorship of *Nesidiocoris tenuis* (Het.: Miridae). Biocontrol Sci Technol 2005; 15: 513-518.
- 317. Sanchez JA, Gillespie DR, McGregor RR. Plant preference in relation to life history traits in the zoophytophagous predator *Dicyphus hesperus*. Entomol Exper Appl 2004; 112: 7-19.
- 318. Alomar O, Goula M, Albajes R. Colonisation of tomato fields by predatory mirid bugs (Hemiptera: Heteroptera) in northern Spain. Agric Ecosystems Environ 2002; 89: 105-115.
- 319. Castañe C, Alomar O, Goula M, Gabarra R. Colonization of tomato greenhouses by the predatory mirid bugs *Macrolophus caliginosus* and *Dicyphus tamaninii*. Biological Control 2004; 30: 591-597.
- 320.Gabarra R, Alomar O, Castañe C, Goula M, Albajes R. Movement of greenhouse whitefly and its predators between in and outside of Mediterranean greenhouses. Agric Ecosys Environ 2004; 102: 341-348.
- 321.Lykouressis DH, Perdikis DCh, Chaljia FCh. The effects of natural enemies on aphid populations on processing tomato in central Greece. Entomologia Hellenica 2000; 13: 35-42.
- 322.Lykouressis D, Giatropoulos A, Perdikis D, Favas C. Assessing the suitability of non-cultivated plants and associated insect prey as food sources for the omnivorous predator *Macrolophus pygmaeus* (Hemiptera: Miridae). Biological Control 2008; 44, 142-148.
- 323.Arno J, Castane C, Riudavets J, Gabarra R. Risk of damage to tomato crops by the generalist zoophytophagous predator *Nesidiocoris tenuis* (Reuter) (Hemiptera: Miridae). Bull Entomol Res 2010; 100: 105-

115.

- 324.Roditakis NE, Perdikis D, Roditakis E, Lykouressis DP, Papadaki M. Beneficial Hemiptera Anthocoridae and Miridae in Crete 10th Panhellenic Entomological Congress Heraklion Crete 4–7 November 2003 (Book of Abstracts, p. 93) (In Greek with English summary).
- 325.Cano M, Vila E, Janssen D, Bretones G, Salvador E, Lara L, Tellez MM. Selection of refuges for *Nesidiocoris tenuis* (Het.: Miridae) and *Orius laevigatus* (Het.: Anthocoridae) virus reservoir risk assessment. IOBC/WPRS Bull 2009; 49: 281-286.